A Comprehensive Review of Ethnomedicinal, Phytochemical and Pharmacological Activity Profile of Achyranthes aspera

Nem Kumar Jain^{1,*}, Shubham Anand^{1,*}, Priyanka Keshri², Santosh Kumar³, Akshay Singh Sengar⁴, Manoj Kumar Bajhaiya¹, Devendra Dhanorya⁵, Sapna Yadav¹, Hema Katra¹, Sajal Mishra¹

¹Department of Pharmacology, School of Pharmacy, ITM University, Gwalior, Madhya Pradesh, INDIA. ²Department of Pharmaceutics, School of Pharmacy, ITM University, Gwalior, Madhya Pradesh, INDIA. ³Department of Life Sciences, School of Sciences, ITM University, Gwalior, Madhya Pradesh, INDIA. ⁴Department of Food Technology, School of Sciences, ITM University, Gwalior, Madhya Pradesh, INDIA. ⁵Department of Pharmacy, Mangalaytan University, Jabalpur, Madhya Pradesh, INDIA.

ABSTRACT

For centuries, humans have harnessed the healing properties of plants to address diverse health conditions. Ancient systems like Ayurveda and Unani medicine hold a rich repository of knowledge extolling the virtues of herbal remedies. *Achyranthes aspera*, one prolific native herbaceous plant in the South America, Asia and Africa often prospers as one common weed. Its many applications in traditional medicine for treating various diseases are at the heart of this global research. *Achyranthes aspera* is diverse many chemical ingredients, including terpenoids, alkaloids, steroids, saponins and flavonoids. This paper provides a thorough examination and elucidation of the phytochemical composition across different parts of *Achyranthes aspera*, delving into its traditional uses and expounding upon its pharmacological properties.

Keywords: Achyranthes aspera, Antidiabetic activity, Anti-oxidant, Anti-inflammatory activity, Anti-microbial activity.

Correspondence:

Mr. Nem Kumar Jain

Department of Pharmacology, School of Pharmacy, ITM University, Gwalior, Madhya Pradesh, INDIA. Email: nemkumar89@gmail.com;

Mr. Shubham Anand

Department of Pharmacology, School of Pharmacy, ITM University, Gwalior, Madhya Pradesh, INDIA. Email: anandshubham582@gmail.com

Received: 02-10-2023; Revised: 21-11-2023; Accepted: 13-05-2024.

INTRODUCTION

Throughout history, nature has served as a wellspring of medicinal compounds, yielding a notable array of innovative drugs.^[1] Recent times have witnessed a remarkable surge in the production and utilization of plant derived health products, both in developed and developing nations. This surge has led to an exponential growth in the global herbal product market. The World Health Organization (WHO) has identified a staggering 22,000 species of medicinal plants in its endeavor to catalogue global usage.^[2]

According to a survey conducted by the WHO, a striking 80% of the population in developing countries predominantly relies on traditional herbal medicine for their healthcare needs.^[3] The exploration of plant chemical constituents and subsequent pharmacological assessments forms the bedrock for discovering potential leads in the development of new therapeutic agents.



DOI: 10.5530/pres.16.3.57

Copyright Information : Copyright Author (s) 2024 Distributed under Creative Commons CC-BY 4.0

Publishing Partner : EManuscript Tech. [www.emanuscript.in]

Many of today's life-saving drugs in modern medicine are derived from plants.^[4]

Because of the cumulative and persistent side effects associated with modern medications, there has been a notable trend toward herbal medicines in recent years. However, this natural reservoir and the traditional knowledge associated with it face escalating threats from urbanization, population surges and the ongoing exploitation of these herbal reserves.^[5] Presently, the evaluation of plants based on their traditional uses plays a pivotal role in the quest for novel drug agents. One such plant under scrutiny is *Achyranthes aspera*, commonly known as prickly chaff flower in English and Chirchira in Hindi. This article aims to provide information about the chemical constituents found in the different parts of *Achyranthes aspera*, as well as elucidate its various pharmacological effects, which underlie its many applications. Its use in the treatment of many diseases is well-documented.

Geographical distribution

Achyranthes aspera thrives as a common weed in India and other regions, growing along roadsides, field edges, and in abandoned areas at altitudes of up to 2,100 m. Its presence is not limited to India but can also be found in Australia, Bangladesh, South



Figure 1: Morphology of Achyranthes aspera.

Andaman Islands, America, Ceylon, Africa and various parts of tropical Asia.^[6]

Plant characteristics

Achyranthes aspera is characterized as a perennial or sometimes annual herbaceous plant, with a prostrate or erect habit (Figure 1).^[7]

Seeds: The seeds possess a brown hue, featuring a truncated apex and a rounded base. They are endospermic and sub-cylindric in shape.^[8]

Height: Typically, it attains a height ranging from 0.2 to 2.0 m, often supported by a woody base.^[9]

Leaf: The leaves are arranged oppositely, bearing a velvety, tomentose texture. They take on an obovate form, with undulating margins and a white hairy surface. The leaf petiole has a crescent-shaped outline and consists of a single-layer cuticle with a thick cuticle. The midrib has a single-layer epidermis, surrounded by 4 to 5 layers of parenchyma on the upper surface and 2 to 3 layers of parenchyma on the lower surface.^[10] Leaf dimensions vary, with an average length of 5.22 cm and width of 2.5 cm.^[11] They can be found in assorted sizes, featuring anomocytic stomata on the lower epidermis.^[12]

Branch: The branches are either terete or quadrangular, marked by striations and a pubescent surface. They support thick leaves.

Roots: The root system comprises secondary and tertiary roots. These roots are cylindrical in shape, exhibiting a diameter ranging from 0.1 to 1.0 cm. They possess a somewhat ribbed texture, gradually tapering towards the ends and are tinted yellowish-brown.^[13] Flowers: Arranged in spikes, the flowers vary in length from 8 to 30 cm, with a width of 3 to 7 mm. They are bisexual, presenting a greenish-white hue and occur in numerous quantities. The flowers are sessile, bracteate and have two bracts, one of which has a spiny lip. They exhibit optical symmetry and subcellular arrangement. The perianth comprises five membranous segments, while there are five stamens with short

filaments and a two-celled anther. Flowering typically occurs in the summer season.

Trunk: The stem is simple or branched, has ribs, angular borders and is often purple in color. Fruit: The fruits are dry indehiscent capsules, enclosed in persistent bracts and perianths. They contain a seeded drupe or utricle.^[14]

Traditional uses

Achyranthes aspera finds mention in both Ayurvedic and Chinese medicinal traditions.^[15] In Ayurvedic texts known as "Nighantus," The plant is described for its medicinal properties as a digestive aid, purgative, medicine for internal inflammation, itching, haemorrhoids, cervical gland hypertrophy and abdominal enlargement.^[16] Hindus use ash taken from the whole plant to make caustic alkaline preparations.^[17]

European and Indian doctors know the plant's diuretic properties.^[18] In cases of general edema and renal edema, a decoction of the plant is used as a diuretic.^[19] In the Philippines, this herb is used to treat toothache, gastrointestinal problems and dysentery.^[20] The plant is used for many therapeutic purposes, including the treatment of asthma, indigestion, bronchitis, flatulence and menstrual disorders. It also acts as an expectorant, analgesic, purifying, anthelmintic, diaphoretic and gastric suppressant.^[21] The roots are used to treat coughs, stomach tumours and stomach-related diseases.^[22]

