

# Antiproliferative and phytochemical analyses of leaf extracts of ten Apocynaceae species

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## ABSTRACT

**Background:** The anticancer properties of Apocynaceae species are well known in barks and roots but less so in leaves. **Materials and Methods:** In this study, leaf extracts of 10 Apocynaceae species were assessed for antiproliferative (APF) activities using the sulforhodamine B assay. Their extracts were also analyzed for total alkaloid content (TAC), total phenolic content (TPC), and radical scavenging activity (RSA) using the Dragendorff precipitation, Folin–Ciocalteu, and 1,1-diphenyl-2-picrylhydrazyl (DPPH) assays, respectively. **Results:** Leaf extracts of *Alstonia angustiloba*, *Calotropis gigantea*, *Catharanthus roseus*, *Nerium oleander*, *Plumeria obtusa*, and *Vallis glabra* displayed positive APF activities. Extracts of *Allamanda cathartica*, *Cerbera odollam*, *Dyera costulata*, and *Kopsia fruticosa* did not show any APF activity. Dichloromethane (DCM) extract of *C. gigantea*, and DCM and DCM:MeOH extracts of *V. glabra* showed strong APF activities against all six human cancer cell lines. Against breast cancer cells of MCF-7 and MDA-MB-231, DCM extracts of *C. gigantea* and *N. oleander* were stronger than or comparable to standard drugs of xanthorrhizol, curcumin, and tamoxifen. All four extracts of *N. oleander* were effective against MCF-7 cells. Extracts of *Kopsia fruticosa* had the highest TAC while those of *Dyera costulata* had the highest TPC and RSA. Extracts of *C. gigantea* and *V. glabra* inhibited the growth of all six cancer cell lines while all extracts of *N. oleander* were effective against MCF-7 cells. **Conclusion:** Extracts of *C. gigantea*, *V. glabra*, and *N. oleander* therefore showed great promise as potential candidates for anticancer drugs. The wide-spectrum APF activities of these three species are reported for the first time and their bioactive compounds warrant further investigation.

**Key words:** Antiproliferative, Apocynaceae, radical scavenging, total alkaloid content, total phenolic content

## INTRODUCTION

The family Apocynaceae consists of about 250 genera and 2000 species of tropical trees, shrubs, and vines.<sup>[1]</sup> With the inclusion of species of Asclepiadaceae, the family has now been enlarged from two to five subfamilies.<sup>[2]</sup> Characteristic features of the family are that almost all species produce milky sap; leaves are simple, opposite, or whorled; flowers are large, colorful, and slightly fragrant with five contorted lobes; and fruits are in pairs.<sup>[1,3]</sup>

In traditional medicine, Apocynaceae species are used to treat gastrointestinal ailments, fever, malaria, pain, and

diabetes.<sup>[1]</sup> Of the 10 species studied, leaves of *Allamanda cathartica* are used as a purgative or emetic in Southeast Asia.<sup>[4]</sup> Leaves are also used as an antidote, and for relieving coughs and headaches. Stems, leaves, and latex of *Alstonia angustiloba* are used for gynecological problems and skin sores in Indonesia.<sup>[5]</sup> Leaves are externally applied to treat headache in Malaysia.<sup>[6]</sup> Roots and leaves of *Calotropis gigantea* are used to treat skin and liver diseases, leprosy, dysentery, worms, ulcers, tumours, and earache.<sup>[7]</sup> Its latex has wound-healing properties.<sup>[8]</sup> A decoction of all parts of *Catharanthus roseus* is used to treat malaria, diarrhea, diabetes, cancer, and skin diseases.<sup>[9]</sup> The species is also well known as an oral hypoglycemic agent. Extracts prepared from leaves have been used as an antiseptic agent for healing wounds and as a mouthwash to treat toothache. In Southeast Asia, leaves of *Cerbera odollam* are used in aromatic bath by women after childbirth.<sup>[10]</sup> Leaves, bark and latex are emetic and purgative, and seeds are toxic and

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strongly purgative. Leaves and bark of *Dyera costulata* have been used for treating fever, inflammation, and pain.<sup>[11]</sup> *Kopsia fruticosa* has cholinergic effects and is used to treat sore and syphilis.<sup>[12]</sup> *Nerium oleander* is highly poisonous with no reported benefits in traditional medicine. In Asia, a decoction of leaves of *Plumeria obtusa* is used for treating wounds and skin diseases.<sup>[13]</sup> Its latex and bark are known to have purgative and diuretic properties. *Vallaris glabra* is well known in Thailand because the scent of its flowers is similar to that of pandan leaves and aromatic rice.<sup>[14]</sup> Its use in traditional medicine is not known.

Apocynaceae species have been reported to possess anticancer properties. They include those of *Allamanda*,<sup>[15]</sup> *Alstonia*,<sup>[16,17]</sup> *Calotropis*,<sup>[18-20]</sup> *Catharanthus*,<sup>[21]</sup> *Cerbera*,<sup>[22,23]</sup> *Nerium*,<sup>[24,25]</sup> *Plumeria*,<sup>[26]</sup> and *Tabernaemontana*.<sup>[27]</sup>

Prompted by the anticancer properties found in many species of Apocynaceae, leaf extracts of 10 species were assessed for antiproliferative activity against six human cancer cell lines. Their extracts were also analyzed for total alkaloid content, total phenolic content, and radical scavenging activity.

