The Ocean's Treasure Trove: Bioactive Compounds from Sea Sponges

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ABSTRACT

Sea sponges are among the oldest living creatures on Earth that have evolved complex chemical defenses to thrive in the ocean's competitive environment. These marine animals have developed an impressive array of bioactive compounds with therapeutic applications. This article explores bioactive compounds derived from sea sponges, showcasing their remarkable therapeutic potential. The article begins by introducing the fascinating world of sea sponges, highlighting their unique biology and ecology. We then delve into the various bioactive compounds isolated from sea sponges. The review highlights the anticancer properties of sea sponge-derived compounds demonstrating their potential in cancer treatment. We also explore their antimicrobial and anti-inflammatory activities, showcasing their potential in combating infectious diseases and inflammatory disorders. The article further discusses the challenges and opportunities in harnessing the bioactive potential of sea sponges. This review serves as a comprehensive resource for researchers, scientists, and students interested in marine biomedicine and natural product drug discovery.

Keywords: Sea sponges, Bioactive compounds, Therapeutic potential, Anticancer, Anti-inflammatory, Marine biomedicine, Drug discovery.

INTRODUCTION

The Earth's surface is predominantly covered by oceans, which encompass a vast array of ecosystems that support an incredible 87% of the planet's biodiversity.^[1] Research on marine-based substances has led to the discovery of many biologically active compounds with possible medical uses.^[2] Scientists have discovered around 30,000 bioactive substances from the ocean. The ocean's unique ecosystem is a rich source of diverse bioactive compounds, with numerous examples already demonstrating therapeutic potential across a range of diseases.^[3] The marine-derived compounds are being explored for their potential to create functional foods and dietary supplements. Marine-derived natural products have demonstrated impressive therapeutic potential, exhibiting anti-inflammatory, antimicrobial, and anticancer activities, among others.^[4] Marine microorganisms are source of novel pharmaceuticals, aligning with traditional drug discovery models that rely on natural products. Additionally, the scalability of microbial fermentation processes offers a viable solution for meeting the demand for large-scale production of



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bioactive compounds derived from marine microorganisms.^[4] Recent advances in marine biotechnology have revealed that microorganisms from marine environments are a rich source of novel natural products.^[1] Microbiomes associated with deep-sea sponges display a distinctive profile, characterized by lower complexity and higher diversity compared to their shallow-water counterparts.^[5] Sea sponges are a treasure trove of bioactive compounds, offering promising leads for therapeutic applications. For instance, certain species like *Axinella donnani* and *Clathria compressa* have been found to harbor antibacterial compounds. Others, such as *Aaptos aaptos* and *Dactylospongia metachromia*, contain antiviral compounds.^[6,7]

OVERVIEW OF DIFFERENT TYPES OF SEA SPONGE AND BIOACTIVE COMPOUND PRESENT IN THEM

Aplysina aerophoba

The Mediterranean sponge *Aplysina aerophoba* thrives in sunlit environments and attaches itself to stable surfaces like rocks. Its vibrant yellow hue is attributed to the unstable pigment uranidine, which transforms when exposed to air. Interestingly, this sponge harbors a vast array of bacteria and contains high levels of brominated alkaloids, particularly isoxazoline alkaloids. These compounds can be broken down into smaller

3,5 dibromotyrosin structures, which exhibit antibacterial properties. Research has shown that the isolated brominated metabolites from Aplysina aerophoba display moderate to potent cytostatic and antimicrobial activities.^[8] Researchers examined the osteogenic potential of a crude acetonic extract derived from the Mediterranean sponge Aplysina aerophoba, with a particular emphasis on its prominent bromotyrosine-derived compounds. The research utilized zebrafish larvae as a model to assess mineralization and bone development. The findings identified aerophobin-1 as a promising pro-osteogenic agent, presenting potential applications in regenerative medicine.^[9] A thorough examination of the sponge's chemical composition led to the identification of four prominent brominated alkaloids, which were characterized as distinct compounds.^[10] It yielded a unique Gram-positive bacterium, which was classified within the Rubrobacter genus following an in-depth examination of its genetic characteristics.^[11] Aeroplysinin-1, a natural compound derived from marine sources, has shown exceptional anti-inflammatory and anti-angiogenic effects. Moreover, Aeroplysinin-1 has demonstrated broad-spectrum antimicrobial activity, inhibiting the growth of diverse microorganisms, including fungi, protozoa, and viruses. Additionally, research has indicated that this compound can suppress tumor growth or induce apoptosis in various cell lines, including those of endothelial, monocytic, and cancerous origin.^[12] Aeroplysinin-1 anti-inflammatory properties by exhibits modulating endothelial cell responses. Its mechanism involves inhibiting key signaling molecules, ultimately hindering the activation of pro-inflammatory pathways. This discovery opens possibilities for Aeroplysinin-1's potential use in managing cardiovascular diseases. Additionally, Aeroplysinin-1 has been found to exhibit anti-angiogenic properties.^[13,14] A team of scientists discovered two new compounds, Microsphaerones A and B, in a fungus that lives inside Aplysina aerophoba. The discovery of Microsphaerones A and B is significant, as they represent the first gamma-pyrone derivatives isolated from this fungal genus.^[15] Researchers discovered beneficial bacteria in Aplysina aerophoba, that produce antimicrobial compounds.^[16] A comprehensive study examined how controlled growth conditions influence the bacterial communities associated with Aplysina aerophoba sponges. Over six months, sponges were cultivated under varying temperatures, light exposure, and nutrient levels. Using denaturing gradient gel electrophoresis and sequencing, researchers identified a diverse bacterial community spanning five phyla. Despite reduced biomass and compromised morphology, the sponges maintained distinct microbial associations. Notably, a Cyanobacteria strain exhibited adaptive responses to changing light conditions. Moreover, the cultivation period saw an increase in characteristic brominated compounds.[17]

Axinella corrugate

Axinella corrugate species are typically found in exposed deep reef environments. They are native to the Indian and Pacific Oceans. Characteristically, these sponges are relatively small, usually under 20 cm in size, and display vibrant yellow or orange hues. Researchers isolated novel compounds from the marine sponge *Axinella corrugata*, including ester derivatives of a coumarin-based carboxylic acid. Notably, these compounds demonstrated potent inhibitory activity against the SARS-coronavirus.^[18,19] A symbiotic fungus associated with it has been found to produce a compound with antibacterial properties.^[20] It contains two unique proteins with distinct abilities to bind to red blood cells. One protein strongly binds to cells from various animals, but this binding is blocked by specific sugars. The other protein targets rabbit red blood cells and is inhibited by certain sugars and compounds.^[21]

Hyrtios erectus

Hyrtios erectus, a marine sponge, is classified within the phylum Porifera. Specifically, it falls under the class Demospongiae, order Dictyoceratida, and family Thorectidae. This sponge is characterized by its distinctive blackish hue.^[22,23] Researchers examined the impact of a Hyrtios erectus marine sponge extract on the proliferation and survival of MCF-7 breast cancer cells.^[22] Scientists extracted a total of 20 sesterterpenes, 12 of which were novel, from Hyrtios erectus found near Chuuk Island. Tests showed that some of these compounds slowed the growth of various cancer cells.^[24] Scientists discovered two new scalarane-type sesterterpenes, Hyrtioscalaranes A and B. These compounds showed promising anti-inflammatory and antioxidant properties. After purifying the extract, researchers found that Hyrtioscalaranes A and B targeted a specific enzyme involved in inflammation more selectively than a commonly used anti-inflammatory drug.^[25] Scientists analyzed extracts from the marine sponge Hyrtios erectus and identified two notable compounds. One was a previously discovered oxysterol, while the other was a new alkyl benzoate derivative. The two key compounds exhibited significant toxicity against human breast cancer cells.^[26] Researchers investigated compounds from Hyrtios erectus for their ability to combat malaria, specifically targeting a strain of *Plasmodium falciparum* that is resistant to chloroquine. Following extraction with 85% methanol, three compounds smenotronic acid, ilimaquinone, and pelorol - were isolated and structurally elucidated using advanced spectroscopic methods. In vitro evaluations of these compounds against P. falciparum revealed promising results, with pelorol demonstrating notable antimalarial activity.^[27] Scientists investigated the chemical composition of Hyrtios erectus, uncovering a range of distinctive compounds, including 5-hydroxy-1H-indole-3-carboxylic acid methyl ester, hyrtiosulawesine, and the hyrtiosin A and B analogs.^[28] Research has shown that hyrtiosal, a compound extracted from the marine sponge Hyrtios erectus, possesses anti-HIV properties by blocking the interaction between