In the tribal communities of Chitoor district of Andhra Pradesh, this herb is used to treat epilepsy, Scorpion bite. Additionally, a remedy made from its seeds and milk, known as Payasam Kheer, is considered effective for brain-related ailments.^[23]

Achyranthes aspera serves various therapeutic roles, acting as a diuretic, astringent, purgative and remedy for conditions such as abdominal pain, ascites, haemorrhoids^[24] and skin rashes.^[25] It also acts as a laxative^[26] and as an antidote for snakebites, In addition, it is also used in the treatment of bone fractures,^[27] whooping cough, respiratory problems^[28] asthma^[29] and leukaemia.^[30] The inflorescence is employed in cases of hydrophobia^[31] and cough,^[32] while the fruit is used to address hydrophobia.^[33] The seeds find application in treating gonorrhea, insect bites, fear of water, cough (especially whooping cough) and anti- asthma.^[34] They are also known for their cathartic, purgative and emetic properties.^[35] The leaves are used to treat conditions such as dog bites, intermittent fever, asthma, ulcers, wounds and typhoid, For whooping cough, tonsillitis, haemorrhage,^[36] cough and hydrophobia,^[37] the roots are used as an anti-asthma,^[38] diuretic, diaphoretic and anti- syphilis drug.[39]

Phytochemistry

The whole *Achyranthes aspera* plant was analyzed, showing its solubility in water and chloroform. Initially, the component known as achyranthine was identified as a betaine derivative of

Table 1: Taxanomic Classification.	
Plantae	
Tracheobionta	
Angiosperms	
Eudicots	
Core Eudicots	
Caryophyllales	

N-methylpyrrolidine-3-carboxylic acid.^[40] Subsequent studies clarified that betaine, not achyranthine, is soluble in water.^[41] Further studies demonstrated that the chloroform-soluble fraction consisted of a mixture of two unidentified alkaloid compounds.^[42] The ethanolic extract of the plant contained alkaloids and saponins, while tannins and flavonoids were not detected.^[43] Employing gas chromatography-mass spectrometry (GC-MS) technology, a separate study elucidated the phytochemical constituents within the hydro- alcoholic extract of *A. aspera's* whole plant.^[44] This analysis revealed the presence of 15 distinct phytocompounds, which are detailed in Table 1.^[45]

Shoot

From the shoots of *Achyranthes aspera*, a unique aliphatic dihydroxyketone named.^[46] Dihydroxyhenpentacontan-4 was synthesized along with tritriacontane. Another study showed the production of dihydroxy ketones from shoots.^[47] identified as 36,37- dihydroxyhenpentacontan-4-one and triacontanol.^[48] Additionally, a new long-chain alcohol, 17-pentatriacontanol.^[48] and an essential oil yield four distinct compounds: 4- methylheptatriacon-1-en-10-ol, 16-hydroxy-2 6-methylheptacosan-2-one and tetracontanol.^[49]

Stem

From the chloroform extract of the stem, unique compounds including pentatriacontanone, 6- pentatriacontanone, hexatriacontane and triacontane were successfully isolated, Ecdysterone, a distinctive component, was identified in the stem, employing ethyl acetate as the extracting agent, the stem yielded 3-acetoxy-6-benzoyloxyapangamide,^[50] a compound characterized by its distinct chemical structure.

Inflorescence

Alkaloids and flavonoids have been found in inflorescences.^[51]

Seeds

The seeds were found to be a rich source of protein, with a content of 24.8% and a calorific value of 3.92/g, comparable to Bengal seeds.^[52] Protein hydrolysis in the seeds revealed the presence of essential amino acids, with leucine, isoleucine, phenylalanine and valine contents equivalent to those of Bengal gram. Notably, tryptophan and sulfur amino acid (methionine and Cystine) contents exceed those of most legumes, while arginine, lysine and threonine are relatively lower than those in whole eggs.^[53] The composition of saponin sugar moieties includes glucose, galactose, xylose and rhamnose, the seeds contain a portion of crude sapogenin, which secretes oleanolic acid.^[54] Further research led to the isolation of two oleanolic saponins,^[55] identified as saponin A and saponin B. They are characterized as α -l-rhamnopyranosyl (1 \rightarrow 4)- β '-D -glucopyranosyl (1 \rightarrow 4)- β '-D glucuronopyranosyl (1 \rightarrow 3)-oleanolic acid and β -D-galactopyranosyl (1 \rightarrow 28) ester of saponin A, respectively.^[56] Acid hydrolysis of total saponins confirmed the presence of oleanolic acid.^[57] A rapid procedure for isolating triterpenoid saponins from plants has been described based on partition chromatography.^[58]

In the seeds, hexatriacontane, 10-octacosanone, 10-triacosanone and 4-triacosanone have been identified as constituents.^[59] The fatty fraction of seed oils includes lauric, myristic, palmitic, stearic, arachidic, behenic, oleic and linoleic acids, Additionally, the seeds were found to contain Ecdysterone.

Roots

Various preliminary studies have been conducted to discern the chemical composition of the root. It was noted that the saponin fraction contains oleanolic acid as its aglycone component, Additionally, analyses revealed the presence of saponins and alkaloids in both the root and shoot, with no detectable flavonoids.^[60] In a separate investigation, alkaloids were identified in the root, while saponins and tannins were not reported, Another preliminary chemical study unveiled the presence of steroids, flavonoids, alkaloids, saponins and terpenoids in the root, with no glycosides detected.^[61] Notably, β -sitosterol was also identified in the root in a separate study.

Pharmacology Antimicrobial properties

Diverse preliminary investigations have unveiled the multifaceted antibacterial attributes of *Achyranthes aspera*. The entire plant, along with the constituent achyranthine, demonstrated significant antibacterial efficacy against *Streptococcus haemolyticus*, *Staphylococcus aureus* and *Bacillus typhosus*.^[62] Aqueous and alcoholic extracts of the leaves showed notable antibacterial activity against *Staphylococcus Aureus* and *E. coli*.^[63] Seeds grown on cow manure showed antibacterial activity against *Salmonella typhimurium*, *Pseudomonas cichorii* and *B. subtilis* strains, In addition, 80% concentrated ethanolic extract of leaves and stems inhibited the growth of *B. subtilis* and *S. yellow*.^[64]

Additionally, diethyl ether extract from the leaves of *A. aspera* has demonstrated inhibitory effectiveness against a variety of microorganisms including *T. rubrum*, *E. floccosum*, *Enterobacter* sp., *S. aureus*, *Salmonella* sp., *Shigella* sp., *Trichophyton*

mentagrophytes, Aspergillus sp., T. tonurans, P. vulgaris, Klebsiella sp. to E. coli.^[65] The finished fabric derived from Achyranthes aspera displayed a notable reduction in bacterial count against Staphylococcus aureus and Escherichia coli.^[66] Furthermore, saponin extracted from the ethyl acetate extract of A. aspera exhibited larvicidal properties against mosquitoes.^[67,68] In in vivo experiments, aqueous extracts of the leaves showed specific antibacterial activity against Proteus vulgaris.^[69] with no apparent effect against Pseudomonas aeruginosa, Escherichia *coli*, or *Klebsiella aerogenes*.^[70] Another water residue from leaves was found to have no effect on a variety of bacteria, including Escherichia coli, Klebsiella aerogenes, Cytophaga sp., Pseudomonas aeruginosa, Vibrio parahaemolytica, Aeromonas hydrophilla, Damsela, Bacillus cereus and Streptococcus pyogenes.^[71] Additionally, leaf extracts also demonstrated antibacterial effects against E. coli, S. citri and aerobic spores found in soft drinks.^[72] Additionally, essential oil extracted from the buds demonstrated antifungal activity against Aspergillus cameus, inhibiting its mycelial growth, In a comparative study of herbal fumigants, Achyranthes aspera significantly reduced microbial colonies in air samples compared to formalin.[73]

