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### ORIGINAL ARTICLE

# Evaluation of the efficacy of 2% curcumin gel in the treatment of experimental periodontitis

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Submitted: 24-04-2014 Revised: 04-06-2014 Published: 06-08-2014

# ABSTRACT

Context: Local drug delivery (LDD) systems have been proposed for the treatment of periodontitis. Curcumin could be a suitable agent as LDD for the treatment of periodontitis. Aim: To formulate, evaluate the anti-inflammatory activity and to assess the duration of the action and the efficacy of 2% curcumin gel in the treatment of experimental periodontitis in Wistar albino rat model. Settings and Design: Twenty-one Wistar albino rats were randomly assigned to three groups. Periodontitis was induced using ligature model. Group 1: Control; group 2: Plain gel, and group 3: 2% curcumin gel. Materials and Methods: About 2% curcumin gel was prepared. The anti-inflammatory activity and duration of action was assessed. Silk ligature 5-0 was used to induce periodontitis. Gingival index (GI) and probing pocket depth (PPD) were measured. Treatment was done. The rats were sacrificed. Morphometric analysis was performed using stereomicroscope and ImageJ software. Statistical Analysis Used: Analysis of variance followed by Bonferroni's test, Wilcoxon's test for inter-group comparison, Mann-Whitney test for P value computation was used. The observations are mean ± standard deviation and standard error of the mean. P < 0.01 when compared to control was considered as statistically significant. Results: About 2% curcumin gel showed 42.98% inhibition of edema and peak activity was noted at 24 h. There was statistically significant change in the GI and PPD. Morphometric analysis did not show any significant difference between groups. No toxic effects were seen on oral administration of 2000 mg/kg of curcumin. Conclusions: About 2% curcumin gel was effective in the treatment of experimental periodontitis.



**Key words:** Anti-inflammatory activity, curcumin gel, experimental periodontitis, local drug delivery, morphometric analysis

#### INTRODUCTION

Periodontitis is one of the most common diseases affecting the teeth. It is seen in individuals who are more susceptible to the disease.<sup>[1]</sup>

The remnants of plaque and calculus can cause failure in periodontal treatment. <sup>[2]</sup> Chemotherapeutic agents can be used to enhance the results achieved by mechanical instrumentation or to improve outcomes at sites not responsive to conventional

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therapy.<sup>[3]</sup> Several research studies have established their efficacy in the treatment of periodontal diseases.<sup>[4]</sup>

### **MATERIALS AND METHODS**

Institutional Animal Ethical Committee, approval was obtained. Dosage of the gel was selected based on the results of acute oral toxicity study of *Curcuma longa* in Wistar albino rats carried out according to Organization for Economic Cooperation and Development Guidelines No. 420. The rats were housed in polypropylene cages under standard conditions with access to food and water ad-libitum.

The aim of this study was to formulate, evaluate the anti-inflammatory activity, to assess the duration of action

and the efficacy of 2% curcumin gel in the treatment of experimentally induced periodontitis in Wistar albino rat model.

# Preparation of 2% curcumin gel by simple dispersion method

Carbopol-940 was soaked in purified water containing 0.2% w/v sodium benzoate overnight. Using tissue homogenizer hydroxypropyl methylcellulose (HPMC) solution was mixed in propylene glycol. 2 mg of curcumin (Rajesh Chemicals, Mumbai) was transferred into HPMC solution and homogenized. This drug solution was transferred to carbopol solution and homogenized. Triethanolamine was added quantity sufficient (q.s.) to neutralize the pH. Then, distilled water was added to make q.s. to 100 ml. Control gel was prepared in the same manner. The gel was stored at ambient temperature. The formulation of 2% curcumin gel was prepared by Department of Pharmaceutics, NGSM Institute of Pharmaceutical Sciences [Figure 1, Table 1]. Shavetha et al. showed in an in-vitro study that this particular combination showed increased bioavailability and a higher percentage of drug diffusion and good rheological and texture properties.[5]

# Evaluation of physicochemical parameters of 2% curcumin gel

The formulations were subjected to tests such as homogeneity, spreadabilty, grittiness, extrudability, pH measurement, drug content, and percentage drug release.