## MATERIALS AND METHODS

### Plant materials

The 10 Apocynaceae species studied were *A. cathartica*, *Alstonia angustiloba*, *Calotropis gigantea*, *Catharanthus roseus*, *Cerbera odollam*, *Dyera costulata*, *Kopsia fruticosa*, *Nerium oleander*, *Plumeria obtusa*, and *Vallaris glabra* [Figure 1]. Their common or vernacular names and brief descriptions are given in [Table 1]. Leaves of the species were collected from Sunway, Puchong, or Kepong, all in the state of Selangor, Malaysia. Identification of species was based on

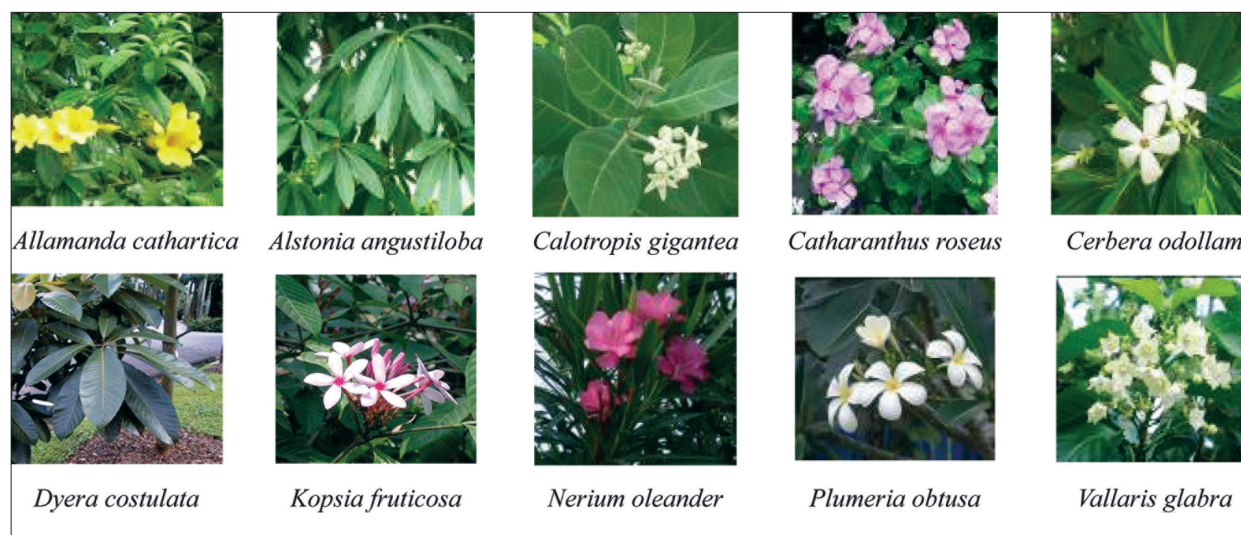
documented descriptions and illustrations.<sup>[1-3]</sup> The voucher specimens of these species were deposited in the herbarium of Monash University Sunway Campus.

### Extraction of leaves

For crude extraction, fresh leaves of each species (40 g) were cut into small pieces and freeze-dried overnight. Dried samples were blended and extracted with 250 ml of methanol (MeOH) three times for 1 h each time. Samples were filtered and the solvent was removed using a rotary evaporator (Eyela). The dried crude extracts were stored at  $-20^{\circ}\text{C}$  for further analysis. For sequential extraction, fresh leaves of each species (40 g) were freeze-dried, ground, and extracted successively with hexane, dichloromethane (DCM), DCM:MeOH (1:1), and MeOH (HmbG Chemicals). For each solvent, the suspension of ground leaves in 250–300 ml of solvent was shaken for 1 h on the orbital shaker. After filtering, the samples were extracted two more times for each solvent. Solvents were removed with a rotary evaporator to obtain the dried extracts, which were stored at  $-20^{\circ}\text{C}$  for further analysis.

### Antiproliferative activity

Antiproliferative (APF) activity of extracts (25  $\mu\text{g}/\text{ml}$ ) was initially screened for growth inhibitory activity against three human cancer cell lines (MCF-7, MDA-MB-231, and HeLa) using the sulforhodamine B (SRB) assay.<sup>[28-30]</sup> Growth inhibitory activity with less than 50% cell growth was considered positive while that with more than 50% cell growth was considered negative. Extracts with positive growth inhibition were further tested against six human cancer lines (MCF-7, MDA-MB-231, HeLa, HT-29, SKOV-3, and HepG2) using six different extract concentrations. Human cancer cell lines were obtained from the American Type Culture Collection (ATCC, Manassas, VA, USA). The



**Figure 1:** The ten Apocynaceae species studied

**Table 1: Common or vernacular names and brief description of Apocynaceae species studied**

Species (common/vernacular name)	Brief description
<i>Allamanda cathartica</i> L. (Common allamanda)	A shrub with trumpet-shaped yellow flowers similar in size as leaves which are in whorls
<i>Alstonia angustiloba</i> Miq. (Pulai)	A medium-sized tree with leaves in whorls and having fine secondary veins
<i>Calotropis gigantea</i> (L.) Aiton (Giant milkweed)	A shrub with pale green leaves and white or lilac flowers with a crown rising from the center
<i>Catharanthus roseus</i> (L.) G. Don (Madagascar periwinkle)	A common ornamental shrub with oval to oblong leaves and white, pink or purple flowers with a dark colored center
<i>Cerbera odollam</i> Gaertner (Pong-pong)	A tree bearing white flowers in clusters and rounded fruits that are green when young and red when mature
<i>Dyera costulata</i> Hook (Jelutong)	A tall timber tree with straight columnar bole, leaves in whorls and latex which was an important source of chewing gum
<i>Kopsia fruticosa</i> (Ker.) A. DC. (Pink kopsia)	A shrub with large glossy leaves and clusters of light pink flowers resembling those of <i>Ixora</i>
<i>Nerium oleander</i> L. (Oleander)	An ornamental shrub with thick narrow leaves in pairs or whorls and bearing clusters of pink, red, or purple flowers
<i>Plumeria obtusa</i> L. (Frangipanni)	A tree producing dark green, glossy and oval leaves and white fragrant flowers with a yellow center
<i>Vallisneria spiralis</i> L. (Kesidang)	A woody climber producing clusters of white flowers with a scent characteristic of pandan leaves or fragrant rice