HIV-1 integrase and viral DNA.^[29] Researchers discovered that a compound extracted from *Hyrtios erectus* effectively blocks a key enzyme involved in insulin signaling. Therefore, is a potential target for treating obesity and type 2 diabetes.^[30]

Spongia officinalis

Spongia officinalis, is a widespread species found in the Mediterranean Sea, northeastern Atlantic Ocean, and beyond. It is a member of the Dictyoceratida order.^[31] Bioactive terpenes were extracted from Spongia officinalis, revealing a diverse range of compounds. The extract contained sesterterpenes and C₂₁ furanoterpenes, including four newly discovered oxidized metabolites. Additionally, six known furospongin-series compounds and three scalarane sesterterpenes were isolated. Researchers made a significant discovery, finding that two key compounds, tetrahydrofurospongin-2 and dihydrofurospongin -2, triggered the formation of biofilms in Escherichia coli. Additionally, the isolated compounds were tested for their ability to combat bacterial and fungal growth.^[32] Researchers isolated a series of bioactive diterpenes like 3-Nor-spongiolide A, Spongiolides A, and Spongiolides B, whose chemical frameworks were elucidated using cutting-edge spectroscopic methods.^[33] Scientists have extracted a variety of bioactive substances from a marine sponge, Spongia officinalis, which was cultivated through aquaculture. The extracted compounds included several known substances, such as Jellynolide A and sponalisolides, as well as new metabolites. Jellynolide A, was obtained as a colorless oil.^[34] Researchers discovered two new compounds in a marine sponge species. The extracted compounds included a unique acetoxy diterpenoid and an unusual 18-nor-spongian derivative. Additionally, six known metabolites were identified.^[35] Scientists isolated anti-inflammatory steroids from Spongia officinalis, including a novel, highly degraded steroid featuring a unique 5/6/5-tricyclic structure.^[36] Researchers examined a unique compound produced by a Bacillus bacterium living in association with the Spongia officinalis. The compound, (3S, 6S)-3,6-diisobutylpiperazine-2,5-dione, was analyzed using advanced spectroscopic techniques. Tests showed that the compound had antibacterial properties.^[37] Researchers evaluated the anticonvulsant and pain-relieving properties of extracts and fractions derived from Spongia officinalis. The results showed promising anticonvulsant and analgesic effects, indicating potential uses in therapy.^[38] Researchers discovered anticancer compounds in a fungus called Penicillium sp., which lives inside a marine sponge. The extracted compounds demonstrated impressive anticancer activity.^[39] Researchers investigated the potential of Spongia officinalis, to yield anti-inflammatory and anticancer compounds. They tested the sponge's methanol extract and fractions in rats with induced inflammation and in human cancer cells grown in the lab. The results showed that the extract and fractions had significant anti-inflammatory and anticancer effects, with the effects increasing as the dose increased.^[40]

Petrosia ficiformis

Petrosia ficiformis, a massive, wine-colored sponge is widely distributed throughout the Mediterranean Sea and Macaronesia, including the Azores, Madeira, and Canary Islands.^[41] A recent study yielded five newly discovered polyacetylenes from Petrosia ficiformis.[42] Scientists identified 12 unusual stanols with a distinctive 5 beta-dihydro nucleus in the marine sponge Petrosia ficiformis. Interestingly, these compounds were absent in earlier samples of the same sponge, leading researchers to speculate that they might be formed through bacterial processing of the sponge's natural sterols.^[43] Scientists discovered a new sterol, (24R)-24,26-Dimethylcholesta-5,26-dien-3 beta-ol, in the marine sponge Petrosia ficiformis. A thorough examination revealed a diverse range of monohydroxy sterols, including common and rare types. Using advanced spectroscopy and comparison with a synthetic version, researchers confirmed the structure of a newly identified minor sterol, (24R)-24,26-dimethylcholesta-5,26-dien-3β-ol, also known as 26(29)-dehydroaplysterol.^[44] Researchers investigated the antimicrobial properties of bacteria living on the marine sponge Petrosia ficiformis. The study aimed to explore the potential link between the sponge's natural products and those produced by its associated microorganisms. Using a combination of traditional and molecular techniques, the researchers identified 57 bacterial strains with distinct characteristics.[45]

Niphates erecta

The Lavender rope sponge, scientifically known as Niphates erecta, is a highly variable species. The surface texture varies from smooth and porous to spiky. The color of Niphates erecta ranges from pale purple, pink, bluish, or grey on the exterior to a lighter shade internally. The sponge is compressible, tough, yet easy to cut. The marine sponge Niphates erecta has a widespread distribution, occurring in areas of Bermuda. Scientists discovered a novel glycoprotein, niphatevirin, from the marine sponge Niphates erecta, demonstrating potent anti-HIV properties. Niphatevirin exhibited significant anti-HIV activity, protecting human lymphoblastoid cells from HIV-1-induced damage.^[46] An exploration of marine organisms in the Colombian Caribbean Sea uncovered promising antiherpes properties. Specifically, three species of marine sponges - Aka cachacrouense, Niphates erecta, and Dragmacidon reticulatum - showed notable potential, justifying further research to isolate and identify the bioactive compounds driving this activity.^[47]

Halichondria panicea

Halichondria panicea exhibits highly variable morphology. Typically found on open coastlines, it can form a low-lying crust with distinctive "volcano-like" exhalant openings. It typically exhibits a cream-yellow hue. A distinctive seaweed-like odor is also characteristic of *Halichondria panicea*. Geographically, this species is distributed across the North Atlantic region, ranging

from the Barents Sea to the Mediterranean.^[48] Scientists found that certain types of sea sponges are home to bacteria that produce antibiotics. These antibiotics can effectively combat strains of bacteria that are resistant to multiple drugs. The discovery of these bacteria and their antibacterial compounds could lead to the development of new medicines.^[49] Scientists have identified new complex lipids in a type of marine sponge. One of these lipids has a unique fatty acid component and was found to have a specific structure. Researchers used advanced techniques to determine the lipid's composition. Additionally, a second complex lipid was discovered in the same sponge species, notable for its unusually long fatty acid chain.^[50] Researchers discovered four novel gamma-pyrones, designated as nocapyrones A-D, in an organic extract of the Nocardiopsis strain HB383.^[51] Scientists discovered a new compound mayamycin, derived from a type of bacteria Streptomyces sp., that shows promise in fighting cancer and antibiotic-resistant bacteria.[52]

Cliona celata

Studies on the ethyl acetate fraction of Cliona celata revealed potential anti-inflammatory properties. Mechanistic its investigations showed that this fraction impeded the nuclear migration of NF-KB p65 subunits, accompanied by decreased phosphorylation and degradation of IkB-a. Additionally, ECC strongly suppressed NF-kB-mediated gene expression and DNA-binding activity, accompanied by decreased nuclear p65 protein levels. Mechanistic investigations revealed that ECC inhibits NF-κB activation by preventing IκB degradation, thereby modulating the inflammatory response.^[53] Researchers assessed the efficacy of Cliona celata marine sponge extracts as larvicides, ovicides, and repellents against Anopheles stephensi, a key malaria vector. The study examined the effectiveness of various solvent extracts of C. celata in controlling Anopheles stephensi populations. The findings revealed that the methanol extract demonstrated the highest larvicidal activity, significantly impacting fourth-instar larvae at a concentration of 500 ppm. Conversely, the hexane extract proved to be the most potent repellent against Anopheles stephensi.^[54] Researchers investigated the multifaceted inhibitory effects of Cliona celata on TNF-amediated MMP-9 production, exploring its influence on NF-κB and AP-1 signaling pathways, enzymatic activity, and cellular migration. MMP-9 plays a pivotal role in extracellular matrix degradation, contributing to the pathological proliferation and migration of vascular smooth muscle cells in atherosclerotic lesions.^[55] A novel aminosteroid, Clionamine B, derived from the marine sponge *Cliona celata*, has been shown to induce autophagy. Researchers developed an efficient synthesis of Clionamine B, utilizing tigogenin, a plant-derived sapogenin, as the starting material. A key step in the synthesis involved the stereoselective introduction of the α -hydroxyl group at C-20, accomplished through the oxidation of a y-lactone enolate with molecular oxygen. The resulting synthetic Clionamine B exhibited robust

autophagy-inducing activity in MCF-7 human breast cancer cells.^[56] Scientists found that *Cliona celata*, possesses notable antioxidant and anti-inflammatory activities.^[57]