#### Spreadability

Spreadabilty was measured by modifying an apparatus suggested by Mutimer et al.<sup>[6]</sup>

#### Homogeneity

Homogeneity was graded. Grades were allotted as +++ good, ++ fair, and + poor.



Figure 1: Curcumin gel

#### Extrudability

The formulation was filled in a clean, lacquered aluminum collapsible one ounce tube with a nasal tip of 5 mm opening. The extrudability was then determined by measuring the amount of gel extruded through the tip when a constant load of 1 kg was placed. The extruded gel was collected and weighed. The percentage of gel extruded was calculated and grades were allotted.<sup>[7]</sup>

#### Determination of pH

About  $5 \pm 0.1$  g of gel is taken in a 100 ml in a beaker and the gel was dispersed in 45 ml of water. pH meter was used to determine the pH.

#### Determination of drug content uniformity

A total of 5 g of the prepared formulation was subjected for analytical assay using UV spectrophotometer at  $\lambda_{\rm max}$  430 nm.

#### Drug release studies

Curcumin release from the gel was studied using permeation apparatus.

# Evaluation of anti-inflammatory activity of 2% curcumin gel

A total of 18 healthy Wistar albino rats of either sex were randomly allocated to test (2% curcumin gel), standard (Voveron® gel), and control group (plain gel) with six animals (n = 6) in each group. The anti-inflammatory activity was assessed by carrageenan induced paw edema method.

# Duration of anti-inflammatory activity of 2% curcumin gel

A total of 36 healthy Wistar albino rats of either sex were divided into six groups (n = 6) based on the time duration at which 2% curcumin gel was applied prior to injection of 0.1% carrageenan. In groups 1-6, 50 mg of 2% curcumin gel was applied at 0, 2, 4, 6, 12, 24, and 48 h prior to administration of carrageenan.

About 50 mg of the 2% gel was divided into two equal parts of 25 mg. The first part of 25 mg gel was applied on the plantar surfaces of their left hind paw surface

Table 1: Formula used to prepare 2% curcumin gel				
Ingredients	Quantity			
Carbapol	2 g			
Polymer (HPMC)	2 g			
Curcumin	2 g			
Propylene glycol	5 ml			
Sodium benzoate	0.2 ml			
Triethnolamine	q.s.			
Distilled water	g s to make 100 ml			

HPMC=Hydroxypropyl methylcellulose; q.s.=Quantity sufficient

by gentle rubbing with the index finger approximately 50 times until no gel was seen or felt on the skin. After 5 min, 25 mg gel was applied in a similar manner at 2, 4, 6, 12, 24, and 48 h prior to injection of 0.1 ml of 1% carrageenan solution. Three hours after the injection the 3<sup>rd</sup> h reading was noted. The percentage inflammation and the percent inhibition of edema at each dosing interval were calculated. In the control group (n = 6) the plain gel and standard (Voveron® gel) group (n = 6) gel was applied 3 h prior to the carrageenan injection.

#### **Experimental periodontitis**

A total of 21 Wistar albino rats (5-10 weeks old) weighing between 150 and 250 g were used in the study. Five rats weighing 150-250 g were taken in a poly propylene cage each day of the experiment for 9 days. The rats were anesthetized with ketamine anesthesia. Preperiodontal examination was done and the upper second molars were ligated using a sterile braided silk suture (5-0) [Figure 2]. Soft tissue indicators were measured.