cells were seeded 24 h prior to treatment in 96-well plates at densities of 10,000–20,000 cells/well. Each cell line was designated one plate. Initial cell population of each cell lines prior to addition of extracts was determined by fixing with trichloroacetic acid (TCA) (Sigma). Extracts were dissolved in dimethyl sulfoxide (DMSO) (Sigma) and serially diluted from 8–25 µg/ml. Control cultures were treated with the same volume of DMSO. The concentration of DMSO was kept within 1% to avoid any interference with cell viability. After the addition of extracts, the plates were incubated for 48 h. After incubation, the cells were fixed with 50 µl of cold 50% TCA and incubated for 1 h at 4°C. The plates were then washed with tap water and air dried. Cells were stained with 100 µl of 0.4% SRB solution (Sigma) diluted with 1% acetic acid followed by incubation for 10 min at room temperature. Unbound dye was removed by washing with 1% acetic acid. Bound stain was then solubilized with 200 µl of 10 mM trizma base (Sigma). Absorbance of each well at 505 nm was obtained using a microplate reader. Dose–response curves were constructed to obtain GI<sub>50</sub> or growth inhibition of cell lines by 50%. Activity is considered to be effective when GI<sub>50</sub> value ≤20 µg/ml.<sup>[31]</sup>

### Analysis of TAC, TPC, and RSA

Total alkaloid content (TAC) of extracts was determined using the Dragendorff precipitation assay.<sup>[32]</sup> For each species, extracts (15 mg) were dissolved in 1 ml of distilled water that was acidified to pH 2.0–2.5 with 0.01 M HCl. Analysis was conducted in triplicate. Alkaloids were then precipitated with 0.4 ml of Dragendorff reagent. Washed with 0.5 ml of distilled water to remove traces of the reagent, the precipitate was later treated with 0.4 ml of 1% sodium sulfide, resulting in a brownish-black precipitate. Precipitates formed at each stage were recovered by centrifugation at 14,000 rpm for 1 min. The resulting

precipitate was dissolved in 0.2 ml of concentrated nitric acid and diluted to 1 ml with distilled water. Addition of 2.5 ml of 3% thiourea to 0.5 ml aliquots of this solution resulted in a yellow-colored complex. Absorbance was measured at 435 nm and TAC was expressed as boldine equivalent in milligram per gram of extract. The calibration equation for boldine (Sigma) was  $y = 1.068x$  ( $R^2 = 0.9959$ ) where  $y$  is absorbance and  $x$  is mg/ml of boldine. Dragendorff reagent was prepared by dissolving 0.8 g of bismuth nitrate (Sigma) in 40 ml of distilled water and 10 ml of glacial acetic acid. The resulting solution was mixed with 20 ml of 40% potassium iodide.

Total phenolic content (TPC) of extracts was determined using the Folin–Ciocalteu (FC) assay.<sup>[33]</sup> Extracts (300 µl in triplicate) were introduced into test tubes followed by 1.5 ml of FC reagent (Fluka) at 10 times dilution and 1.2 ml of sodium carbonate (Fluka) at 7.5% w/v. The tubes were allowed to stand for 30 min in the dark before absorbance was measured at 765 nm. TPC was expressed as gallic acid (GA) equivalent in milligram per gram of extract. The calibration equation for GA (Fluka) was  $y = 0.0111x - 0.0148$  ( $R^2 = 0.9998$ ) where  $y$  is absorbance and  $x$  is mg/ml of GA.

Radical scavenging activity (RSA) of extracts was determined using the 1,1-diphenyl-2-picrylhydrazyl (DPPH) assay.<sup>[33]</sup> Different dilutions of extracts (1 ml in triplicate) were added to 2 ml of DPPH (Sigma). The concentration of DPPH used was 5.9 mg in 100 ml of methanol. Absorbance was measured at 517 nm after 30 min. RSA was calculated as IC<sub>50</sub>, the concentration of extract to scavenge 50% of the DPPH radical. RSA was then expressed as ascorbic acid equivalent antioxidant capacity (AEAC) using the equation of AEAC (mg ascorbic

acid/g of extract) =  $IC_{50(\text{ascorbate})}/IC_{50(\text{sample})} \times 10^5$ .  $IC_{50}$  of ascorbic acid used for calculation of AEAC was 0.00387 mg/ml.

## RESULTS AND DISCUSSION

Initial screening of leaf extracts of 10 Apocynaceae species against three human cancer cell lines (MCF-7, MDA-MB-231, and HeLa) showed that extracts of *A. angustiloba*, *C. gigantea*, *C. roseus*, *N. oleander*, *P. obtusa*, and *V. glabra* displayed positive growth inhibitory activity, that is, inhibition with  $\leq 50\%$  cell growth [Table 2]. DCM extract of *C. gigantea*, and DCM and DCM:MeOH extracts of *N. oleander* and *V. glabra* inhibited all three cancer cell lines. All four extracts of *N. oleander* were effective against MCF-7 cells. Only DCM:MeOH extract of *C. roseus* was active against MCF-7 and HeLa cells. Hexane extract of *N. oleander* inhibited MCF-7 cells while hexane extract of *P. obtusa* inhibited MCF-7 and HeLa cells. MeOH extracts of *N. oleander* and *V. glabra* inhibited MCF-7 cells. In general, DCM and DCM:MeOH extracts of these species were the most effective. All extracts of *A. cathartica*, *C. odollam*, *D. costulata*, and *K. fruticosus* did not show any APF activity.