Corticium candelabrum

Corticium candelabrum, a Homosclerophorida-order sponge, is native to the eastern Atlantic Ocean and Mediterranean Sea. Typical habitats include coralline algae communities in shady positions, such as vertical walls, under overhangs, and in caves, at depths of up to 20 meters.^[58] *Corticium candelabrum* typically inhabits rocky areas beneath the littoral zone in the Mediterranean region. *Corticium candelabrum* is a prolific source of bioactive compounds, featuring a diverse array of terpenoids, alkaloids, and peptides. The therapeutic potential of these secondary metabolites has sparked significant interest in biomedical research. Investigations have revealed that compounds isolated from *Corticium candelabrum* exhibit a broad spectrum of biological activities, including antimicrobial, antiviral, and anticancer properties.^[59]

Crella spinulata

Crella spinulata is a demosponge species within the Crella genus and Crellidae family. Notably, research has demonstrated that Crella spinulata exhibits anti-proliferative effects and induces cell cycle arrest in colon cancer cells under in vitro conditions. A recent study investigated the anticancer properties of Crella spinulata's mesohyl against the Caco-2 colon cancer cell line. Using MTT assays and flow cytometry, the results showed that Crella spinulata significantly inhibited Caco-2 cell proliferation.^[60] Crella spinulata has shown significant antimicrobial and cytotoxic properties.^[61] Studies have found that extracts and mesohyls from marine sponges, including Crella spinulata, can inhibit the growth and migration of liver cancer cells. A novel 3D cell culture system, known as primmorph, provides a sustainable method for producing bioactive compounds from marine sponges for potential use in cancer therapies. The anticancer effects of primmorph extracts and mesohyls from Crella spinulata were assessed through various assays, including cell viability, colony formation, cell cycle analysis, and apoptosis. Treatment with these extracts significantly reduced the migration and proliferation of liver cancer cells (HepG2). Notably, the primmorph extract of Crella spinulata exhibited the most promising anticancer activity, demonstrating both antiproliferative and antimigratory effects.^[62] Researchers have discovered a novel class of compounds, designated as shishicrellastatins, derived from the marine sponge Crella spinulata.^[63]

Cribrochalina vasculum

Cribrochalina vasculum, a marine sponge belonging to the Niphatidae family and Haplosclerida order, exhibits a unique morphology. Its shape is variable, often resembling a funnel or bowl, but can also be irregular.^[64] Research has identified

compounds from the marine sponge Cribrochalina vasculum as potential anticancer agents. Notably, two acetylene alcohols derived from the sponge disrupted tumor cell signaling mediated by IGF-1R, offering a promising avenue for cancer treatment.^[65] Scientists have identified a range of bioactive compounds in the Caribbean sponge Cribrochalina vasculum, including a distinct class of hydroxyalkynyl lipids with significant biological activity. Detailed analysis revealed three prominent compounds: 3-hydroxydocosa-(4E, 15E)-dien-1-yne, 3-hydroxy-16-methyleicos-(4E)-en-1yne, and 3-hydroxy-19-methyleicos-(4E)-en-1-yne. Further examination of the sponge's lipophilic extract led to the discovery of four novel acetylene metabolites, each displaying unique structural characteristics.^[66] A novel compound, 23-methyl-5,9-pentacosadienoic acid, was isolated and characterized from the marine sponge Cribelochalina vasculum through a comprehensive extraction and analysis process.^[67] Scientists have detected a small quantity of the sterol 23-Epidihydrocalysterol in the marine sponge Cribrochalina vasculum. A novel sterol featuring a cyclopropane ring was isolated from C. vasculum and characterized using advanced analytical techniques.^[68]

Fasciospongia cavernosa

Fasciospongia cavernosa, a species of sea sponge, is distinguished by its robust, fleshy walls and expansive, cavernous interior. Its unique appearance is marked by prominent, longitudinal ridges, or fascioles, that run along its surface. The marine sponge Fasciospongia cavernosa inhabits tropical waters, typically in shallow coral reefs and rocky crevices, and is widespread across the Indo-Pacific region, including nations such as Australia, Indonesia, and the Philippines. Researchers have isolated a novel cacospongionolide derivative from this sponge, designated as cacospongionolide F. Spectroscopic analysis and chemical transformations revealed its unique structure, identifying it as a bioactive sesterterpene. The absolute configuration of cacospongionolide F was elucidated employing a variant of the Mosher esterification approach. The antimicrobial properties and toxicity of cacospongionolide F to brine shrimp and fish have been investigated.^[69] Scientists have isolated a unique lectin from the marine sponge Fasciospongia cavernosa, which exhibits specificity for D-galactose and N-acetyl-D-galactosamine.^[70] Scientists have discovered a novel compound, Cacospongionolide E, in Fasciospongia cavernosa. This compound demonstrated potent inhibition of human secretory phospholipase A2 (PLA2), surpassing the efficacy of the well-established compound manoalide. Notably, Cacospongionolide E exhibited no toxicity towards human neutrophils. Biological assessments revealed pronounced activity in the Artemia salina bioassay and moderate toxicity in the Gambusia affinis fish lethality test.^[71] Scientists have discovered a new sesterterpene, cacospongionolide B, in the marine sponge Fasciospongia cavernosa, native to

the Adriatic region.^[91] Scientists have discovered that two sesterterpenoids, cacospongionolide and scalaradial, derived from marine sources, exhibit potent pro-apoptotic effects, inducing programmed cell death in human carcinoma cell lines.^[72] Furthermore, Cacospongionolide B, a sesterterpene has shown strong anti-inflammatory properties.^[73] Scientists have identified Cacospongionolide B, a distinctive compound found in the *Fasciospongia cavernosa*, as a potent suppressor of human synovial phospholipase A2 (sPLA2) activity. This suppression has been demonstrated to modulate inflammatory responses over both acute and chronic timeframes.^[74] Researchers investigated the anti-inflammatory properties of cavernolide, a unique C₂₁ terpene lactone isolated from the marine sponge *Fasciospongia cavernosa*.^[75]

Halichodrian bowerbanki

Halichondria bowerbankii, a marine species known by several common names, thrives in bays and harbors. Originating in the North Atlantic, its native range extends from Iceland and Norway to the Mediterranean, Azores, and eastern North America.^[76] The marine sponge *Halichondria bowerbanki* has been found to contain a bioactive compound known as Halichondamide A. Research has shown that Halichondamide A exhibits notable anticancer properties, with moderate inhibitory effects observed in certain cancer cell lines, particularly those linked to hepatic and mammary tumors.^[77]

Hyattella intestinalis

Hyattella intestinalis, a demosponge, boasts a distinctive body composition featuring pebble needles and spongine fibers, allowing for substantial water absorption. This species is classified within the genus Hyattella and family Spongiidae. Researchers have isolated spongian diterpenes from Hyattella intestinalis, revealing moderate antiviral activity against adenovirus. Advanced chromatographic and spectroscopic techniques facilitated the identification of three previously unknown compounds. Notably, one of these compounds exhibited significant antiviral properties.^[78] Scientists have identified a distinct group of norsesterterpenes, termed mooloolabenes A-E, in the Australian marine sponge Hyattella intestinalis. Furthermore, a previously unknown sesterterpene, mooloolaldehyde, displaying structural similarities to scalarane compounds, was also isolated from this species. Biological evaluation of the extracted compounds revealed significant anticancer potential, characterized by pronounced cytotoxic effects against the P388 cancer cell line.^[79] Scientists have found that certain bacteria living in the marine sponge Hyattella intestinalis show strong potential in combating malaria. Laboratory tests revealed that these bacteria effectively inhibit the growth of Plasmodium falciparum, the parasite responsible for the disease.[80]

lotrochota baculifera

Iotrochota baculifera, a demosponge species, boasts a distinctive skeletal structure composed of elongated, slender spicules. Characterized by its porous body and intricate network of canals and chambers, this sea sponge belongs to the Iotrochotidae family. Chemical analysis of specimens collected from the Indian Ocean revealed a range of sphingolipids, including the novel glycosphingolipid iotroridoside-B and a mixture of four sphingolipid components, featuring two newly discovered compounds.^[81] A chemical study of the marine sponge *Iotrochota* birotulata, sampled from the waters off Port Royal, Jamaica, aimed to identify its primary constituents and assess the bioactivity of its crude extracts. The analysis revealed a range of compounds, notably renierapurpurin, a carotenoid derivative previously unreported in this species. Additional isolated compounds included a tyrosine derivative and the ubiquitous steroid β-sitosterol.^[82]