#### Treatment of experimental periodontitis

Four weeks after ligature placement the rats were divided into three groups. The rats in the control group did not receive any treatment, group 2 received plain gel, group 3 received 2% curcumin gel. The gels were applied with a tuberculin syringe with a blunt tip. The application was done every alternate day for 6 days. The soft tissue indicators were measured prior to euthanizing the rats.<sup>[10]</sup>

#### Soft tissue indicators of periodontitis

#### Gingival index

Gingival index (GI) was recorded on maxillary second molar on four surfaces: Mesial, buccal, distal, and palatal surface. GI was recorded 1 week after ligature placement and 1 week after treatment.<sup>[11]</sup>



Figure 2: Ligature placed

#### Probing pocket depth

Probing pocket depth (PPD) was taken from the gingival margin to the bottom of the pocket using a modified graduated silver cone with the round-ended tip of approximately 0.4 mm in diameter. Six values were recorded and averaged (mesiobuccal, midbuccal, distobuccal, mesiopalatal, midpalatal, and distopalatal). PPD was examined 1 week after ligature placement and 1 week after treatment.<sup>[10]</sup>

#### Sacrificial indicator

#### Morphometric analysis of alveolar bone loss

After the rats were euthanized with ketamine overdose the specimens were dissected carefully to maintain their integrity then, immersed in sodium hypochlorite for 4 h and manual scavenging of the remaining tissue was done. The specimens were stained with methylene blue dye (1 g/100 mL, Sigma-Aldrich, Saint Louis, MO, USA) for 1 min to demarcate the cemento-enamel junction (CEJ) and examined under a stereomicroscope. In order to ensure reproducibility of the alignment of the image, the buccal cusp tip of the first and second molars were placed such that they superimposed on the corresponding lingual/palatal cusp tip. Photographs were obtained with a 6.1-megapixel digital camera (Nikon D100, Ayutthaya, Thailand). Measurements were made on the maxillary second molar in a blinded fashion, 3 times using Java based image processing software, NIH, USA, and the mean values were used in statistical analysis.<sup>[12]</sup> The distance method applied by Crawford, Taubman, and Smith on digitalized images was used to perform linear measurements from the CEJ to the alveolar bone crest, on half of each root following the axis. Six measurements were obtained for the maxillary second molar [Figures 3 and 4]. The data was subjected to statistical analysis.

#### **RESULTS**

All statistical analysis was performed using InStat-GraphPad software [GraphPad software Inc. CA, USA]. P < 0.001 was considered as highly significant and P < 0.05 was considered as significant.

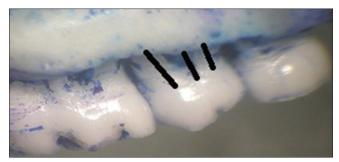


Figure 3: Morphometric analysis: Buccal view

# Evaluation of physicochemical parameters of the 2% curcumin gel

The gels showed optimum spreadability of 15 s, homogeneity of +++ grade, good extrudability. The pH was within the acceptable range of 6.9-7.2 even at the end of 30 days. Curcumin release was 61% [Table 2].

# Evaluation of anti-inflammatory action of 2% curcumin gel

Percentage inflammation was calculated using the formula:

Percentage inflammation =  $\{V - V/V\} \times 100$ 

where  $V_i = 0$  h reading and V = 3 h reading. The average paw thickness in the drug treated group was compared within the control group.

Percent inhibition of the edema was calculated using the formula:

 $\frac{\text{Percentage swelling of drug treated group }(V_{\text{t}})}{\text{Percentage swelling of control group }(V_{\text{c}})} \times 100$ 

The mean weight of the control group was  $238.33 \pm 9.83$  g, the test group was  $237.50 \pm 22.30$  g and the standard was  $227.33 \pm 62.19$  g. Analysis of variance was used for multiple comparisons. F value was 0.151. P value was 0.861. There was no statistically significant difference between the groups.