Contrary to findings of this study, cytotoxic activities have been reported in species of *Cerbera*, *Allamanda*, and *Kopsia*. Methanol extract of leaves of *C. odollam* strongly inhibited MCF-7 and T47D cells.<sup>[23]</sup> Cardenolides from seeds of *C. odollam* had cytotoxic activity against KB, BC, and NCI-H187 cells.<sup>[34]</sup> Potent cytotoxic activity was reported in ethanol extracts of fruits and leaves of *Cerbera*

*manghas*.<sup>[35]</sup> Cardenolides from roots of *C. manghas* also showed APF activity.<sup>[22]</sup> Ethanol root extracts of *A. schottii* and *A. blanchetti* displayed stronger cytotoxicity against K-562 cells than leaf and stem extracts.<sup>[15]</sup> Valparicine from the stem bark of *Kopsia arborea* showed cytotoxic effects against KB and Jurkat cells<sup>[36]</sup> while kopsimaline from leaves and stem bark of *Kopsia singaporensis* was found to inhibit KB cells.<sup>[37]</sup>

Extracts of the six species were further tested against six human cancer cell lines (MCF-7, MDA-MB-231, HeLa, HT-29, SKOV-3, and HepG2). Results showed that DCM extract of *A. angustiloba* inhibited only MDA-MB-231, HeLa, and SKOV-3 cells with  $GI_{50}$  values of 20, 20, and 16  $\mu\text{g/ml}$ , respectively [Table 3]. DCM extract of *C. gigantea* strongly inhibited all cancer cell lines with  $GI_{50}$  values ranging from 1.3–3.3  $\mu\text{g/ml}$ . Against MCF-7 and MDA-MB-231 cells,  $GI_{50}$  of DCM extract of *C. gigantea* (1.9 and 1.3  $\mu\text{g/ml}$ ) was much stronger than that of xanthorrhizol (11 and 8.7  $\mu\text{g/ml}$ ), curcumin (4.1 and 8.7  $\mu\text{g/ml}$ ), and tamoxifen (8.3 and 4.6  $\mu\text{g/ml}$ ), respectively.<sup>[30]</sup> DCM:MeOH extract of *C. roseus* strongly inhibited MCF-7 and HeLa cells with  $GI_{50}$  of 3.5 and 4.7  $\mu\text{g/ml}$ , respectively. All four extracts of *N. oleander* were effective against MCF-7 cells with  $GI_{50}$  ranging from 3.7–12  $\mu\text{g/ml}$ . DCM and DCM:MeOH extracts inhibited all cell lines except HepG2 cells. Hexane extract of *P. obtusa* was effective against MCF-7 and HeLa cells, while its DCM extract was effective against HeLa cells. DCM and DCM:MeOH extracts of *V. glabra* inhibited all cell lines with  $GI_{50}$  values ranging from 7.5–12  $\mu\text{g/ml}$  and 5.8–13  $\mu\text{g/ml}$ , respectively. In addition, MeOH extract of *V. glabra* also inhibited the growth of MCF-7 and HepG2 cells. Against MCF-7 cells,  $GI_{50}$  of DCM and DCM:MeOH extracts of *V. glabra* (7.7 and 7.0  $\mu\text{g/ml}$ ) was stronger than xanthorrhizol (11  $\mu\text{g/ml}$ ) and comparable to tamoxifen (8.3  $\mu\text{g/ml}$ ), respectively.<sup>[30]</sup>

To the best of our knowledge, this study represents the first report of APF activities from leaf extracts of *A. angustiloba*, *P. obtusa*, and *V. glabra*. Earlier studies have reported cytotoxic activity from the root bark of *Alstonia macrophylla*<sup>[16]</sup> and from the stem bark of *Alstonia scholaris*.<sup>[17]</sup> Iridoids isolated from the bark of *Plumeria rubra* were cytotoxic.<sup>[26]</sup> A recent study reported potent cell growth inhibition of cardenolide glycosides isolated from *Vallisneria spiralis*.<sup>[38]</sup> The finding of strong APF activities from DCM and DCM:MeOH extracts of *C. gigantea* from this study is supported by earlier reports that leaf and root extracts of *C. gigantea* had strong inhibitory activity against cancer cells.<sup>[18-20]</sup> Isolated from leaves of *N. oleander*, pentacyclic triterpenoids were cytotoxic to KB cells.<sup>[24]</sup> Against HL60 and K562 cells, the stem extract of *N. oleander* displayed stronger cytotoxic activity than leaf and root extracts.<sup>[39]</sup>

**Table 2: Leaf extracts of Apocynaceae species with positive growth inhibitory activity against three human cancer cell lines**

Species	Leaf extract <sup>3</sup>	Growth inhibitory activity <sup>1</sup>		
		MCF-7 <sup>2</sup>	MDA-MB-231 <sup>2</sup>	HeLa <sup>2</sup>
<i>Alstonia angustiloba</i>	DCM	-	+	+
<i>Calotropis gigantea</i>	DCM	+	+	+
<i>Catharanthus roseus</i>	DCM:MeOH	+	-	+
<i>Nerium oleander</i>	DCM:MeOH	+	-	+
	MeOH	+	-	-
<i>Plumeria obtusa</i>	HEX	+	-	+
<i>Vallisneria glabra</i>	DCM	-	-	+
	DCM	+	+	+
	DCM:MeOH	+	+	+
	MeOH	+	-	-

<sup>1</sup>Growth inhibitory activity with  $<50\%$  cell growth is considered positive (+) while that with  $>50\%$  cell growth is considered negative (-); <sup>2</sup>MCF-7 and MDA-MB-231 are human breast cancer cells, and HeLa are human cervical cancer cells; <sup>3</sup>HEX = Hexane, DCM = Dichloromethane, and MeOH = Methanol.