The leaf extracts of *Achyranthes aspera*, along with other plant species, displayed varying levels of larvicidal activity against *Aedes aegypti* L. and *Culex quinquefasciatus* early fourth- instar larvae.^[74] In addition, the plant has exhibited significant antibacterial and antifungal properties in dried leaf extracts, particularly in chloroform, petroleum ether and methanol.^[75] Phytochemical analyses revealed that *Achyranthes aspera* extracts demonstrated the most prominent inhibition of *E. coli* growth.^[76] Additionally, the root extract was found to possess potent hormonal actions associated with insect molting, displaying high larvicidal activity against *Boophilis microplus* tick larvae.^[77]

Anti-inflammatory activity

"The alcohol extract of Achyranthes aspera exhibited significant anti-inflammatory effects in albino male rats, ^[78] as demonstrated in studies utilizing cotton pellet granuloma and carrageenin-induced hind paw oedema models.^[79] Additionally, the ethanolic extract of A. aspera demonstrated anti-inflammatory and anti-arthritic activities within the dosage range of 100-200 mg/kg.[80] Achyranthine, a water-soluble alkaloid found in A. aspera, was evaluated for its anti-inflammatory and antiarthritic properties in rats across various models,^[81] showing notable efficacy although not surpassing the performance of phenylbutazone and betamethasone.^[82] Moreover, achyranthine led to alterations in organ weights, including reductions in the thymus, spleen and adrenal glands,^[83] accompanied by elevated concentrations of ascorbic acid and cholesterol in the adrenal gland,^[84] similar to the effects of betamethasone.[85] All three medications under examination induced a decrease in food intake without significant effects on urination, mortality rate, or fecal output.^[86]

Notably, betamethasone exhibited a higher incidence of gastric ulcers compared to achyranthine.^[87] Furthermore, additional studies have corroborated the anti-inflammatory properties of *A. aspera*.^[88] In a chronic study, the alcohol extract of *A. aspera* demonstrated notable inhibition of edema in carrageenin-induced rat paw oedema.^[89] with 65.38% and 72.37% reductions observed at doses of 375 and 500 mg/kg, respectively.^[90]

In terms of immunomodulatory activity, the extract of Achyranthes aspera exhibited an enhanced induction of humoral antibody responses specific to Ovalbumin (OVA) in mice. This response displayed a dose-dependent trend, resulting in significantly elevated levels of IgM, IgG 1 and IgG 3 antibodies, albeit with a decrease in anti-OVA PCA titers. Notably, extracts from the seed and root of the plant demonstrated particularly robust activity.^[91] In studies involving fish, Achyranthes aspera seed supplementation led to notable enhancements in various immune parameters, including RNA/DNA ratios, hemagglutination antibody titters and anti-trypsin activity. Additionally, serum globulin levels were significantly elevated in the Achyranthes-treated group, indicating an augmentation of immune function in catla.^[92] Similar positive effects on immune parameters were observed in Labeo rohita, rohu fingerlings, where Achyranthes supplementation resulted in increased superoxide anion generation, lysozyme levels and serum bactericidal activity, among other markers. Furthermore, the extract showed potential in enhancing resistance to infection.^[93] Finally, it has been shown that hydroalcoholic extract of A. aspera enhances the cell-mediated immune system by enhancing phagocytic function.^[94]

Ant-ifertility activity

The ethanol extract of *Achyranthes aspera* root demonstrated potent antifertility activity when administered orally to fertile albino female rats during days 1-7 of pregnancy. At a dosage of 200 mg/kg body weight, it exhibited an impressive 83.3% anti-implantation activity, resulting in complete prevention of successful pregnancies.^[95] In addition, when tested in immature female albino mice that had undergone ovariectomy, the ethanolic extract showed estrogenic properties.^[96]

In a separate investigation, the acetone and methanolic extracts from the roots of *A. aspera* exhibited notable anti-implantation effects, with 50% and 60% inhibition observed, respectively, in mice.^[97] Additionally, a synthetic extract derived from the leaves and roots of *A. aspera* and *Stephania hernandifolia*, in a ratio of 1:3, showed that sperm had the ability to quickly immobilize within 2 min after administration at a concentration of 0.32 g/mL. This effect was irreversible, indicating spermicidal rather than spermiostatic properties.^[98] Moreover, the extract significantly reduced sperm viability, rendering them nonviable after 30 min. At the highest concentration, it also led to a notable decrease in hypo-osmotic swelling, suggesting potential damage to the sperm plasma membrane.^[99] The methanolic extract of *A. aspera* leaves was assessed for its impact on various indicators of antifertility activity in female rats. The extract induced significant increases in pituitary and uterine weights in ovarectomized rats, demonstrating abortifacient effects. However, it had no noticeable influence on serum levels of ovarian hormones or different lipid profiles, except for a reduction in HDL at the doses tested.^[100]

In a distinct trial, the benzene extract of *A. aspera* stem, when orally administered, resulted in 100% prevention of conception on post-coitum or day 1 in rats.^[101] The stem's crude benzene extracts also exhibited a strong abortifacient effect in mice.^[102] Additionally, the ethanolic extract (excluding the root) of the plant demonstrated a 60% antifertility effect in rats when administered orally at doses ranging from 100 to 200 mg/kg body weight. Subsequent tests further supported its potential.^[103]

The aerial fraction of n-butanol has demonstrated an anti-pregnancy effect in female rats when administered orally at doses of 75 mg/kg or more on days 1 to 5 postcoitus, although it was found to be ineffective in hamsters, even at higher doses.^[104] up to 300 mg/kg. The aqueous portion did not show an anti-fertility effect in hamsters or rats. In ovariectomized rats, the extract showed strong estrogenic activity, leading to noticeable stimulation of uterine weight even at a dosage as low as 3.75 mg/kg.^[105]

In another study, male rats fed a 50% ethanolic *Achyranthes aspera* extract showed no significant effects on sperm motility or HMG CoA reducing activity. However, this leads to decreased sperm count, epididymal weight, serum testosterone levels and 3beta-hydroxysteroid dehydrogenase activity in the testicles. Elevated urinary 17-ketosteroid concentrations, fecal bile acids, testicular liver cholesterol concentrations and increased incorporation of labeled acetate into cholesterol suggest reproductive toxicity, possibly through inhibition of synthesis protein Androgen.^[106]

Whole plant extracts exhibited abortifacient effects in mice, with the benzene extract demonstrating the highest activity, specifically by interfering with corpus luteum function in the ovaries. However, no such effects were observed in rats. Furthermore, co-administration of progesterone or pituitary extract did not prevent abortions in mice, indicating species-specific action.^[107] In rabbits, a benzene fraction of the extract demonstrated abortifacient effects at a single dose of 50 mg/kg.^[108] Additionally, oral administration of both ethanol and chloroform extracts of *A. aspera* exhibited estrogenic properties as well as a 100% anti-implantation effect.^[109]