Ircinia strobilina

Ircinia strobilina, a marine sponge belonging to the Irciniidae family, is recognizable by its grey or glossy black hue and unique conular spiny surface features. This species is predominantly found in the Caribbean Sea, with documented occurrences near Florida, the Virgin Islands, Cuba, and Venezuela. Researchers have isolated and characterized a lectin from *Ircinia strobilina*, discovering its potential to combat biofilm formation. Specifically, this lectin demonstrated significant inhibitory effects on biofilm formation in certain bacterial strains, including *Staphylococcus aureus* and *Staphylococcus epidermidis*.^[83] The sponge's fatty acid composition was characterized by elevated levels of demospongic acids, a distinct group of fatty acids typical of certain sponge species.^[84]

Jaspis johnstoni

Jaspis johnstoni, a sea sponge belonging to the Ancorinidae family, exhibits a distinctive astrophorid morphology. Characterized by its robust, globular body and siliceous spicule-based skeleton, this species thrives in tropical and subtropical waters, including coral reef environments and rocky crevices. A chemical examination of Fijian *Jaspis johnstoni* specimens revealed two cytotoxic compounds, toyocamycin and 5-(methoxycarbonyl)tubercidin, which are part of the pyrrolo[2,3-d]pyrimidine nucleoside family.^[85] Jasplakinolide possesses antifungal and antiproliferative properties.^[86]

Lendenfeldia chondrodes

Lendenfeldia chondrodes, a vibrant blue sea sponge, is a popular aquarium species. Belonging to the Thorectidae family, this sponge boasts a distinctive appearance.^[87] Scientists have identified polybrominated diphenyl ethers (PBDEs) in the marine sponge *Lendenfeldia chondrodes*. These compounds have shown potential therapeutic value due to their ability to inhibit

specific protein kinases, CDK7 and FynB, which play critical roles in cellular processes.^[88] Researchers have identified novel compounds in *Lendenfeldia chondrodes*. Two previously unknown 1-deoxynojirimycin derivatives were discovered through analysis of the sponge's aqueous extract.^[89] Researchers have identified a novel epidioxy sterol with antifouling properties in the marine sponge *Lendenfeldia chondrodes*.^[90]

Luffariella variabilis

Scientists have identified a range of cytotoxic manoalide-type sesterterpenes in Luffariella variabilis. An in-depth examination of the sponge's chemical constituents led to the isolation of 13 previously unknown linear terpenes, featuring 11 distinctive manoalide derivatives characterized by their acyclic frameworks. Additionally, a polyprenylphenol derivative and a polyprenylbenzaldehyde derivative were identified.^[91] Researchers identified two novel furanosesterterpenoids in the marine sponge Luffariella variabilis. One of these compounds exhibited considerable cytotoxic effects against NBT-T2 cells, suggesting its potential as a valuable candidate for future studies.^[92] An investigation into the chemical composition of Luffariella variabilis led to the discovery of acetylated sesterterpenes.^[93] Scientists discovered a novel manoalide-related sesterterpene, (4E,6E) -dehydro-25-O-methylmanoalide, in extracts of the marine sponge Luffariella variabilis. This finding was complemented by the identification of the established compound (4E,6E)dehydromanoalide.^[94] A subsequent examination of a variant of this sponge species resulted in the isolation of an additional new manoalide-related sesterterpene, 24-O-ethylmanoalide.^[95] Researchers have discovered two novel β-carboline alkaloids, variabines A and B, in tLuffariella variabilis. Spectroscopic analysis revealed the structural composition of these compounds, with variabine A featuring a sulfonated group distinct from variabine B. Notably, variabine A's sulfate group is a unique characteristic among previously identified β-carboline alkaloids. Assessment of variabine B's biological activity revealed potent inhibition of proteasome function and disruption of a key protein-protein interaction. Conversely, variabine A exhibited negligible impact on these biological pathways.^[96] The marine sponge Luffariella variabilis yields a compound called manoalide, which boasts a dual pharmacological profile. Notably, manoalide has been found to block calcium channels and exhibit antimicrobial properties.^[97] Researchers have discovered a new class of anti-inflammatory sesterterpenes, known as luffariellins, which provide significant contributions to the field of chemotaxonomy.[98]

Pachychalina alcaloidifera

The tropical sea sponge *Pachychalina alcaloidifera*, a member of the Niphatidae family, is native to the Indo-Pacific region. It typically thrives in shallow coral reef environments within this geographic range. Investigations into ingenamine G, a marine alkaloid isolated from the *Pachychalina alcaloidifera*, uncovered its dual capacity for inducing cytotoxic and genotoxic effects. Notably, ingenamine G displayed moderate toxicity towards human lymphocytes. Additional analyses suggested that this compound may also possess genotoxic potential, leading to chromosomal instability and genetic alterations. Human lymphocytes were exposed to varying concentrations of ingenamine G, and the results indicated that all concentrations tested exhibited cytotoxicity, reduced mitotic activity, and induced chromosomal damage. DNA strand breaks increased at higher concentrations, and high concentrations of ingenamine G disrupted mitotic spindle formation, inducing chromosomal instability.^[99] A thorough examination of the sponge's chemical constituents revealed a subset of four bis-piperidine alkaloids, including madangamine F, haliclonacyclamine F, arenosclerin D, and arenosclerin E. Employing spectroscopic techniques enabled the characterization of these compounds, which exhibited pronounced cytotoxic effects against a range of cancer cell lines.^[100]

Pandaros acanthifolium

Pandaros acanthifolium belongs to Pandaros genus, part of the Microcionidae family. A detailed chemical examination of this species uncovered a diverse array of steroidal glycosides, including four novel compounds, designated as acanthifoliosides G-J. Characteristic structural features of these compounds include a highly oxygenated D ring and unique sugar moieties. Interestingly, acanthifolioside G demonstrated remarkable antioxidant and cytoprotective activities.^[101] Researchers have identified novel antiprotozoal compounds in the Pandaros acanthifolium sponge. A thorough chemical analysis yielded 12 new steroidal glycosides with potential therapeutic benefits. Laboratory tests revealed that several of these compounds demonstrated significant efficacy against various parasitic protozoa. Notably, two compounds showed potent growth-inhibiting effects against specific parasites.^[102] A comprehensive chemical examination of Pandaros acanthifolium led to the identification of seven previously unknown steroidal glycosides. This investigation uncovered a subset of four novel compounds, termed pandarosides A-D, alongside three methyl ester analogs corresponding to pandarosides A, C, and D.^[103] A detailed re-examination of the chemical constituents of the Caribbean sponge Pandaros acanthifolium uncovered six novels steroidal saponins. Spectroscopic analysis and comparative studies enabled the identification of these compounds, comprising Pandarosides K-M and their corresponding methyl esters. Biological evaluations revealed that these new compounds exhibited moderate to weak antiprotozoal activity. Additionally, their cytotoxic properties against human cancer cell lines, as well as their capacity to induce hemolysis and liposome permeabilization, were assessed in conjunction with previously characterized pandarosides and acanthifoliosides. The findings

indicated that select pandarosides displayed notable cytotoxic effects, whereas certain acanthifoliosides exhibited pronounced hemolytic activity.^[104]

CONCLUSION

The ocean's depths hold a treasure trove of bioactive compounds, with sea sponges emerging as a rich source of novel and diverse molecules. These compounds have shown immense potential in addressing various human health challenges, including cancer, inflammation, and infectious diseases. The unique structural and functional properties of these compounds make them attractive candidates for pharmaceutical development. As research continues to unravel the secrets of sea sponge-derived bioactives, it is essential to adopt sustainable and eco-friendly approaches to harness these treasures. By doing so, we can unlock the full potential of the ocean's treasure trove, leading to the discovery of new life-saving medicines and a deeper appreciation for the importance of marine conservation.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

NF-κB: Nuclear factor kappa B; IκB: inhibitor of nuclear factor kappa B; MMP-9: Matrix metalloproteinase-9; MTT: 3-(4,5-Dimethylthiazol-2-yl)-2,5-Diphenyltetrazolium Bromide; NSCLC: Non-small-cell lung cancer; SCLC: Small-cell lung cancer; PARP: Poly (ADP-ribose) polymerase; IL-1β: Interleukin-1 beta; IL-6: Interleukin 6; IL-8: Interleukin 8; PPAR-α: Peroxisome proliferator-activated receptor alpha; PPAR-γ: Peroxisome proliferator-activated receptor gamma.