**Table 3: Antiproliferative activity of leaf extracts of Apocynaceae species with positive growth inhibitory activity against six human cancer cell lines**

Species	Leaf extract <sup>3</sup>	Growth inhibition (GI <sub>50</sub> ) <sup>1</sup>					
		MCF-7 <sup>2</sup>	MDA-MB-231 <sup>2</sup>	HeLa <sup>2</sup>	SKOV-3 <sup>2</sup>	HT-29 <sup>2</sup>	HepG2 <sup>2</sup>
<i>Alstonia angustiloba</i>	DCM	-	20 ± 1.7	20 ± 1.1	16 ± 1.4	-	-
<i>Calotropis gigantea</i>	DCM	1.9 ± 0.2	1.3 ± 0.3	2.5 ± 0.5	2.5 ± 0.2	3.3 ± 0.2	1.8 ± 1.7
	DCM:MeOH	13 ± 0.3	-	15 ± 1.0	20 ± 2.3	24 ± 0.7	16 ± 3.5
<i>Catharanthus roseus</i>	DCM:MeOH	3.5 ± 0.1	-	4.7 ± 0.6	-	-	-
<i>Nerium oleander</i>	HEX	11 ± 0.9	-	-	12 ± 2.8	-	-
	DCM	3.7 ± 0.1	5.2 ± 0.7	5.1 ± 0.7	4.5 ± 1.0	6.1 ± 0.9	-
	DCM:MeOH	4.3 ± 0.2	18 ± 2.3	6.8 ± 0.9	7.6 ± 2.1	9.2 ± 2.2	-
	MeOH	12 ± 1.1	-	-	-	-	-
<i>Plumeria obtusa</i>	HEX	5.7 ± 0.8	-	10 ± 1.4	-	-	-
	DCM	-	-	19 ± 4.0	-	-	-
<i>Vallis glabra</i>	DCM	7.7 ± 1.3	12 ± 2.0	9.8 ± 1.5	7.5 ± 4.5	9.3 ± 2.0	7.6 ± 0.2
	DCM:MeOH	7.0 ± 2.5	13 ± 6.3	8.5 ± 2.9	7.7 ± 2.4	12 ± 1.2	5.8 ± 1.2
	MeOH	16 ± 2.1	-	-	-	-	19 ± 0.9

<sup>1</sup>GI<sub>50</sub> (µg/ml) is growth inhibition of cancer cell lines by 50% and inhibition is not effective (-) if values >20 µg/ml; <sup>2</sup>MCF-7 and MDA-MB-231, HeLa and SKOV-3, HT-29, and HepG2 are human breast, cervical, colon, and liver cancer cells, respectively; <sup>3</sup>HEX = Hexane, DCM = Dichloromethane, and MeOH = Methanol

It is interesting to note that out of four leaf extracts of *C. roseus* tested against six cell lines, only DCM:MeOH extract inhibited MCF-7 and HeLa cells. The species is well known for its indole alkaloids notably vinblastine and vincristine, which are used to treat Hodgkin's disease and acute leukemia in children, respectively.<sup>[1]</sup> It can be inferred that the APF activities of *C. roseus* may be cell line specific unlike those of *C. gigantea*, *N. oleander*, and *V. glabra* which are wide spectrum, inhibiting most or all cell lines tested.

Of the 10 species analyzed, MeOH crude and DCM extracts of *K. fruticososa* had the highest TAC (100 and 129 mg BE/g of extract), respectively [Table 4]. Other species with moderately high TAC were *D. costulata*, *C. roseus*, and *A. angustiloba* with MeOH crude and DCM:MeOH extracts having values ranging from 23–58 and 58–68 mg BE/g of extract, respectively. Based on TAC, the species can be ranked as high (*K. fruticososa*), moderate (*D. costulata*, *C. roseus*, and *A. angustiloba*), and low (*C. odollam*, *C. gigantea*, *V. glabra*, *P. obtusa*, *A. cathartica*, and *N. oleander*). Extracts of *D. costulata* had the highest TPC and strongest RSA. MeOH crude, DCM:MeOH, and MeOH extracts of *D. costulata* yielded TPC values of 319, 354, and 279 mg GAE/g of extract, and RSA values of 377, 349, and 278 mg AA/g of extract, respectively. Compared to *D. costulata*, extracts of other species can be categorized as moderate to low.

The high TAC of leaf extracts of *K. fruticososa* may be attributed to the presence of alkaloids identified as fruticosamine, fruticosine, and kopsine.<sup>[40]</sup> The presence of flavonols identified as 3',7'-dimethoxyquercetin and quercetin-3-O- $\alpha$ -L-rhamnopyranoside<sup>[11]</sup> may contribute to the high TPC and RSA of leaf extracts of *D. costulata*.

Overall, there is a strong correlation between TPC and RSA of extracts ( $R^2 = 0.992$ ) but not with TAC ( $R^2 =$

0.135). The strong correlation between TPC and RSA of extracts affirms that Apocynaceae species with higher concentration of phenolic compounds in the leaves also have stronger radical scavenging capacity. This would mean that the phenolic compounds are the main contributors to the antioxidant potential of leaves. Similar findings have been reported in medicinal plants and herbs,<sup>[41-43]</sup> plants of industrial interest,<sup>[44]</sup> wild edible fruits,<sup>[45]</sup> and mushrooms.<sup>[46]</sup> The correlation of results of phytochemical analysis with APF activities remains unclear. Extracts of *C. gigantea*, *N. oleander*, and *V. glabra* which showed strong APF activities had low TAC, and moderate to low TPC and RSA.