The mean percentage of inflammation as measured using change in paw thickness in the control, standard,

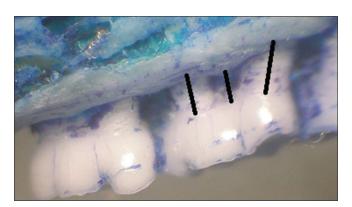


Figure 4: Morphometric analysis: Palatal view

and test group was  $68.38 \pm 25.05$  mm,  $15.74 \pm 4.63$  mm, and  $38.99 \pm 22.20$  mm, respectively. Percent inhibition of edema in the standard group was 76.97% and in the test group was 42.98%. The test group showed moderate anti-inflammatory activity [Tables 3-5].

# Duration of the anti-inflammatory activity of 2% curcumin gel

The gel base was used as control, 1% Voveron® gel was used as standard and the test formulation was 2% curcumin gel. The test groups 1–6, the mean percentage of inflammation was  $36.65 \pm 11.98$  mm,  $23.05 \pm 4.79$ ,  $22.46 \pm 6.63$ ,  $21.78 \pm 7.91$ ,  $19.88 \pm 14.69$ , and  $48.79 \pm 15.72$  mm. Percent inhibition of edema was 46.39%, 66.28%, 67.14%, 68.15%, 70.91%, and 28.65%, respectively [Table 6].

### Soft tissue indicators of periodontitis

#### Gingival index

The mean GI at the end of 7 days was in the control, plain gel, and curcumin gel group was  $2.19 \pm 0.33$ ,  $2.21 \pm 0.33$ , and  $2.67 \pm 0.49$ . The mean GI at the end of 35 days in the control, plain gel, and curcumin gel group was  $3.08 \pm 0.34$ ,  $3.08 \pm 0.36$ , and  $1.23 \pm 0.18$ . The mean percentage change was -40.35%, -39.62%, and 53.75%. There was a statistically significant change in the GI at the end of 35 days, P value was 0.001 [Table 7].

#### Probing pocket depth

Mean PPD in control group, plain gel group, curcumin group, and Tulsi group, 1 week after placement of ligature, was  $2.75\pm0.121, 2.73\pm0.120$  and  $2.681\pm0.098$  mm, respectively. Mean PPD in control group, plain gel group 1 week after placement of ligature, in the control group was, plain gel group and curcumin group was  $2.79\pm0.137, 2.73\pm0.120$  and  $1.79\pm0.094$ , respectively. On comparison between

Table 2: Physicochemical characters of curcumin gel	2%
Dhysicachomical parameters	

Physicochemical parameters	
Homogeneity	++
Grittiness	-
Extrudability	++
Spreadability (s)	15.0
рH	7.0
Drug content	98.1
Percentage release	61

Table 3: Weight of the animals selected for the study								
Weight (g) n Mean SD SE 95% CI for mean							ANOVA F	P value
					Lower bound	Upper bound		
Standard	6	227.3	62.19	25.39	162.06	292.6	1.51	0.86 NS
Test	6	237.5	22.30	9.10	214.09	260.9		
Control	6	238.3	9.832	4.01	228.02	248.6		
Total	18	234.3	36.593	8.625	216.19	252.5		

SD=Standard deviation; SE=Standard error; CI=Confidence interval; ANOVA=Analysis of variance; NS=Not significant

control and plain gel and curcumin gel using Tukey's multiple comparison 1 week after ligature placement, mean difference was 0.018, 0.072, and 0.027, respectively. There was no statistical significant difference between groups [Tables 8 and 9].

#### Sacrificial indicators

#### Morphometric analysis

The mean bone loss at mesiobuccal, midbuccal, distobuccal, mesiopalatal, midpalatal, and distopalatal site treated with plain gel was  $0.30 \pm 0.05$  mm,  $0.28 \pm 0.05$ ,  $0.22 \pm 0.05$ ,  $0.37 \pm 0.06$ ,  $0.29 \pm 0.03$ , and  $0.29 \pm 0.05$  mm respectively.