SUMMARY

Sea sponges are a rich source of bioactive compounds with potential therapeutic benefits. This review provides an overview of the various bioactive compounds isolated from sea sponges, including their structural diversity and pharmacological properties. The potential applications of these compounds in treating diseases are discussed, along with the challenges and opportunities in developing sea sponge-derived therapeutics. The importance of sustainable approaches in harnessing the medicinal potential of sea sponges is also highlighted.

REFERENCES

- Zhou Q, Hotta K, Deng Y, Yuan R, Quan S, Chen X. Advances in biosynthesis of natural products from marine microorganisms. Microorganisms. 2021;9(12):2551. doi: 10.33 90/microorganisms9122551, PMID 34946152.
- Little RD, Nishiguchi GA. Synthetic efforts toward, and biological activity of, thyrsiferol and structurally related analogues. Stud Nat Prod Chem. 2008;35:3-56. doi: 10.1016 /S1572-5995(08)80003-8.
- Banday AH, ul AN, Farooq R, Sheikh SA, Ganie MA, Parray MN, *et al.* Exploring the potential of marine natural products in drug development: A comprehensive review. Phytochem Lett. 2024;59:124-35. doi: 10.1016/j.phytol.2024.01.001.
- Avhad AB, Bhangale CJ. Marine natural products and derivatives. RPS Pharm Pharmacol Rep. 2023;2(2):rqad008. doi: 10.1093/rpsppr/rqad008.
- Busch K, Slaby BM, Bach W, Boetius A, Clefsen I, Colaço A, et al. Biodiversity, environmental drivers, and sustainability of the global deep-sea sponge microbiome. Nat Commun. 2022;13(1):5160. doi: 10.1038/s41467-022-32684-4, PMID 36056000.
- Varijakzhan D, Loh JY, Yap WS, Yusoff K, Seboussi R, Lim SE, *et al.* Bioactive compounds from marine sponges: fundamentals and applications. Mar Drugs. 2021;19(5):246. doi: 10.3390/md19050246, PMID 33925365.
- Varijakzhan D, Loh JY, Yap WS, Yusoff K, Seboussi R, Lim SE, *et al.* Bioactive compounds from marine sponges: fundamentals and applications. Mar Drugs. 2021;19(5):246. doi: 10.3390/md19050246, PMID 33925365.
- Orfanoudaki M, Hartmann A, Alilou M, Mehic N, Kwiatkowski M, Jöhrer K, et al. Cytotoxic compounds of two demosponges (*Aplysina aerophoba* and *Spongia* sp.) from the Aegean Sea. Biomolecules. 2021;11(5):723. doi: 10.3390/biom11050723, PMID 34065941.
- 9. Carnovali M, Ciavatta ML, Mollo E, Roussis V, Banfi G, Carbone M, *et al*. Aerophobin-1 from the marine sponge *Aplysina aerophoba* modulates osteogenesis in zebrafish larvae. Mar Drugs. 2022;20(2):135. doi: 10.3390/md20020135, PMID 35200664.
- Sacristan-Soriano O, Banaigs B, Becerro MA. Relevant spatial scales of chemical variation in *Aplysina aerophoba*. Mar Drugs. 2011;9(12):2499-513. doi: 10.3390/md 9122499, PMID 22363236.
- Kämpfer P, Glaeser SP, Busse HJ, Abdelmohsen UR, Hentschel U. Rubrobacter aplysinae sp. nov., isolated from the marine sponge Aplysina aerophoba. Int J Syst Evol Microbiol. 2014; 64(3)(Pt_3):705-9. doi: 10.1099/ijs.0.055152-0, PMID 24170773.
- García GV JA, Martínez-Póveda BA, Rodríguez-Quesada AM, Medina-Torres MÁ. (+)-Aeroplysinin-1 Modulates the Redox Balance of Endothelial Cells.
- Vidal I, Castilla L, Marrero AD, Bravo-Ruiz I, Bernal M, Manrique I, et al. The sponge-derived brominated compound Aeroplysinin-1 impairs the endothelial inflammatory response through inhibition of the NF-κB pathway. Mar Drugs. 2022;20(10):605. doi: 10.3390/md20100605, PMID 36286429.
- Rodríguez-Nieto S, González-Iriarte M, Carmona R, Muñoz-Chápuli R, Medina MA, Quesada AR. Antiangiogenic activity of aeroplysinin-1, a brominated compound isolated from a marine sponge. FASEB J. 2002;16(2):261-3. doi: 10.1096/fj.01-0427fje , PMID 11772945.
- Wang CY, Wang BG, Brauers G, Guan HS, Proksch P, Ebel R. two novel γ-pyrone derivatives from the sponge-derived fungus *Microsphaeropsis* sp. J Nat Prod. 2002;65(5):772-5. doi: 10.1021/np0104828, PMID 12027766.
- Hentschel U, Schmid M, Wagner M, Fieseler L, Gernert C, Hacker J. Isolation and phylogenetic analysis of bacteria with antimicrobial activities from the Mediterranean sponges *Aplysina aerophoba* and *Aplysina cavernicola*. FEMS Microbiol Ecol. 2001;35(3):305-12. doi: 10.1111/j.1574-6941.2001.tb00816.x, PMID 11311441.
- Gerçe B, Schwartz T, Voigt M, Rühle S, Kirchen S, Putz A, et al. Morphological, bacterial, and secondary metabolite changes of *Aplysina aerophoba* upon long-term maintenance under artificial conditions. Microb Ecol. 2009;58(4):865-78. doi: 10.1007 /s00248-009-9560-6, PMID 19588186.
- Hamoda AM, Fayed B, Ashmawy NS, El-Shorbagi AA, Hamdy R, Soliman SS. Marine sponge is a promising natural source of anti-SARS-CoV-2 scaffold. Front Pharmacol. 2021;12:666664. doi: 10.3389/fphar.2021.6666664, PMID 34079462.
- Lira SP, Seleghim MH, Williams DE, Marion F, Hamill P, Jean F, et al. A SARS-corono virus 3CL protease inhibitor isolated from the marine sponge Axinella cf. corrugata: structure elucidation and synthesis. J Braz Chem Soc. 2007;18(2):440-3. doi: 10.1590 /S0103-S0532007000200030.
- Trianto A, Widyaningsih S, Radjasa OK, Pribadi R. Symbiotic fungus of marine sponge Axinella sp. producing antibacterial agent. IOP Conf S Earth Environ Sci. 2017 (Vol. 55, No. 1, p. 012005).
- Dresch RR, Lerner CB, Mothes B, Trindade VM, Henriques AT, Vozári-Hampe MM. Biological activities of ACL-I and physicochemical properties of ACL-II, lectins isolated from the marine sponge *Axinella corrugata*. Comp Biochem Physiol B Biochem Mol Biol. 2012;161(4):365-70. doi: 10.1016/j.cbpb.2012.01.001, PMID 22245532.
- Muthiyan R, Nambikkairaj B, Mahanta N, Immanuel T, Mandal RS, Kumaran K, et al. Antiproliferative and proapoptotic activities of marine spongeHyrtios erectus extract on breast carcinoma cell line (MCF-7). Pharmacogn Mag. 2017; 13 Suppl 1:S41-7. doi: 10.4103/0973-1296.203983 [ePub]. PMID 28479725, PMCID PMC5407115.
- Abd El-Moneam NM, Shreadah MA, El-Assar SA, Nabil-Adam A. Protective role of antioxidants capacity of *Hyrtios* aff. Erectus sponge extract against mixture of persistent organic pollutants (POPs)-induced hepatic toxicity in mice liver: biomarkers and ultrastructural study. Environ Sci Pollut Res Int. 2017;24(27):22061-72. doi: 10.10 07/s11356-017-9805-8, PMID 28791578.