## CONCLUSION

Out of 10 species of Apocynaceae, leaf extracts of *A. angustiloba*, *C. gigantea*, *C. roseus*, *N. oleander*, *P. obtusa*, and *V. glabra* displayed positive APF activities. DCM extract of *C. gigantea*, and DCM and DCM:MeOH extracts of *V. glabra* inhibited the growth of all six human cancer cell lines. Against MCF-7 and MDA-MB-231 breast cancer cells, DCM extracts of *C. gigantea* and *N. oleander* were stronger than or comparable to standard drugs of xanthorrhizol, curcumin, and tamoxifen. All four extracts of *N. oleander* were effective against MCF-7 cells. With wide-spectrum APF activities, leaves of these three species are therefore promising candidates as alternative resources for anticancer drugs. Their wide-spectrum APF activities are reported for the first time and this warrants further investigation into their bioactive compounds.

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**Table 4: Total alkaloid content, total phenolic content, and radical scavenging activity of leaf extracts of Apocynaceae species**

Species	MeOH crude extract <sup>1</sup>	Sequential extract <sup>1</sup>			
		HEX	DCM	DCM:MeOH	MeOH
TAC (BE mg/g extract) <sup>2</sup>					
<i>Kopsia fruticosa</i>	100 ± 4.2	63 ± 1.1	129 ± 4.0	99 ± 2.5	46 ± 1.6
<i>Dyera costulata</i>	58 ± 2.5	2.4 ± 0.6	11 ± 1.8	68 ± 2.7	41 ± 2.4
<i>Catharanthus roseus</i>	37 ± 2.8	18 ± 0.9	9.1 ± 0.8	76 ± 5.5	35 ± 3.3
<i>Alstonia angustiloba</i>	23 ± 0.3	13 ± 2.3	27 ± 3.1	58 ± 1.2	27 ± 1.0
<i>Cerbera odollam</i>	2.8 ± 0.9	3.6 ± 0.5	3.0 ± 0.5	5.0 ± 1.5	3.2 ± 0.4
<i>Calotropis gigantea</i>	2.7 ± 0.3	3.7 ± 1.0	8.6 ± 1.2	9.6 ± 1.9	9.2 ± 2.7
<i>Vallis glabra</i>	2.7 ± 0.4	4.4 ± 0.8	8.9 ± 0.6	9.2 ± 0.2	8.7 ± 2.2
<i>Plumeria obtusa</i>	2.3 ± 0.3	2.3 ± 0.4	2.5 ± 0.1	4.3 ± 1.3	5.8 ± 1.7
<i>Allamanda cathartica</i>	2.0 ± 0.1	4.4 ± 1.3	8.7 ± 0.9	12 ± 1.0	8.7 ± 2.1
<i>Nerium oleander</i>	1.4 ± 0.7	1.6 ± 0.0	2.8 ± 0.8	3.9 ± 0.7	8.3 ± 0.5
TPC (GAE mg/g extract) <sup>2</sup>					
<i>Dyera costulata</i>	319 ± 5.5	21 ± 0.2	23 ± 0.1	354 ± 6.0	279 ± 3.0
<i>Vallis glabra</i>	99 ± 3.8	15 ± 0.9	24 ± 0.5	134 ± 1.0	164 ± 13
<i>Plumeria obtusa</i>	85 ± 0.9	21 ± 1.7	52 ± 1.3	104 ± 4.0	134 ± 3.0
<i>Kopsia fruticosa</i>	83 ± 1.1	39 ± 0.5	20 ± 0.5	129 ± 1.0	84 ± 1.0
<i>Alstonia angustiloba</i>	68 ± 2.0	17 ± 0.6	24 ± 0.6	96 ± 1.1	94 ± 1.1
<i>Nerium oleander</i>	56 ± 3.8	18 ± 0.3	23 ± 0.1	57 ± 0.3	29 ± 0.3
<i>Catharanthus roseus</i>	53 ± 2.5	24 ± 0.5	46 ± 1.4	75 ± 0.7	61 ± 1.1
<i>Allamanda cathartica</i>	37 ± 0.8	11 ± 0.4	27 ± 0.2	39 ± 0.9	42 ± 0.5
<i>Cerbera odollam</i>	30 ± 4.1	19 ± 0.7	26 ± 0.5	30 ± 0.6	40 ± 0.9
<i>Calotropis gigantea</i>	28 ± 0.8	14 ± 0.6	44 ± 1.7	42 ± 0.8	33 ± 0.5
RSA (AA mg/g extract) <sup>2</sup>					
<i>Dyera costulata</i>	377 ± 25	15 ± 0.3	9.3 ± 0.5	349 ± 21	278 ± 5.4
<i>Vallis glabra</i>	84 ± 0.5	6.0 ± 0.3	8.4 ± 0.4	77 ± 2.2	119 ± 6.8
<i>Plumeria obtusa</i>	66 ± 3.5	4.9 ± 0.2	10 ± 0.9	58 ± 0.1	115 ± 5.3
<i>Kopsia fruticosa</i>	63 ± 3.2	12 ± 1.4	7.5 ± 0.3	70 ± 2.0	48 ± 1.5
<i>Nerium oleander</i>	42 ± 1.3	6.4 ± 0.1	8.5 ± 0.6	48 ± 1.2	33 ± 0.2
<i>Alstonia angustiloba</i>	29 ± 0.9	10 ± 0.2	5.7 ± 0.7	50 ± 1.2	46 ± 2.2
<i>Catharanthus roseus</i>	24 ± 0.8	13 ± 0.5	21 ± 0.5	33 ± 1.1	31 ± 0.3
<i>Allamanda cathartica</i>	17 ± 1.1	11 ± 0.1	7.9 ± 0.8	18 ± 1.4	24 ± 2.0
<i>Cerbera odollam</i>	15 ± 1.0	16 ± 0.4	5.5 ± 0.1	14 ± 1.3	25 ± 0.7
<i>Calotropis gigantea</i>	7.6 ± 0.4	5.6 ± 0.3	6.1 ± 0.4	8.0 ± 0.6	14 ± 1.3