The alkaloid fraction obtained from the alcoholic extract of the root bark inhibited the oxytocin response in isolated rat uterus. However, it did not affect uterine responses to histamine or serotonin in guinea pigs, nor to acetylcholine in rats.^[110] Benzene extract of the whole plant (excluding roots) also showed anti-implantation activity in female albino rats.^[111]

Anti-hyperlipidemic activity

The alcoholic extract derived from *Achyranthes aspera* demonstrated significant lipid-lowering effects in both hyperlipidemic and healthy rats. In rats with triton-induced hyperlipidemia, the extract led to notable decreased Phospholipid (PL), Triglyceride (TG), Total blood Cholesterol (TC) and Total Lipid (TL). Specifically, PL, TC, TL and TG% serum concentrations decreased by 62%, 56%, 67% and 68%, respectively. After chronic administration for 30 days. Furthermore, hepatic lipids also showed a significant decrease. This effect is attributed to the extract's ability to prompt rapid bile acid excretion, resulting in reduced cholesterol absorption, indicating a potential mechanism of action for its cholesterol-lowering properties.^[112]

Additionally, the hypolipidemic potential of *Achyranthes aspera* was assessed in rats fed with sesame oil, known to induce lipid peroxidation. The alcoholic extract, administered at a dose of 100 mg/kg, effectively reduced blood levels of triglycerides, phospholipids, total lipids and total cholesterol, highlighting its efficacy in mitigating lipid abnormalities.^[113]

Anti-feedent activity

The crude ethanolic extract of *A. aspera* was evaluated for its effectiveness against cauliflower borer and brinjal borer larvae. Detailed observations were made on the initial and final weights of the larvae, mortality rates and death rates. Notably, the extract led to a significant reduction in both food consumption and excreted feces. Interestingly, there was a sudden increase in the total body weight of the larvae by 600 μ g and 800 μ g. However, at the higher concentration of 800 μ g, the larvae exhibited signs of distress and unhealthy growth. By the third day, the larvae exposed to the 1000 μ g concentration had succumbed to the extract's effects. Furthermore, the amount of excreted feces was notably decreased at the highest concentration of 1000 μ g. In conclusion, the plant extract demonstrated substantial antifeedant properties and exhibited a lower level of larvicidal activity against *Spodopter litura*.^[114]

Anti-diabetic activity

The 50% ethanolic extract of the entire plant demonstrated hypoglycemic effects in rats during preliminary biological investigations.^[115] However, it showed no discernible impacts on respiration, Cardiovascular System (CVS), Central Nervous System (CNS), or isolated guinea pig ileum. Additionally, the extract did not exhibit anti-helminthic, anti-tumor, anti-protozoal, or anti-viral properties. The Maximum Tolerated Dose (MTD) of the extract in rats was determined to be 1000 mg/kg body. Weight when administered orally, in another study, a substantial dose-dependent hypoglycemic effect was observed with oral administration of 2-4 g/kg of whole plant powder in both normal and alloxan-induced diabetic rabbits. Additionally, water and methyl alcohol extracts of the plant reduced blood sugar

levels in both healthy rabbits and rabbits with alloxan-induced diabetes. $^{\left[116\right] }$

The hypoglycemic effect of the plant in diabetic rabbits may be due to the fact that it provides essential nutrients such as zinc, manganese, magnesium, calcium and copper to beta cells. Furthermore, administration of plant seeds to high-dose fructose-fed rats showed alterations in the redox and oxidation states of plasma and other tissues.^[117]

In experiments involving normoglycemic albino rats, diabetes was induced by intraperitoneal injection of a dose of 150 mg/kg body weight of alloxan monohydrate after a 12 hr fasting periods. This dose of alloxan resulted in sustained hyperglycemia after four days of monitoring blood and urine samples to determine glucose levels.^[118] The *A. aspera* water extract group (administered at a dose of 500 mg/kg) showed significantly lower blood glucose and HbA_{1c} levels than the control group.

To induce diabetes in adult Wistar rats, *Streptozotocin* was administered intravenously at a dose of 60 mg/kg body weight. This agent caused diabetes to develop within three days by damaging the beta cells.^[119] It was reported that an ethanol extract of *A. aspera* (administered at 600 mg/kg) significantly reduced blood glucose levels.

Diuretic activity

Mice were treated with 10 to 20 mg/kg intramuscularly (i.m) of the saponin derived from *A. aspera* seeds exhibited significantly increased urine production at 2, 6 and 24 hr compared to untreated rats. Notably, Mersalyl at a concentration of 3 mg/ kg also induced a diuretic effect of comparable magnitude. The optimal dosage for the saponin was determined to be 10 mg/kg. Furthermore, rats administered oral doses of the saponin (at 5-10 mg/kg) demonstrated a substantial elevation in urine output, akin to the effect observed with acetazolamide at 10 mg/kg. Importantly, the diuretic effects of the saponin were comparable to those of acetazolamide and were accompanied by an increase in the excretion of potassium and sodium in the urine.^[120]

Activity on Cardiovascular system

Incorporation of isolated saponins extracted from *A. aspera* seeds demonstrated a significant increase in contractile force in hearts isolated from guinea pigs, frogs and rabbits. Pronethalol effectively blocks the stimulant effects of lower doses of saponins (from 1 to 50 μ g), while mepyramine partially attenuated these effects. This resulted in enhanced contraction force of failing papillary muscle and heightened tone in hypodynamic hearts due to the presence of saponins. Notably, compared to the effects of digoxin, the onset of action of saponins was quicker and of shorter duration.^[120] Further investigations into the phosphorylase activity of perfused rat hearts revealed that saponin, much like adrenaline, stimulated this activity, indicating a comparable effect on cardiac function.^[121]

In an earlier study, the roots of *A. aspera* were found to induce a sudden reduction in blood pressure in anesthetized dogs, without notable effects on respiration. However, at higher doses, a slight depression in respiratory rate was observed. The hypotensive effects of the extracts could be counteracted with atropine sulfate. Additionally, the extracts exerted temporary negative chronotropic and ionotropic effects on the frog's heart. In addition, the extract increased both the intensity and amplitude of contractions in the uterus of pregnant and non- pregnant albino rats, guinea pigs and rabbits, as well as induced contractions in the ileum of an isolated rabbit. When administered orally to rabbits, the plant extract significantly increased urine output.^[122]

In anesthetized dogs, the water-soluble alkaloid achyranthine, found in *A. aspera*, is reported to increase respiratory rate and amplitude while lowering blood pressure, impairing cardiac function and dilating the heart. Blood vessels, another study showed that saponins derived from *A. aspera* seeds exhibited cardio stimulatory activity and increased contractile force in both isolated and intact hypokinetic hearts.^[123] However, it should be noted that cardiovascular toxicity has been reported from leaf decoctions. In tropical West Africa, this plant was observed to activate the cardiovascular system.^[124]

Analgesic and Anti-pyretic activities

Achyranthes aspera leaves exhibit both analgesic and antipyretic effects, equivalent to those of aspirin, at doses of 25 mg/kg for analgesic effects and 125 mg/kg for antipyretic effects. These effects were determined using the hot plate method and the yeast induction method.^[125] Another study corroborates the analgesic activity of *Achyranthes aspera* leaves and seeds.^[126] Specifically, when evaluated by the hot plate method and the acetic acid-induced torsion response, the leaves and seeds demonstrated analgesic effects in rats. Furthermore, hydroalcoholic extract obtained from the roots and leaves of *Achyranthes aspera* showed centrally acting analgesic effects in adult male albino rats, as evidenced by the results of tail flicking, hot plates and acetic acid-induced contraction of peripheral analgesic activity, aspirin. Is the standard drug.^[127]