- Kwon OS, Kim D, Kim CK, Sun J, Sim CJ, Oh DC, et al. Cytotoxic scalarane sesterterpenes from the sponge Hyrtios erectus. Mar Drugs. 2020;18(5):253. doi: 10.3390/md18050 253, PMID 32414015.
- Chakraborty K, Francis P. Hyrtioscalaranes A and B, two new scalarane-type sesterterpenes from *Hyrtios* erectus with anti-inflammatory and antioxidant effects. Nat Prod Res. 2021;35(24):5559-70. doi: 10.1080/14786419.2020.1795854, PMID 32713194.
- Hawas UW, Abou El-Kassem LT, Abdelfattah MS, Elmallah MI, Eid MA, Monier MM, et al. Cytotoxic activity of alkyl benzoate and fatty acids from the red sea sponge Hyrtios erectus. Nat Prod Res. 2018;32(12):1369-74. doi: 10.1080/14786419.2017.13 44662, PMID 28669229.
- 27. Ju E, Latif A, Kong CS, Seo Y, Lee YJ, Dalal SR, *et al.* Antimalarial activity of the isolates from the marine sponge *Hyrtios erectus* against the chloroquine-resistant Dd2 strain of Plasmodium falciparum. Z Naturforsch C J Biosci. 2018;73(9-10):397-400. doi: 10.1 515/znc-2018-0025, PMID 29813035.
- Sauleau P, Martin MT, Dau ME, Youssef DT, Bourguet-Kondracki ML. Hyrtiazepine, an azepino-indole-type alkaloid from the Red Sea marine sponge *Hyrtios erectus*. J Nat Prod. 2006;69(12):1676-9. doi: 10.1021/np060132r, PMID 17190441.
- Du L, Shen L, Yu Z, Chen J, Guo Y, Tang Y, *et al.* Hyrtiosal, from the marine sponge *Hyrtios erectus*, inhibits HIV-1 integrase binding to viral DNA by a new inhibitor binding site. Chem Med Chem. 2008;3(1):173-80. doi: 10.1002/cmdc.200700223, PMID 17943714.
- Sun T, Wang Q, Yu Z, Zhang Y, Guo Y, Chen K, et al. Hyrtiosal, a PTP1B inhibitor from the marine sponge Hyrtios erectus, shows extensive cellular effects on PI3K/ AKT activation, glucose transport, and TGF-β/Smad2 signaling. Chem bio chem. 2007;8(2):187-93. doi: 10.1002/cbic.200600349, PMID 17183521.
- Karimi E, Ramos M, Gonçalves JM, Xavier JR, Reis MP, Costa R. Comparative metagenomics reveals the distinctive adaptive features of the *Spongia officinalis* endosymbiotic consortium. Front Microbiol. 2017;8:2499. doi: 10.3389/fmicb.2017. 02499, PMID 29312205.
- Manzo E, Ciavatta ML, Villani G, Varcamonti M, Sayem SM, Van Soest R, *et al.* Bioactive terpenes from *Spongia officinalis*. J Nat Prod. 2011;74(5):1241-7. doi: 10.1021/np200 226u, PMID 21548580.
- Han GY, Sun DY, Liang LF, Yao LG, Chen KX, Guo YW. Spongian diterpenes from Chinese marine sponge Spongia officinalis. Fitoterapia. 2018;127:159-65. doi: 10.1016/j.fitote .2018.02.010, PMID 29447982.
- Jin T, Li P, Wang C, Tang X, Yu X, Sun F, et al. pokepola esters, and sponalisolides from the aquaculture sponge Spongia officinalis L. Phytochemistry. 2022;194:113006.
- Jin T, Li P, Wang C, Tang X, Yv X, Li K, et al. Two new spongian diterpene derivatives from the aquaculture sponge Spongia officinalis Linnaeus., 1759. Nat Prod Res. 2023;37(2):216-26. doi: 10.1080/14786419.2021.1961137, PMID 34348550.
- Liu J, Liu ZY, Fu Y, Gu YC, Li SW, Zhang HY et al. Anti-inflammatory steroids from the South China sea sponge Spongia officinalis. Chem Bio divers. 2024;21(6):e202400519. doi: 10.1002/cbdv.202400519, PMID 38576052.
- Bhattacharya D, Lai TK, Saha A, Selvin J, Mukherjee J. Structural elucidation and antimicrobial activity of a diketopiperazine isolated from a *Bacillus* sp. associated with the marine sponge *Spongia officinalis*. Nat Prod Res. 2021;35(14):2315-23. doi: 1 0.1080/14786419.2019.1672684, PMID 31583909.
- Dellai A, Mansour HB, Clary-Laroche A, Deghrigue M, Bouraoui A. Anticonvulsant and analgesic activities of crude extract and its fractions of the defensive secretion from the Mediterranean sponge, *Spongia officinalis*. Cancer Cell Int. 2012;12(1):15. doi: 10. 1186/1475-2867-12-15, PMID 22494441.
- Kaliaperumal K, Salendra L, Liu Y, Ju Z, Sahu SK, Elumalai S, et al. Isolation of anticancer bioactive secondary metabolites from the sponge-derived endophytic fungi Penicillium sp. and in silico computational docking approach. Front Microbiol. 2023;14:1216928. doi: 10.3389/fmicb.2023.1216928, PMID 37849927.
- Dellai A, Deghrigue M, Laroche-Clary A, Masour HB, Chouchane N, Robert J, *et al.* Evaluation of antiproliferative and anti-inflammatory activities of methanol extract and its fractions from the Mediterranean sponge. Cancer Cell Int. 2012;12(1):18. doi: 10.1186/1475-2867-12-18, PMID 22587650.
- Riesgo A, Taboada S, Pérez-Portela R, Melis P, Xavier JR, Blasco G, et al. Genetic diversity, connectivity and gene flow along the distribution of the emblematic Atlanto-Mediterranean sponge Petrosia ficiformis (Haplosclerida, Demospongiae). BMC Evol Biol. 2019;19(1):24. doi: 10.1186/s12862-018-1343-6, PMID 30651060.
- Guo Y, Gavagnin M, Salierno C, Cimino G. Further Petroformynes from Both Atlantic and Mediterranean Populations of the Sponge *Petrosia ficiformis*. J Nat Prod. 1998;61(3):333-7. doi: 10.1021/np970424f, PMID 9548871.
- Seidel SB, Proudfoot JR, Djerassi C, Sica D, Sodano G. Minor and trace sterols from marine invertebrates 56. Novel coprostanols from the marine sponge *Petrosia ficiformis*. Steroids. 1986;47(1):49-62. doi: 10.1016/0039-128x(86)90076-0, PMID 3101232.
- Khalil MW, Djerassi C, Sica D. Minor and trace sterols in marine invertebrates XVII.
 Steroids. 1980; 35(6); 24R(24), 26-dimethylcholesta-5, 26-dien-3β-01, a new sterol from the sponge *Petrosia ficiformis*:707-19.
- Chelossi E, Milanese M, Milano A, Pronzato R, Riccardi G. Characterisation and antimicrobial activity of epibiotic bacteria from *Petrosia ficiformis* (Porifera, *Demospongiae*). J Exp Mar Biol Ecol. 2004;309(1):21-33. doi: 10.1016/j.jembe.2004.0 3.006.