The mean bone loss at mesiobuccal, midbuccal, distobuccal, mesiopalatal, midpalatal, and distopalatal site treated with 2% curcumin gel was  $0.25 \pm 0.04$  mm,  $0.20 \pm 0.05$ ,  $0.14 \pm 0.03$ ,  $0.35 \pm 0.17$ ,  $0.25 \pm 0.12$ , and  $0.25 \pm 0.12$  mm respectively.

On comparison there was no statistically significant bone loss between the various groups at the mesiobuccal,

Table 4: Percentage inflammation in control, standard, and test (curcumin) group

	Control	Standard	Test (curcumin)
Mean percentage of	68.387±25.05	15.74±4.634	38.992±22.20
inflammation+SEM			

SEM=Standard error of mean

Table 5: Percent inhibition of edema								
Formulation Dose Number Percent Percent (mg/paw) of rats swelling inhibition (mm) of edema								
Control	50	6	38.99±42.98	-				
Standard	50	6	15.74±4.63	76.975				
Test	50	6	38.99±23.22	42.983				

midbuccal, distobuccal, mesiopalatal, midpalatal, and distopalatal site.

Bonferroni's multiple comparisons test was used to compare the effect of control, plain gel and 2% curcumin gel at mesiobuccal, midbuccal, mesiopalatal, midpalatal, and distopalatal sites. There was no statistically significant difference between the mean bone loss at mesiobuccal, midbuccal, mesiopalatal, midpalatal, and distopalatal sites treated with control, plain gel, and 2% curcumin gel [Tables 10-16].

#### Acute oral toxicity

All the animals survived for the period of 14 days. They appeared healthy throughout the study. All the animals appeared to gain weight during the observation period of 14 days. There were no signs of gross toxicity, adverse pharmacological events or changes in behavior. Gross necropsy findings did not show any abnormalities.

# **DISCUSSION**

Periodontitis is a chronic inflammatory disease caused by bacterial infection of the supporting tissues around the teeth. [13,14] The concept of locally delivering chemotherapeutic agents to the periodontal pocket as a method to treat periodontal disease has been studied for over few decades. Several herbal drugs have been an area of interest in the treatment of periodontal diseases. [4]

Periodontal disease can be induced in the rats by tying a ligature of 2-0–5-0 braided silk around the cervix of the maxillary or mandibular molars or by injecting lipopolysaccharides into the papilla or combination of both.<sup>[15]</sup> Souza *et al.* used 4 weeks for periodontitis induction in the maxilla. It was similar to the study period used in our study.<sup>[16]</sup>

Table 6: Duration of anti-inflammatory action of 2% curcumin gel							
Curcumin 2 h 4 h 6 h 12 h 24 h						48 h	
Mean percentage of inflammation±SEM Percent inhibition	36.65±11.98 46.39	23.05±4.79 66.28	22.46±6.63 67.14	21.78±7.918 68.15	19.88±14.69 70.91	48.791±15.72 28.65	
SEM=Standard error of mean	EM=Standard error of mean						

Table 7: Comp	Table 7: Comparison of GI before and after treatment								
Group	n	Minimum	Maximum	Mean	SD	Median	Wilcoxon signed rank test value	P value	
Control group			-						
GI at 7 days	12	2.00	3.00	2.19	0.33	2.00	3.24	0.001** HS	
GI at 35 days	12	2.50	3.50	3.08	0.34	3.00			
Plain gel									
GI at 7 days	12	2.00	3.00	2.21	0.33	2.00	3.11	0.002** HS	
GI at 35 days	12	2.50	3.50	3.08	0.36	3.00			
2% curcumin gel									
GI at 7 days	12	2.00	3.00	2.67	0.49	3.00	1.43	0.002** HS	
GI at 35 days	12	1.00	1.50	1.23	0.18	1.20			