Anti-carcinogenic activity

Scientists have explored the leaves of the *Achyranthes aspera* plant for their potential chemo preventive properties. In laboratory studies, methanol extract, alkaloid, non-alkaloid fractions and saponins were shown to effectively reduce early Epstein-Barr virus antigen activation by the tumor promoter 12-O-tetradecanoylphorbol-13-acetate in Raji cells (in concentration 100 µg). Notably, the alkaloid-free fraction, consisting mainly of nonpolar molecules, had the most significant inhibitory activity (96.9%; with a viability of 60 µL). Furthermore, in a two-step *in vivo* skin carcinogenesis assay in mice, the whole methanol extract demonstrated potent anticancer activity (76%).^[128]

Additionally, non-alkaloid constituents of the plant have been identified as potential anti- cancer agents, exhibiting inhibitory activity on human pancreatic cancer cells when extracted in methanol, highlighting these properties. Its anti-proliferative and anti-cancer effects.^[129] In experiments evaluating the anti-cancer activity of *A. aspera*, Swiss albino mice were intraperitoneally injected with mineral oil.^[130] While the whole methanol extract showed significant anti-cancer effects in *in vivo* studies using rat skin carcinogens, the alkaloid-free fraction, consisting only of non-polar molecules, demonstrated the strongest inhibitory effect in *in vitro* tests.

Renal Disorders

Achyranthes aspera shows an inhibitory effect on the mineralization of urinary stones, including calcium oxalate, calcium carbonate and calcium phosphate. Specifically, methanol extract demonstrated protective properties against lead-induced nephrotoxicity in albino rats.^[131] The plant root was evaluated for its effectiveness in inhibiting the nucleation and growth of calcium oxalate crystals *in vitro*, as well as protecting against oxalate-induced injury in NRK-52E (kidney epithelial model). (Mouse tube). Additionally, a hydroalcoholic extract of the plant has been studied for its effect in inhibiting the crystallization of calcium oxalate in synthetic urine, suggesting its potential as an anti-kidney stone drug.^[132]

Anti-Dandruff Activity

The efficacy of polyherb hair conditioner containing methanol leaf extract of *A. aspera* (PHO) was evaluated for anti-dandruff effectiveness. Clinical studies have demonstrated that coumarin; a component found in the crude extract of *Achyranthes aspera*, effectively reduces dandruff and inhibits the growth of Pityrosporum ovule.^[133]

Anti-Depressant Activity

Rats were subjected to doses of 200, 400 and 600 mg/kg of the methanolic extract derived from *A. aspera* and their total immobility time was recorded. Notably, the oral administration of the methanolic extract of *Achyranthes aspera* at a dosage of 600 mg/kg led to a significant reduction in immobility time, indicating its potential for producing antidepressant-like effects.

Miscellaneous

The effects of *Achyranthes aspera* leaf extract were studied on various parameters including body weight, Lipid Peroxidation (LPO), liver protein content, Catalase (CAT) activity, Superoxide Dismutase (SOD), as well as blood Thyroxine (T4), Triiodothyronine (T3) and glucose concentration. The extract demonstrated significant thyroid activity, elevating thyroid hormone levels, serum glucose levels, body weight and liver protein content. Interestingly, it did not cause any notable effects on the antioxidant enzymes SOD and CAT, suggesting its direct

free radical scavenging ability.^[134] The water-soluble alkaloid of the plant, achyranthine, isolated from it, has a spasmolytic effect on the rectal muscles of frogs and a cathartic and diuretic effect on albino rats. However, no significant effects were observed in isolated guinea pig, rat and rabbit ileum, as above the Central Nervous System (CNS), with only a slight antipyretic effect noted,^[135] Furthermore, experiments revealed that the leaf extract lacked anti-protozoal and antiviral properties and had no effects on Cardiovascular System (CVS), respiratory, central nervous system and membranes initiate preganglionic stimulation. The LD₅₀ of the extract in rats was found to be greater than 1000 mg per kg i.p.^[136] Additionally, fresh leaf extracts of *Achyranthes aspera* were assessed for their impact on Alternaria alternate, the pathogen causing *Vicia faba* leaf spot disease, resulting in observed growth inhibition.^[137]

The alkaloid fraction derived from the alcoholic extract of the root bark of *Achyranthes aspera* showed an inhibitory effect on the oxytocin response in isolated rat uterus. However, this fraction showed no inhibitory effect on the response to histamine in the guinea pig uterus, nor to serotonin and acetylcholine in the rat uterus.^[138] Additionally, the total chloroform-soluble fraction (alkaloid residue) of *Achyranthes aspera* has demonstrated mild antidiuretic activity in rats and has an antispasmodic effect against various spasmolytic agents on the myocardium. guinea pigs' intestines and intestines. No specific effects on the central nervous system were observed in rats and rats exposed to this fraction did not exhibit analgesic effects.

CONCLUSION

For centuries, medicinal plants have played an important role in humanity's survival. Knowledge about using plants to treat various diseases has been passed down from generation to generation. In modern times, a combination of indigenous wisdom and modern techniques are used to explore the medicinal properties, effectiveness and safety of plants.

Achyranthes aspera is one of the herbs widely used in Unani, Ayurvedic and Siddha systems of medicine to treat a variety of diseases. This herb acts as an astringent, diuretic and cleanser and is used in the treatment of conditions such as hemorrhoids, ascites and skin rashes. In addition, it is also used to treat snakebites, broken bones, respiratory problems and whooping cough. It also acts as a laxative in the treatment of leukemia and asthma. With a rich variety of chemical constituents including flavonoids, alkaloids, steroids, saponins and terpenoids, there is significant potential to develop innovative therapies and drugs to treat various diseases.

However, further exploration of the chemical composition of *Achyranthes aspera* is necessary as very little research has been done on this subject. The available botanical and analytical studies on this plant are rare. This provides many opportunities to conduct further research such as phytopharmacology of different

extracts, standardization of extracts, identification and isolation of specific compounds. These studies may be followed by the development of lead molecules, as well as their incorporation into specific botanical formulations for intended use.

AUTHOR CONTRIBUTION

NKJ and SA planned the review outline, content, and structure. NKJ, SA, PK, and SK wrote the manuscript and created the figures. AS, MB, DD, and SY contributed to the literature search and data collection. SY, HK, and SM created the figures and tables. All authors approved the manuscript for publication.

ACKNOWLEDGEMENT

The authors extend their appreciation to the authorities of ITM University for providing the necessary support and resources.

CONFLICT OF INTEREST

All authors declare that there is no conflict of interest.

ABBREVIATIONS

GC-MS: Gas Chromatography-Mass Spectrometry; **TCA:** Tricarboxylic Acid Cycle; **HDL:** High-Density Lipoprotein; **IgG:** Immunoglobulin G; **HMG-CoA:** 3-Hydroxy-3-Methyl-Glutaryl-Coenzyme A; **OAR:** Oxidation and Reduction; **CNS:** Central Nervous System; **CVS:** Cardiovascular System; **PL:** Phospholipids; **TG:** Triglycerides; **TL:** Total Lipids; **MTD:** Maximum Tolerated Dose; **HbA**_{1c}: Hemoglobin A1c; **NRK52E:** A specific cell line used in research; **Pho:** Phosphorylation; **LD**₅₀: Median Lethal Dose; **ED**₅₀: Effective Dose 50%; **LPO:** Lipid Peroxidation; **CAT:** Catalase; **SOD:** Superoxide Dismutase; **T3 and T4:** Triiodothyronine and Thyroxine (Hormones produced by the thyroid gland).