- 46. O'Keefe BR, Beutler JA, Cardellina JH, Gulakowski RJ, Krepps BL, Mcmahon JB, et al. Isolation and characterization of niphatevirin, a human-immunodeficiency-virusinhibitory glycoprotein from the marine sponge Niphates erecta. Eur J Biochem. 1997;245(1):47-53. doi: 10.1111/j.1432-1033.1997.t01-1-00047.x, PMID 9128723.
- Silva IT, Caon T, Lückemeyer DD, Ramos FA, Tello E, Arévalo-Ferro C, *et al*. Antiherpes screening of marine organisms from Colombian Caribbean Sea. Rev Bras Farmacognosia. 2011;21(4):608-14. doi: 10.1590/S0102-695X2011005000094.
- 48. Hiscock K. Halichondria panicea (Halichondria) Breadcrumb sponge. In: Plymouth: Marine Biological Association of the United Kingdom; 2008. Tyler-Walters H. Marine life information network: biology and sensitivity key information reviews [online]. [cited 14-1-2025] Available from: https://www.marlin.ac.uk/species/detail/1407.
- Rodriguez Jimenez A, Dechamps E, Giaux A, Goetghebuer L, Bauwens M, Willenz P, et al. The sponges Hymeniacidon perlevis and Halichondria panicea are reservoirs of antibiotic-producing bacteria against multi-drug resistant Staphylococcus aureus. J Appl Microbiol. 2021;131(2):706-18. doi: 10.1111/jam.14999, PMID 33421270.
- Nagle DG, McClatchey WC, Gerwick WH. New glycosphingolipids from the marine sponge *Halichondria panicea*. J Nat Prod. 1992;55(7):1013-7. doi: 10.1021/np50085 a032, PMID 1402953.
- Schneemann I, Ohlendorf B, Zinecker H, Nagel K, Wiese J, Imhoff JF, et al. γ-pyrones from a Nocardiopsis strain isolated from the marine sponge Halichondria panicea. J Nat Prod. 2010;73(8):1444-7. doi: 10.1021/np100312f, PMID 20695474.
- Schneemann I, Kajahn I, Ohlendorf B, Zinecker H, Erhard A, Nagel K, et al. Mayamycin, a cytotoxic polyketide from a Streptomyces strain isolated from the marine sponge Halichondria panicea. J Nat Prod. 2010;73(7):1309-12. doi: 10.1021/np100135b, PMID 20545334.
- Yang JH, Suh SJ, Lu Y, Li X, Lee YK, Chang YC, et al. Anti-inflammatory activity of ethylacetate fraction of *Cliona celata*. Immunopharmacol Immunotoxicol. 2011;33(2):373-9. doi: 10.3109/08923973.2010.520716, PMID 20929426.
- Reegan AD, Kinsalin AV, Paulraj MG, Ignacimuthu S. Larvicidal, ovicidal and repellent activities of marine sponge *Cliona celata* (Grant) extracts against *Anopheles stephensi* Liston (*Diptera: Culicidae*). Asian Pac J Trop Med. 2015;8(1):29-34. doi: 10.1016/ S1995-7645(14)60183-8, PMID 25901921.
- 55. Suh SJ, Kwak CH, Song KH, Kwon KM, Chung TW, Cho SH, et al. Triple inhibitory activity of *Cliona celata* against TNF-α-induced matrix metalloproteinase-9 production via downregulated NF-κB and AP-1, enzyme activity, and migration potential. Inflammation. 2012;35(2):736-45. doi: 10.1007/s10753-011-9369-6, PMID 21845471.
- Forestieri R, Donohue E, Balgi A, Roberge M, Andersen RJ. Synthesis of clionamine B, an autophagy stimulating aminosteroid isolated from the sponge *Cliona celata*. Org Lett. 2013;15(15):3918-21. doi: 10.1021/ol4016783, PMID 23869546.
- 57. Alves J, Gaspar H, Silva J, Alves C, Martins A, Teodoro F, *et al.* Unravelling the anti-inflammatory and antioxidant potential of the marine sponge *Cliona celata* from the Portuguese coastline. Mar Drugs. 2021;19(11):632. doi: 10.3390/md19110 632, PMID 34822503.
- Corticium-candelabrum. Available from: https://www.inaturalist.org/taxa/ 604291-Corticium-candelabrum.
- 59. de Caralt S, Agell G, Uriz MJ. Long-term culture of sponge explants: conditions enhancing survival and growth, and assessment of bioactivity. Biomol Eng. 2003;20(4-6):339-47. doi: 10.1016/s1389-0344(03)00045-5, PMID 12919818.
- 60. Rady H. Sponge mesohyl induces anti-proliferation activity and cell cycle arrest in colon cancer *in vitro*. Res J Pharm Biol Chem Sci. 2014;5(6):1070-4.
- Soapi K, Feussner KD, Aalbersberg WG. Antimicrobial and cytotoxic activities of marine plants and invertebrates from the coast of Espirito Santo in Vanuatu. S Pac J Nat App Sci. 2013;31(1):89-95. doi: 10.1071/SP13010.
- Rady H, Salem S, Ez El-Arab ME. Primmorph extracts and mesohyls of marine sponges inhibit proliferation and migration of hepatocellular carcinoma cells *in vitro*. J Pharm Anal. 2019;9(4):284-91. doi: 10.1016/j.jpha.2019.03.008, PMID 31452967.
- Murayama S, Imae Y, Takada K, Kikuchi J, Nakao Y, Van Soest RW, et al. Shishicrellastatins, inhibitors of cathepsin B, from the marine sponge Crella spinulata (Yvesia). Bioorg Med Chem. 2011;19(22):6594-8. doi: 10.1016/j.bmc.2011.06.052, PMID 21764589.
- Aiello A, Fattorusso E, Menna M, Pansini M. Further bioactive acetylenic compounds from the Caribbean sponge Cribrochalina vasculum. J Nat Prod. 1992;55(9):1275-80. doi: 10.1021/np50087a015, PMID 1431945.
- 65. Zovko A, Novak M, Hååg P, Kovalerchick D, Holmlund T, Färnegårdh K, et al. Compounds from the marine sponge Cribrochalina vasculum offer a way to target IGF-1R mediated signaling in tumor cells. On cotarget. 2016;7(31):50258-76. doi: 10.1 8632/oncotarget.10361, PMID 27384680.
- Aiello A, Fattorusso E, Menna M, Pansini M. Further bioactive acetylenic compounds from the Caribbean sponge *Cribrochalina vasculum*. J Nat Prod. 1992;55(9):1275-80. doi: 10.1021/np50087a015, PMID 1431945.
- Carballeria NM, Reyes ED. Identification of the new 23-methyl-5, 9-pentacosadienoic acid in the sponge *Cribrochalina vasculum*. Lipids. 1990;25(1):69-71. doi: 10.1007/BF 02562431, PMID 2325509.
- Giner JL, Djerassi C. Minor and trace sterols in marine invertebrates 65. 23-Epidihydrocalysterol: a new cyclopropane-containing sponge sterol. Steroids. 1992;57(6):258-61. doi: 10.1016/0039-128x(92)90057-g, PMID 1440695.
- 69. De Rosa S, Crispino A, De Giulio A, Iodice C, Amodeo P, Tancredi T. A new cacospongionolide derivative from the sponge *Fasciospongia cavernosa*. J Nat Prod. 1999;62(9):1316-8. doi: 10.1021/np990125I, PMID 10514323.