<sup>1</sup>HEX = Hexane, DCM = Dichloromethane, and MeOH = Methanol; <sup>2</sup>BE = Boldine equivalent, GAE = Gallic acid equivalent, AA = Ascorbic acid, TAC = Total alkaloid content, TPC = Total phenolic content, and RSA = Radical scavenging activity

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## REFERENCES

- Wiert C. Medicinal Plants of Asia and the Pacific. Boca Raton: CRC Press/Taylor and Francis; 2006.
- Endress ME, Bruyns PV. A revised classification of the Apocynaceae. Bot Rev 2000;66:1-56.
- Ng FS. Tropical Horticulture and Gardening. Kuala Lumpur, Malaysia: Clearwater Publications; 2006.
- Rahayu SS. *Allamanda cathartica* L. In: Valkenburg JL, Bunyaphatsara N, editors. Plant Resources of South-East Asia No. 12(2): Medicinal and poisonous plants 2. Leiden, Netherlands: Backhuys Publisher; 2001. p. 51.
- Mulyoutami E, Rismawan R, Joshi L. Local knowledge and management of simpukng (forest gardens) among the Dayak people in East Kalimantan, Indonesia. For Ecol Manage 2009;257:2054-61.
- Lin KW. Ethnobotanical study of medicinal plants used by the Jah Hut people in Malaysia. Indian J Med Sci 2005;59:156-61.
- Rajakaruna N, Harris CS, Towers GH. Antimicrobial activity of plants collected from serpentine outcrops in Sri Lanka. Pharm Biol 2002;40:235-44.
- Nalwaya N, Pokharna G, Deb L, Jain NK. Wound healing activity of latex of *Calotropis gigantea*. Int J Pharm Pharm Sci 2009;1:176-81.
- Sutarno H, Rudjiman SU. *Catharanthus roseus* (L.) G. Don. In: de Padua LS, Bunyaphatsara N, Lemmens RH, editors. Plant Resources of South-East Asia No. 12(1): Medicinal and poisonous plants 1. Leiden, Netherlands: Backhuys Publisher; 1999. p. 185-90.
- Khanh TC. *Cerbera odollam* Gaertner. In: Valkenburg JL, Bunyaphatsara N, editors. Plant Resources of South-East Asia No. 12(2): Medicinal and poisonous plants 2. Leiden, Netherlands: Backhuys Publisher; 2001. p. 154-5.
- Subhadhirasakul S, Jankeaw B, Malinee A. Chemical constituents and antioxidative activity of the extract from *Dyera costulata* leaves. Songkla J Sci Technol 2003;25:35-7.
- Johnson T. CRC Ethnobotany Desk Reference. CRC Press LLC; 1999.
- Burkill IH. A Dictionary of the Economic Products of the Malay Peninsula. Vol 2. (I-Z). London: Crown Agents for the Colonies; 1935.
- Wongpornchai S, Sriseadka T, Choovisase S. Identification and quantitation of the rice aroma compound, 2-acetyl-1-