SUMMARY

Achyranthes aspera, belonging to the family Amaranthaceae. Native to Asia, including India, it is also found in various parts of Africa, Australia and the Americas. This robust, erect and often prickly herbaceous plant is characterized by its distinctive lance-shaped leaves, which are typically arranged oppositely along the stem.

The plant has a long history of traditional medicinal use in various cultures. It contains a rich array of bioactive compounds, including alkaloids, flavonoids, saponins and triterpenoids, which contribute to its therapeutic properties. In traditional medicine, *Achyranthes aspera* has been employed to treat a wide range of ailments, including inflammatory conditions, digestive disorders, respiratory issues and skin problems.

One of its notable uses is in traditional Ayurvedic medicine, where it is known as "Apamarga." It is valued for its diuretic,

analgesic and anti-inflammatory properties. Additionally, the plant has been investigated for its potential as an antimicrobial and anti- diabetic agent.

In modern times, *Achyranthes aspera* has attracted attention from the scientific community due to its pharmacological potential. Research has focused on various aspects, including its cancer-fighting, anti-oxidative, anti-microbial and anti-inflammatory qualities. Additionally, studies have explored its potential applications in the treatment of conditions like diabetes, hypertension and gastrointestinal disorders.

Furthermore, *Achyranthes aspera* is used in traditional agricultural practices. It is known to possess allelopathic properties, meaning it can release chemicals that inhibit the growth of competing plants. This characteristic has led to its use in organic farming as a natural herbicide.

REFERENCES

- Anand S, Chaudhuri A, Chopra N, Kataria U, Dhanorya D, Bajhaiya MK, et al. A Comprehensive Review of Therapeutical and Ethnobotanical Aspects, Phytoconstituent and Pharmacological Activity of Aesculus indicia. Pharmacog Res. 2023;16(2):2024
- Pandey MM, Rastogi S, Rawat AK. Indian herbal drug for general healthcare: an overview. Internet J Altern Med. 2008;6(1):3. doi: 10.5580/1c51.
- 3. Arun V, Liju VB, Reena JJV, Parthipan B, Renuka C. Indian J Trad Knowl. 2007;6(4):589-94.
- John D. One hundred useful raw drugs of the Kani tribes of Trivendrum forest division, Kerala, India. Int J Crude Drug Res. 1984;22(1):17-39. doi: 10.3109/138802 08409070646.
- 5. Pande PC, Tiwari L, Pande HC. Indian J Trad Knowl. 2007;6(3):444-58.
- 6. Jain JB, Kumane SC, Bhattacharya S. Indian J Trad Knowl. 2006;5(2):237-42.
- 7. Anonymous. The wealth of India raw materials. New Delhi: Council of Scientific and Industrial Research; 2005. p. 55-7.
- 8. Zafar R. Medicinal plants of India. CBS Publ Distributors. 2009:1-15.
- Paul D, De D, Ali KM, Chatterjee K, Nandi DK, Ghosh D. Comparative study on the spermicidal activity of organic solvent fractions from hydroethanolic extracts of Achyranthes aspera and Stephania hernandifolia in human and rat sperm. Contraception. 2010;81(4):355-61. doi: 10.1016/j.contraception.2009.09.001, PMID 20227555.
- Sutar NG, Sutar UN, Sharma YP, Shaikh IK, Kshirsagar SS. Biosci Biotechnol Res Asia. 2008;5(2):841-4.
- 11. Neogi NC, Garg RD, Rathor RS. Indian J Pharmacol. 1970;32(2):43-6.
- Srivastav S, Singh P, Mishra G, Jha KK, Khosa RL. Achyranthes aspera -An important medicinal plant: a review. J Nat Prod Plant Resour. 2011;1(1):1-4.
- Singh N, Mrinal PS, Gupta VK. A review on pharmacological aspects of Achyranthes aspera. IPCM. 2019; 3(4): 000188: 1-10. doi: 10.23880/ipcm-16000188.
- Dhanoriya D, Jain NK, Dhurandhar Y, et al. In vitro Anti-Urolithiasis Activity of the Hibiscus rosa-sinensis L. Leaves Hydro-Alcoholic Extract on Calcium Oxalate Crystallization. Community Pract. 2024;21(3):1101-1112.
- L. V. Asolkar, K.K. Kakkar, O. J Chakre; "Second supplement to Glossary of Indian medicinal plants with active principles"; I (A- K); Publication and Information Directorate (CSIR) New Delhi, 15-16; 1992.
- 16. Sharma V, Chaudhary U. An overview on indigenous knowledge of *Achyranthes aspera*. Crit Rev. 2015;2(1):7-19.
- 17. Gupta V, Krishna CM, Bansal P, Kumar S, Prasad GP, Ravi KD. Phytochemistry and pharmacological potential of *Achyranthes aspera* -A. review. Int J Ayurvedic Med. 2010;1(1):1.
- Goyal BR, Goyal RK, Mehta AA. PHCOG rev.: plant review phyto-pharmacology of Achyranthes aspera: a review. Pharmacogn Rev. 2007;1(1).
- 19. Saini S. Review on *Achyranthes aspera* L. international education and Research [journal]. 2016;2(2):84-6.
- Bhinde NK, Altekar WW, Tnvedi JC, Slieth UK. Potassium diuretics in the Ayurvedic system of medicine. J Postgrad Med. 1958;4:21-7.
- Basu NK, Singh HK, Aggarwal OP. A chemical investigation of Achyranthes aspera. J Proc Inst Chem. 1957a;29: 55-8.
- 22. Kapoor VK, Singh H. Isolation of Betain from *Achyranthes aspera*. Ind J Chem. 1966;4:461-3.