GI=Gingival index; SD=Standard deviation; \*\*HS=Highly significant

Table 8: Probi	ng pocket depth					
Mean probing	Mean±SD					
depth	7 days postligature placement	At the end of the treatment				
Control	2.75±0.121	2.79±0.137				
Plain	2.73±0.120	2.73±0.120				
Curcumin gel	2.681±0.098	1.79±0.094				

SD=Standard deviation

Table 9: Comparison between control and treatment group at the end of treatment

Turkey's multiple comparisons test	Mean difference	95% CI of difference	Summary
Control versus plain gel	0.05455	-0.1094 to 0.2185	NS
Control versus curcumin gel	0.6818	0.5178 to 0.8458	HS**

CI=Confidence interval; NS=Not significant; \*\*HS=Highly significant

Table 10: Mean bone level at sites treated with control, plain gel, and 2% curcumin gel

Site		Mean±SD	
	Control	Plain gel	Curcumin gel
Mesiobuccal	0.30±0.056	0.30±0.055	0.25±0.046
Midbuccal	0.28±0.056	0.24±0.064	0.20±0.055
Distobuccal	0.22±0.057	0.17±0.036	0.14±0.031
Mesiopalatal	0.37±0.063	0.42±0.203	0.35±0.171
Midpalatal	0.29±0.030	0.33±0.152	0.25±0.129
Distopalatal	0.29±0.053	0.31±0.153	0.25±0.126

SD=Standard deviation

# Table 11: Comparison of the effect of control, plain gel, and 2% curcumin gel on bone loss (mesiobuccal site)

Bonferroni's multiple comparisons test	Mean difference	95% CI of difference	Statistical significance
Mesiobuccal Control versus plain gel	0.0492	-0.05154 to 0.1499	NS
Control versus curcumin gel	0.0534	-0.04734 to 0.1541	NS
Plain gel versus curcumin gel	0.0042	-0.09654 to 0.1049	NS

CI=Confidence interval; NS=Not significant

Studies have supported the positive role of anti-inflammatory agents in the treatment of periodontal disease.[17] The anti-inflammatory properties of curcumin are mediated by the modulating the activity of signaling pathways and transcription factors, especially nuclear factor kappa-β, activating protein-1 and mitogen-activated protein kinases. Curcumin suppresses the expression of interleukin (IL)-6, IL-1 $\beta$ , tumor necrosis factor- $\alpha$ , prostaglandins, matrix metalloproteinase 2 (MMP-2) and MMP-9.[18-23] Curcumin was also shown to improve wound healing by increasing

Table 12: Comparison of the effect of control, plain gel, and 2% curcumin gel on bone loss (midbuccal site)

Bonferroni's multiple comparisons test	Mean difference	95% CI of difference	Statistical significance
Midbuccal			
Control versus	0.0734	-0.02734 to 0.1741	NS
plain gel			
Control versus	0.0793	-0.02144 to 0.1800	NS
curcumin gel			
Plain gel versus	0.0059	-0.09484 to 0.1066	NS
curcumin gel			

CI=Confidence interval; NS=Not significant

### Table 13: Comparison of the effect of control, plain gel and 2% curcumin gel on bone loss (distobuccal site)

Bonferroni's multiple comparisons test	Mean difference	95% CI of difference	Statistical significance
Distobuccal			
Control versus	0.0804	-0.02034 to 0.1811	NS
plain gel			
Control versus	0.0814	-0.01934 to 0.1821	NS
curcumin gel			
Plain gel versus	0.001	-0.09974 to 0.1017	NS
curcumin gel			

CI=Confidence interval; NS=Not significant

# Table 14: Comparison of the effect of control, plain gel, and 2% curcumin gel on bone loss (mesiopalatal site)

Mean difference	95% CI of difference	Statistical significance
0.0169	-0.08384 to 0.1176	NS
0.0197	-0.08104 to 0.1204	NS
0.0028	-0.09794 to 0.1035	NS
	0.0169 0.0197	difference difference   0.0169 -0.08384 to 0.1176   0.0197 -0.08104 to 0.1204   0.0028 -0.09794 to 0.1035