- Sadanandan R, Rauf AA. Isolation, Purification and Characterisation of a D-galactose and N-acetyl-D-galactosamine Specific Lectin from Marine Sponge *Fasciospongia cavernosa*. Protein Pept Lett. 2018;25(9):871-7. doi: 10.2174/0929866525666180905 111452, PMID 30182831.
- 71. De Rosa S, Crispino A, De Giulio A, lodice C, Benrezzouk R, Terencio MC, et al. A new cacospongionolide inhibitor of human secretory phospholipase A2 from the Tyrrhenian sponge *Fasciospongia cavernosa* and absolute configuration of cacospongionolides. J Nat Prod. 1998;61(7):931-5. doi: 10.1021/np980122t, PMID 9677277.
- 72. De Stefano D, Tommonaro G, Malik SA, Iodice C, De Rosa S, Maiuri MC, *et al.* Cacospongionolide and scalaradial, two marine sesterterpenoids as potent apoptosis-inducing factors in human carcinoma cell lines. PLOS One. 2012;7(4):e33031. doi: 10.1371/journal.pone.0033031, PMID 22509253.
- 73. Posadas I, De Rosa S, Terencio MC, Payá M, Alcaraz MJ. Cacospongionolide B suppresses the expression of inflammatory enzymes and tumour necrosis factor-α by inhibiting nuclear factor-κB activation. Br J Pharmacol. 2003;138(8):1571-9. doi: 1 0.1038/sj.bjp.0705189, PMID 12721113.
- 74. García Pastor PG, De Rosa S, De Giulio A, Payá M, Alcaraz MJ. Modulation of acute and chronic inflammatory processes by cacospongionolide B, a novel inhibitor of human synovial phospholipase A2. Br J Pharmacol. 1999;126(1):301-11. doi: 10.1038/sj.bjp. 0702302, PMID 10051149.
- Posadas I, Terencio MC, De Rosa S, Payá M. Cavernolide: a new inhibitor of human sPLA2 sharing unusual chemical features. Life Sci. 2000;67(24):3007-14. doi: 10.1016/ s0024-3205(00)00875-4, PMID 11133013.
- 76. 76. Available from: https://invasions.si.edu/nemesis/species_summary/48398.
- 77. Zhong W, Olugbami JO, Rathakrishnan P, Mohanty I, Moore SG, Garg N, et al. Discovery and Folding Dynamics of a Fused bicyclic cysteine Knot undecapeptide from the Marine Sponge Halichondria bowerbanki. J Org Chem. 2024;89(17):12748-52. doi: 1 0.1021/acs.joc.4c01104, PMID 39189383.
- Ahmadi P, Haruyama T, Kobayashi N, de Voogd NJ, Tanaka J. Spongian diterpenes from the Sponge Hyattella aff. intestinalis. Chem Pharm Bull (Tokyo). 2017;65(9):874-7. doi: 10.1248/cpb.c17-00297, PMID 28652548.
- Somerville MJ, Hooper JN, Garson MJ, Mooloolabenes AE. Mooloolabenes A-E, norsesterterpenes from the Australian sponge *Hyattella intestinalis*. J Nat Prod. 2006;69(11):1587-90. doi: 10.1021/np060244i, PMID 17125226.
- Inbaneson SJ, Ravikumar S. In vitro antiplasmodial activity of marine sponge Hyattella intestinalis associated bacteria against Plasmodium falciparum. Asian Pac J Trop Biomed. 2011;1(1):S100-4. doi: 10.1016/S2221-1691(11)60133-0.
- Muralidhar P, Krishna N, Kumar MM, Rao CB, Rao DV. New sphingolipids from marine sponge lotrochota baculifera. Chem Pharm Bull (Tokyo). 2003;51(10):1193-5. doi: 10. 1248/cpb.51.1193, PMID 14519929.
- 82. Thompson MN, Gallimore WA. Constituents of the Jamaican Sponge *lotrochota birotulata*. World J Org Chem. 2016;1;4:13-6.
- Almeida AS, Mendonça DN, Carneiro RF, Pinheiro U, Nascimento EF, Andrade AL, et al. Purification, biochemical characterization of a lectin from marine sponge *Ircinia* strobilina and its effect on the inhibition of bacterial biofilms. An Acad Bras Cienc. 2023;95 Suppl 2:e20220619. doi: 10.1590/0001-3765202320220619, PMID 38088730.
- Carballeira NM, Shalabi F, Cruz C, Rodriguez J, Rodriguez E. Comparative study of the fatty acid composition of sponges of the genus Ircinia. Identification of the new 23-methyl-5, 9-tetracosadienoic acid. Comp Biochem Physiol B. 1991;100(3):489-92. doi: 10.1016/0305-0491(91)90209-v, PMID 1814678.
- Zabriskie TM, Ireland CM. The isolation and structure of modified bioactive nucleosides from Jaspis johnstoni. J Nat Prod. 1989;52(6):1353-6. doi: 10.1021/np5 0066a032, PMID 2693614.
- Bubb MR, Senderowicz AM, Sausville EA, Duncan KL, Korn ED. Jasplakinolide, a cytotoxic natural product, induces actin polymerization and competitively inhibits the binding of phalloidin to F-actin. J Biol Chem. 1994;269(21):14869-71. doi: 10.101 6/S0021-9258(17)36545-6, PMID 8195116.
- Vargas S, Leiva L, Wörheide G. Short-term exposure to high-temperature water causes a shift in the microbiome of the common aquarium sponge *Lendenfeldia chondrodes*. Microb Ecol. 2021;81(1):213-22. doi: 10.1007/s00248-020-01556-z, PMID 32767091.
- Saïd Hassane C, Tintillier F, Campos PE, Herbette G, de Voogd NJ, Ouazzani J, et al. Polybrominated diphenyl ethers isolated from the marine sponge *Lendenfeldia chondrodes* collected in Mayotte. Nat Prod Res. 2024;38(17):2973-82. doi: 10.1080/14 786419.2023.2204431, PMID 37086477.
- Sakai R, Kamiya H. 1-deoxynojirimycin derivatives from the marine sponge Lendenfeldia chondrodes. J Antibiot (Tokyo). 2006;59(8):507-11. doi: 10.1038/ja.200 6.71, PMID 17080688.
- Sera Y, Adachi K, Shizuri Y. A new Epidioxy sterol as an antifouling substance from a Palauan marine sponge, *Lendenfeldia chondrodes*. J Nat Prod. 1999;62(1):152-4. doi: 1 0.1021/np980263v, PMID 9917306.
- Luo X, Wang Q, Tang X, Xu J, Wang M, Li P, et al. Cytotoxic manoalide-type sesterterpenes from the sponge Luffariella variabilis collected in the South China Sea. J Nat Prod. 2021;84(1):61-70. doi: 10.1021/acs.jnatprod.0c01026, PMID 33371684.
- Ahmadi P, Higashi M, de Voogd NJ, Tanaka J. Two furanosesterterpenoids from the sponge Luffariella variabilis. Mar Drugs. 2017;15(8):249. doi: 10.3390/md15080249, PMID 28796183.

- Ettinger-Epstein P, Motti CA, de Nys R, Wright AD, Battershill CN, Tapiolas DM. Acetylated sesterterpenes from the Great Barrier Reef sponge *Luffariella variabilis*. J Nat Prod. 2007;70(4):648-51. doi: 10.1021/np060240d, PMID 17295541.
- 94. Hamada T, Harada D, Hirata M, Yamashita K, Palaniveloo K, Okamura H, *et al.* Manoalide-related sesterterpene from the marine sponge *Luffariella variabilis*. Nat Prod Commun. 2015; 10(6): 1934578X1501000616: 863-4. doi: 10.1177/1934578X15 01000616, PMID 26197501.
- Gauvin-Bialecki A, Aknin M, Smadja J. 24-O-Ethylmanoalide, a manoalide-related sesterterpene from the marine sponge *Luffariella* cf. variabilis. Molecules. 2008;13(12):3184-91. doi: 10.3390/molecules13123184, PMID 19078858.
- 96. Sakai E, Kato H, Rotinsulu H, Losung F, Mangindaan RE, de Voogd NJ, et al. Variabines A and B: new β-carboline alkaloids from the marine sponge Luffariella variabilis. J Nat Med. 2014;68(1):215-9. doi: 10.1007/s11418-013-0778-8, PMID 23686294.
- Yeom JH, Kim HY, Lim JH, Yoon KW, Kim HM, Jeong HJ. A calcium channel blocker, manoalide exerts an anti-allergic inflammatory effect through attenuating NF-κB activity. Immunopharmacol Immunotoxicol. 2021;43(6):799-805. doi: 10.1080/0892 3973.2021.1988101, PMID 34708672.
- Kernan MR, Faulkner DJ, Jacobs RS. The luffariellins, novel antiinflammatory sesterterpenes of chemotaxonomic importance from the marine sponge *Luffariella* variabilis. J Org Chem. 1987;52(14):3081-3. doi: 10.1021/jo00390a021.

- Cavalcanti BC, Sombra CM, de Oliveira JH, Berlinck RG, de Moraes MO, Pessoa C. Cytotoxicity and genotoxicity of ingenamine G isolated from the Brazilian marine sponge *Pachychalina alcaloidifera*. Comp Biochem Physiol C Toxicol Pharmacol. 2008;147(4):409-15. doi: 10.1016/j.cbpc.2008.01.005, PMID 18291725.
- de Oliveira JH, Nascimento AM, Kossuga MH, Cavalcanti BC, Pessoa CO, Moraes MO, et al. Cytotoxic alkylpiperidine alkaloids from the Brazilian marine sponge Pachychalina alcaloidifera. J Nat Prod. 2007;70(4):538-43. doi: 10.1021/np060450q, PMID 17346073.
- Berrué F, McCulloch MW, Boland P, Hart S, Harper MK, Johnston J, et al. Isolation of steroidal glycosides from the Caribbean sponge Pandaros acanthifolium. J Nat Prod. 2012;75(12):2094-100. doi: 10.1021/np300520w, PMID 23245401.
- Regalado EL, Tasdemir D, Kaiser M, Cachet N, Amade P, Thomas OP. Antiprotozoal steroidal saponins from the marine sponge *Pandaros acanthifolium*. J Nat Prod. 2010;73(8):1404-10. doi: 10.1021/np100348x, PMID 20614907.
- Cachet N, Regalado EL, Genta-Jouve G, Mehiri M, Amade P, Thomas OP. Steroidal glycosides from the marine sponge *Pandaros acanthifolium*. Steroids. 2009;74(9):746-50. doi: 10.1016/j.steroids.2009.03.009, PMID 19541002.
- Regalado EL, Turk T, Tasdemir D, Gorjanc M, Kaiser M, Thomas OP, et al. Cytotoxic and haemolytic steroidal glycosides from the Caribbean sponge Pandaros acanthifolium. Steroids. 2011;76(12):1389-96. doi: 10.1016/j.steroids.2011.07.010, PMID 21820457.

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