- Kapoor VK, Singh H. Investigation of Achyranthes aspera. Ind J Pharmacol. 1967;29:285-8.
- Kumar S, Singh J. P, Kumar S. Phytochemical screening of some plants of Manipur-I. J Econ Bot Phytochem. 1990;1:13-6.
- Misra TN, Singh RS, Pandey HS, Prasad C, Singh BP. Antifungal essential oil and a long chain alcohol from *Achyranthes aspera*. Phytochemistry. 1992;31(5):1811-2. doi: 10.1 016/0031-9422(92)83153-P.
- Misra TN, Singh RS, Pandey HS, Prasad C, Singh BP. Two long chain compounds from Achyranthes aspera. Phytochemistry. 1993;33(1):221-3. doi: 10.1016/0031-9422(93) 85427-S.
- Misra TN, Singh RS, Pandey HS, Prasad C, Singh S. Isolation and characterization of two new compounds from Achyranthes aspera. Ind J Chem. 1996;35B:637-9.
- Varadharaj V, Kuppan M. Identification and determination of bioactive phytochemical constituents from the hydro-alcoholic extract of *Achyranthes aspera* whole plant by gas chromatography-mass spectrometry analysis. Asian J Pharm Clin Res. 2015;8:125-9.
- 29. Pahuja M, Mehla J, Reeta KH, Joshi S, Gupta YK. Hydroalcoholic extract of *Achyranthes aspera* Linn. roots ameliorates hypoxic pulmonary hypertension in rats. J Ethnopharmacol. 2016;189:179-86. doi:10.1016/j.jep.2016.05.044
- Wal A, Wal P, Gupta D, Kumar A, Sharma P, Jain NK. Natural products: a rising star for treating primary dysmenorrhea? Tradit Med Res. 2022;7(6):55. doi:10.53388/ tmr20220410001
- 31. Ali M. Chemical investigation of Achyranthes aspera. Orient JClient. 1993;9:84-5.
- 32. Banerji A, Chintalwar GJ, Joshi NK, Chadha MS. Isolation of ecdysterone from Indian plants. Phytochemistry. 1971;10(9):2225-6. doi: 10.1016/S0031-9422(00)97227-3.
- Sinha SKP, Dogra JVV. A survey of plants of Bhagalpur and Santhal pargana for saponin, flavonoids and alkaloids. Int J Crude Drug Res. 2002;23:77-86.
- Pathak MK, Jha V, Jain NK, Shrivastava N, Yadav N, Purukayastha D. Review on peptidomimetics: a drug designing tool. Am J Pharm Res. 2015;5(12):3859-3867.
- 35. Gopalachari R, Dhar M. L. Chemical examination of the seeds of *Achyranthes aspera*. J Sci Ind Res. 1952;11B:209-10.
- Gopalachari R, Dhar M. L. Studies in the constitution of the saponin from the seeds of *Achyranthes aspera*: Part I - Identification of the sapogenin. J Sci Ind Res. 1958;17B:276-8.
- Khastgir H. N, Sengupta S. K, Sengupta P. The sapogenin from seeds of Achyranthes aspera. J Ind Chem Soc. 1958;35:693-4.
- Hariharan V, Rangaswamyi S. Structure of saponins A and B from the seeds of Achyranthes aspera. Phytochemistry. 1970;9(2);409-14. doi: 10.1016/S0031-9422(00))85154-7.
- Batta AK and, Rangaswami S (1973):. Angiospermae dicotlyedonae: Amaranthaceae etc. Crystalline chemical components of some vegetable drugs. Phytochemistry. 1973;12(1):214-6. doi: 10.1016/S0031-9422(00)84654-3.
- Wal P, Vig H, Wal A, et al. Role of Nutraceuticals and Physical Activity in Parkinson's Disease Risk and Lifestyle Management. Curr Aging Sci. 2023;16(3):170-187. doi:http://dx.doi.org/10.2174/1874609816666230515121717
- Daulatabad CD and, Ankalgi RF (1985):. Minor seed oils. EL fatty acid components of some seed oils. Fette Seifen Anstrichm. 1985;87:196-7 (Chem. Abstr. 1985; 103,:34909p).
- Joshi M. C, Sabnis S. D. A phytochemical study of South Gujarat forests plants with special reference to the medicinal and of ethnobotanical interest. Bull Med Ethnobot Res. 1989;10;61-82.
- Agrawal R. G, Pant P, Tewari L. C, Singh J, Pandey M. J, Tiwary D. N. Preliminary phytochemical screening of medicinal plants of hilly district of U.P. Bull Med Ethnobot Res. 1989;10:176-6.
- Sharma S. K, Vasudeva N, Ali M. A new aliphatic acid from Achyranthes aspera. roots. Indian Journal of Chemistry. 2009;48B:1164-9.
- Banerji A, Chadha MS. S. Insect moulting hormone from Achyranthes aspera. Phytochemistry. 1970;9(7):1671-3. doi: 10.1016/S0031-9422(00)85295-4.
- 46. Muhammed AA, Muhammed MR, Anjon KM, Tanvir M, Azizur R and Muhammed AQ (2005):. 3-Acetoxy- 6~benzoyloxyapangamide from *Achyranthes aspera*. Dhaka University Journal of Pharmaceutical Sciences. 2005;4:21.
- Olaf K, Ernst H, Martin GS, Josef R, Frans B, Erfem M, Dawit A and Asfaw D, et al. (2000): Three saponins, a steroid and a flavanol glycoside from Achyranthes aspera. Monatsjefte fur Chemie. 2000;131:195-204.
- V. Seshadri V, A.K. Batta AK, S. Rangaswami S. Structure of two new saponins from Achyranthes aspera. Indian J Chem. 1981; 20B:773-5.
- 49. Saini S. Review on Achyranthes aspera. International Education and Research [Journal]. 2016;2(2):84-6.
- Krishnaveni A, Thaakur SR. Pharmacognostical and preliminary phytochemical studies of *Achyranthes aspera* Linn. Ancient Science of Life. 2006;26(1-2):1-5. PMID 22557217.
- 51. Basu NK, et al., Indian J. Pharm.acol;32: 43.
- Kirtikar, K.R. and, Basu. B.D.;. "Indian Mmedicinal Pplants"; Dehra Dun, India: International Book Distributors - Dehra Dun India. 2nd Editioned. 1981;3:2065-9.
- Dr. S. Vedavathy S, V. Mrudula V and, A. Sudhakar A; "Tribal Medicines of Chittoor District, A.P. (India)". Tirupati: Herbal Folklore Research Centre, Tirupati. 1997;16:17.