- pyrroline, in bread flowers (*Vallis glabra* Ktze). *J Agric Food Chem* 2003;51:457-62.
15. Schmidt DF, Yunes RA, Schaab EH, Malheiros A, Filho VC, Franchi GC Jr, *et al.* Evaluation of the anti-proliferative effect of extracts of *Allamanda blanchetti* and *A. schottii* on the growth of leukemic and endothelial cells. *J Pharm Pharm Sci* 2006;9:200-8.
  16. Keawpradub N, Eno-Amooquaye E, Burke PJ, Houghton PJ. Cytotoxic activity of indole alkaloids from *Alstonia macrophylla*. *Planta Med* 1999;65:311-5.
  17. Jagetia GC, Baliga MS. Evaluation of anticancer activity of the alkaloid fraction of *Alstonia scholaris* (Sapthaparna) *in vitro* and *in vivo*. *Phytother Res* 2006;20:103-9.
  18. Lhinhatrakool T, Sutthivaiyakit S. 19-Nor- and 18,20-epoxy-cardenolides from the leaves of *Calotropis gigantea*. *J Nat Prod* 2006;69:1249-51.
  19. Wang ZH, Wang MY, Mei WL, Han Z, Dai HF. A new cytotoxic pregnanone from *Calotropis gigantea*. *Molecules* 2008;13:3033-9.
  20. Seeka C, Sutthivaiyakit S. Cytotoxic cardenolides from the leaves of *Calotropis gigantea*. *Chem Pharm Bull* 2010;58:725-8.
  21. Siddiqui MJ, Ismail, Z, Aisha AF, Abdul Majid AM. Cytotoxic activity of *Catharanthus roseus* (Apocynaceae) crude extracts and pure compounds against human colorectal carcinoma cell line. *Int J Pharmacol* 2010;6:43-7.
  22. Chang LC, Gills JJ, Bhat KP, Luyengi L, Farnsworth NR, Pezzuto JM, *et al.* Activity-guided isolation of constituents of *Cerbera manghas* with anti-proliferative and anti-estrogenic activities. *Bioorg Med Chem Lett* 2000;10:2431-4.
  23. Nurhanan MY, Asiah O, Mohd Ilham MA, Siti Syarifah MM, Norhayati I, Lili Sahira H. Anti-proliferative activities of 32 Malaysian plant species in breast cancer cell lines. *J Trop For Sci* 2008;20:77-81.
  24. Siddiqui BS, Begum S, Siddiqui S, Lichter W. Two cytotoxic pentacyclic triterpenoids from *Nerium oleander*. *Phytochemistry* 1995;39:171-4.
  25. Pathak S, Multani AS, Narayan S, Kumar V, Newman RA. Anvirel™, an extract of *Nerium oleander*, induces cell death in human but not murine cancer cells. *Anticancer Drugs* 2000;11:455-63.
  26. Kardono LS, Tsauri S, Padmawinata K, Pezzuto JM, Kinghorn AD. Cytotoxic constituents of the bark of *Plumeria rubra* collected in Indonesia. *J Nat Prod* 1990;53:1447-55.
  27. Lee CC, Houghton P. Cytotoxicity of plants from Malaysia and Thailand used traditionally to treat cancer. *J Ethnopharmacol* 2005;100:237-43.
  28. Skehan P, Storeng R, Scudiero D, Monks A, McMahon J, Vistica D, *et al.* New colorimetric cytotoxicity assay for anticancer drug screening. *J Natl Cancer Inst* 1990;82:1107-12.
  29. Vichai V, Kirtikara K. Sulforhodamine B colorimetric assay for cytotoxicity screening. *Nat Protoc* 2006;1:1112-6.
  30. Cheah YH, Nordin FJ, Tee TT, Azimahtol HL, Abdullah NR, Ibrahim Z. Antiproliferative property and apoptotic effect of xanthorrhizol on MDA-231 breast cancer cells. *Anticancer Res* 2008;28:3677-90.
  31. Gaidhani SN, Lavekar GS, Juvekar AS, Sen S, Singh A, Kumari S. *In-vitro* anticancer activity of standard extracts used in Ayurveda. *Phcog Mag* 2009;5:425-9.
  32. Ribeiro B, Lopes R, Andrade PB, Seabra RM, Goncalves RF, Baptista P, *et al.* Comparative study of phytochemicals and antioxidant potential of wild edible mushroom caps and stipes. *Food Chem* 2008;110:47-56.
  33. Wong SK, Lim YY, Chan EW. Antioxidant properties of *Hibiscus*: Species variation, altitudinal change, coastal influence and floral colour change. *J Trop For Sci* 2009;21:307-15.
  34. Laphookhieo S, Cheenpracha S, Karalai C, Chantrapromma S, Rat-a-pa Y, Ponglimanont C, *et al.* Cytotoxic cardenolide glycoside from the seeds of *Cerbera odollam*. *Phytochemistry* 2004;65:507-10.
  35. Ali AM, Mackeen MM, Ei-Sharkawy SH, Hamid JA, Ismail NH, Ahmad FB, *et al.* Antiviral and cytotoxic activities of some plants used in Malaysian indigenous medicine. *Pertanika J Trop Agric Sci* 1996;19:129-36.
  36. Lim KH, Hiraku O, Komiyama K, Koyano T, Hayashi M, Kam TS. Biologically active indole alkaloids from *Kopsia arborea*. *J Nat Prod* 2007;70:1302-7.
  37. Subramaniam G, Hiraku O, Hayashi M, Koyano T, Komiyama K, Kam TS. Biologically active aspidofractinine alkaloids from *Kopsia singapurensis*. *J Nat Prod* 2008;71:53-7.
  38. Ahmed F, Sadhu SK, Ohtsuki T, Khatun A, Ishibashi M. Glycosides from *Vallis solanacea* with TRAIL resistance overcoming activity. *Heterocycles* 2010;80:477-88.
  39. Turan N, Akgün-Dar K, Kuruca SE, Kiliçaslan-Ayna T, Seyhan VG, Atasever B, *et al.* Cytotoxic effects of leaf, stem and root extracts of *Nerium oleander* on leukemia cell lines and role of the p-glycoprotein in this effect. *J Exp Ther Oncol* 2006;6:31-8.
  40. Kam TS, Lim KH. Alkaloids of *Kopsia*. In: Cordell GA, editor. *The Alkaloids: Chemistry and Biology*. Amsterdam, Netherlands: Elsevier Inc; 2008. p. 1-111.
  41. Akinmoladun AC, Obuotor EM, Farombi EO. Evaluation of antioxidant and free radical scavenging capacities of some Nigerian indigenous medicinal plants. *J Med Food* 2010;13:444-51.
  42. Guo DJ, Cheng HL, Chan SW, Yu PH. Antioxidative activities and the total phenolic contents of tonic Chinese medicinal herbs. *Inflammopharmacology* 2008;16:201-7.
  43. Raj JX, Bajjapai PK, Kumar PG, Murugan PM, Kumar J, Chaurasia OP, *et al.* Determination of total phenols, free radical scavenging and antibacterial activities of *Mentha longifolia* Linn. Hudson from the Cold Desert, Ladakh, India. *Phcog J* 2010;2:470-5.
  44. Dudonné S, Vitrac X, Coutière P, Woillez M, Mérillon JM. Comparative study of antioxidant properties and total phenolic content of 30 plant extracts of industrial interest using DPPH, ABTS, FRAP, SOD, and ORAC assays. *J Agric Food Chem* 2009;57:1768-74.
  45. Lamien-Meda A, Lamien CE, Compaoré MM, Meda RN, Kiendrebeogo M, Zeba B, *et al.* Polyphenol content and antioxidant activity of fourteen wild edible fruits from Burkina Faso. *Molecules* 2008;13:581-94.
  46. Cheung LM, Cheung PC, Ooi VE. Antioxidant activity and total phenolics of edible mushroom extracts. *Food Chem* 2003;8: 249-55.

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