CI=Confidence interval; NS=Not significant

### Table 15: Comparison of the effect of control, plain gel, and 2% curcumin gel on bone loss (mid palatal site)

multiple comparisons test	difference	difference	significance	
Midpalatal				
Control versus	0.0191	-0.08164 to 0.1198	NS	
plain gel				
Control versus	0.0417	-0.05904 to 0.1424	NS	
curcumin gel				
Plain gel versus	0.0226	-0.07814 to 0.1233	NS	
curcumin gel				

CI=Confidence interval; NS=Not significant

Table 16: Comparison of the effect of control, plain gel, and 2% curcumin gel on bone loss (distopalatal site)

Bonferroni's multiple comparisons test	Mean difference	95% CI of difference	Statistical significance
Distopalatal			
Control versus	0.0367	-0.06404 to 0.1374	NS
plain gel			
Control versus	0.0461	-0.05464 to 0.1468	NS
curcumin gel			
Plain gel versus	0.0094	-0.09134 to 0.1101	NS
curcumin gel			

CI=Confidence interval; NS=Not significant

collagen deposition, angiogenesis and the density of fibroblasts, reducing the radiation-induced delay in wound repair. [24] These pharmacological properties make it an effective drug to treat periodontal disease and hence, we fabricated a 2% curcumin gel by simple dispersion method.

The gel showed a standard physicochemical profile with a drug release of 61% inhibition of edema was 42.98%. Maximum inhibition was seen at 24 h. The anti-inflammatory activity lasted for 48 h. Hence, the gel was applied on alternate days to treat experimental periodontitis.

The GI scores in the control group and plain gel group increased indicating that the inflammation progressed. In the 2% curcumin gel group, a highly significant mean percentage decrease was noted. This showed that the curcumin gel had good anti-inflammatory effect.

Morphometric analysis did not show any significant difference between the groups. Whether or not the local drug delivery (LDD) system results in bone regeneration is controversial. In our study, we did not notice any significant difference in the residual periodontal bone level among various groups.

#### Limitations of the study

- Pressure sensitive probe and stent needs to be used
- No scaling and root planning was done
- Microbial profiling, biochemical, and immuno-histochemical parameters have to be evaluated
- It is worthwhile to do chronic toxicity study. But, the animals were monitored closely during the entire study period for any signs of toxicity. No signs of chronic toxicity were noted.

#### CONCLUSION

About 2% curcumin gel has shown good anti-inflammatory effect for 24-48 h despite the limitations in our study. The

2% curcumin gel showed significant reduction in gingival inflammation and pocket depth. The periodontal bone loss was not statistically significant. This is consistent with the use of LDD systems as they help in disease limitation and not significant bone regeneration. Hence, 2% curcumin can be used as useful adjunct to enhance the results of standard periodontal therapy.

### **ACKNOWLEDGMENTS**

I would like to acknowledge the technical and intellectual support provided by Dr. B. H. Sripathi Rao, Principal and Dean, Dr. A. Shashikanth Hegde, Prof. and Head, Dr. K. S. Rajesh, Sr. Professor, Department of Periodontics, Dr. Maji Jose, Prof. and Head, Department of Oral and Maxillofacial Pathology, Yenepoya Dental College, Mangalore, Mr. E. P. Rejeesh, Department of Pharmacology, Yenepoya Medical College, Dr. Sucharita Suresh, statistician, Father Mullers College, Mangalore.

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**Cite this article as:** Hosadurga RR, Rao SN, Jose J, Rompicharla NC, Shakil M, Shashidhara R. Evaluation of the efficacy of 2% curcumin gel in the treatment of experimental periodontitis. Phcog Res 2014;6:326-33.

Source of Support: Nil, Conflict of Interest: None declared.