- Jain NK, Tailang M, Kumar S, et al. Appraising the therapeutical potentials of Alchornea laxiflora (Benth.) Pax & K. Hoffm., an underexplored medicinal herb: A systematic review. Front Pharmacol. 2022;13:958453. doi:10.3389/fphar.2022.958453
- Bhatnagar L. S, Singh V. K, Pandey G. Medico-botanical studies on the flora of Ghaigaon forests, Gwalior, Madhya Pradesh. J Res Indian Med. 1973;8:67-100.
- Raj K. P. S, Patel M. R. Some medicinal plants of Cambay and its immediate vicinity and their uses in Indian indigenous system of medicine. Indian Drugs. 1978;15:145-52.
- Khanna K. K, Mudgal V, Shukla G, Srivastava P. K. Unreported ethno medicinal uses of plants as aphrodisiac from the folklores of Uttar Pradesh plains, India. Bull Bot Surv India. 1994;36:91-4.
- Singh V. K, Ali Z. A. Folk medicines of Aligarh (Uttar Pradesh), India. Fitoterapia. 1989;60:483-90.
- 59. Girach R. D, Aminuddin A, Khan S. A. Ethno medicinal uses of *Achyranthes aspera* in Orissa (India). Int J Pharmacog.n. 1992;30:113-5.
- Anis M, Iqbal M. Medicinal plantlore of Aligarh, India. Int J Pharmacog.n. 1994;32(1):59-64. doi: 10.3109/13880209409082973.
- Husain W, Siddiqui M. B. Ethnobotanical approach of North-Western U.P. Acta Bot Indica. 1987;15:94-7.
- 62. Reddy M. B, Reddy K. R, Reddy M. N. A survey of medicinal plants of Chenchu tribes of Andhra Pradesh, India. Int J Crude Drug Res. 1988;26(4):189-96. doi: 10.3109/138 80208809053918.
- 63. Pal D. C, Jain S. K. Notes on Lodha medicine in Midnapur district, W. B., India. Econ Bot. 1989;43(4):464-70. doi: 10.1007/BF02935920.
- John D. One hundred useful raw drugs of the Kani tribes of Trivendrum forest division, Kerala. India. Int J Crude Drug Res. 1984;22(1):17-39. doi: 10.3109/138802 08409070646.
- Elvanayagum Z. E, Gnavanendham S. G, Balakrishna K, Bhima R. R, Usman S. A. Survey of medicinal plants with anti snake venom activity in Chengalpattu district, Tamil Nadu, India. Fitoterapia. 1995;66:488-92.
- Sebastnian M. K, Bhandari M. M. Medico ethno botany of Mount Abu, Rajasthan, India. J Ethnopharmacol. 1984;12(2):223-30. doi: 10.1016/0378-8741(84)90050-3, PMID 6521495.
- 67. Singh V, Pandey R. P. Medicinal plant-lore of the tribals of eastern Rajasthan (India). J Eecon Ttax Bbot. 1980;1:137-47.
- Neogi N. C, Rathor R. S, Shreshtha A. D, Banerjee B. K. Studies on the anti inflammatory and antiarthritic activity of achyranthine. Indian J Pharmacol. 1969;1:37-47.
- Basu NK, Neogi NC and, Srivastava VP (1957b):. Biological investigations of Achyranthes aspera Lmn. and its constituent achyranthine. J Proc Inst Chem. 1957b;29:161-5.
- George M, Venkatraman PR and Pandalai KM (1947):. Investigation on plant antibiotics: Part II. A search for antibiotic substances in some Indian medicinal plants. J Sci Ind Res. 1947;68:42-6.
- K. SushilKumar S, G.D. Bagchi GD, M.P. Darokar MP. Antibacterial activity observed in the seeds of some coprophilous plants. Int J Pharmacog.n. 1997;35(3):179-84. doi: 1 0.1076/phbi.35.3.179.13293.
- Valsaraj R, Pushpangadan P, Smitt UW, Adsersen A, Nyman U. W andersen A, Nyman U. Antimicrobial screening of selected medicinal plants from India. J Ethnopharmacol. 1997;58(2):75-83. doi: 10.1016/s0378-8741(97)00085-8, PMID 9406894.
- 73. Saravanan P, Ramasamy V, Shivakumar T. Antimicrobial activity of leaf extracts of *Achyranthes aspera*. Asian Journal of Chemistry. 2008;20(1):823-5.
- 74. Thilagavathi G, Kannaian T. Application of Prickly chaff (*Achyranthes aspera.*) leaves as herbal antimicrobial finish for cotton fabric used in healthcare textiles. Natural Product Radiance. 2008;7(4):330-4.
- 75. Bagavan A, Rahuman A. A, Kamaraj C, Geetha K. Larvicidal activity of saponin from *Achyranthes aspera* against *Aedes aegypti* and *Culex quinquefasciatus* (Diptera: Culicidae). Parasitology Research. 2008;103(1):223-9.
- Perumal Sarnmy R, Ignacimuthu S and, Sen A (1998):. Screening of 34 mdian medicinal plants for antibacterial properties. J Ethnopharmacol. 1998;62(2):173-82. doi: 10.1016/s0378-8741(98)00057-9, PMID 9741889.
- Perumal Sarny R, Ignacimuthu S and Patric Raja D Perumal Samy R, Ignacimuthu S, Raja DP. Preliminary screening of ethnomedicinal plants from India. J Ethnopharmacol. 1999;66(2):235-40. doi: 10.1016/s0378-8741(99)00038-0, PMID 10433484.
- Meera P, Amta Dora P and, Karunyal Sameul J (1999):. Antibacterial effect of selected medicinal plants on the bacteria isolated from fruit juices. Geobios. 1999;26:17-23.
- Bisht LSB, Brindavanam NB and, Kimothic GP (1988).: Comparative study of herbal agents used for fumigation in relation to formalin. Ancient Sci Life. 1988;8(2):125-32. PMID 22557643.
- Bagavan, A, Rahuman, A.A, Kamaraj C, Geetha K and Kannappan. Larvicidal activity of saponin from *Achyranthes aspera* against *Aedes aegypti* and *Culex quinqaefasciatus* (Diptera: Culicidae). Parasitology Research. 2008;103(1):223-9. doi: 10.1007/ s00436-008-0962-z, PMID 18392726.
- Bharathi KN, Manikandan S, Suresh K, Lakshmi T. Antidiabetic and antihyperlipidemic activity of *Achyranthes aspera* (Linn.) in streptozotocin induced diabetic rats. Asian Pac J Trop Biomed. 2012;2(12):940-3. doi:10.1016/S2221-1691(12)60215-9
- Londonkar, R.L., Reddy, C., and Abhaykumar, K. (2011). Potential antibacterial and antifungal activity of *Achyranthes aspera*; 2011.

- Manjula M, Indira V, Dhasarathan P. *In vitro* Aaction of Coccinia Grand's against bacterial organisms. Asian J of Microbiology, Biotechnology and Environmental Sciences. 2009;11(2):317-20.
- Chungsamarnyart N, Jiyajinda S, Jangsawan W (1991). Larvicidal effect of plant crude extracts on the tropical cattle tick (*Boophilus microplus*). Kasetsart. 1991;J 25:80-9.
- Vetrichelvian T, Jegadeesan M. Effect of alcohol extract of Achyranthes aspera Linn. on acute and subacute inflammation. on acute and subacute inflammation. Phytother Res. 2003;17(1):77-9. doi: 10.1002/ptr.1070, PMID 12557252.
- Gokhale A. B, Damre A. S, Kulkami K. R, Saraf M. N. Preliminary evaluation of anti-inflammatory and anti-arthritic activity of *S. lappa, A. speciosa* and *A. aspera*. Phytomedicine. 2002;9(5):433-437. doi: 10.1078/09447110260571689, PMID 12222664.
- Wal P, Shukla V, Uzzaman Khan MM, Gaur K, Wal A, Jain NK. Medicinal Properties of Crotalaria burhia: A Review. Curr Tradit Med. 2024;10(3):90-100. doi:http://dx.doi.org /10.2174/2215083810666230428095559
- Iwalewa, E. O., et al. "Inflammation: the foundation of diseases and disorders. A review of phytomedicines of South African origin used to treat pain and inflammatory conditions.". African Journal of Biotechnology 6.25. (2007):6.25.
- Kumar SP, Sucheta S, Deepa VS, Selvamani P, Latha SP SK, S S, V SD, P S, S L (2008). Antioxidant activity in some selected Indian medicinal plants. African J of Biotechnology. 2008;7(12):1826-8. doi: 10.5897/AJB2008.000-5030.
- Vasudeva R. Y, Duddukuri G. R, Sunil B. G, Athota R. R. Immunomodulatory Activity of Achyranthes aspera on the Elicitation of Antigen-Specific Murine antibody Response. Pharm. Biol. 2002;40(3):175-8. doi: 10.1076/phbi.40.3.175.5831.
- 91. Rao Y. VRao Y V, Chakrabarti R. Stimulation of immunity in Indian major carp *Catla catla* with herbal feed ingredients. Fish Shellfish Immunol. 2005;18(4):327-34. doi: 10 .1016/j.fsi.2004.08.005, PMID 15561562.
- Vasudeva N, Sharma SK. K. Post-coital antifertility activity of Achyranthes aspera. root. J. EthanopharmacolJ2006; 107(2):179-81. doi: 10.1016/j.jep.2006.03.009, PMID 16725289.
- Vasudeva Rao Y, Das B. K, Jyotyrmayee P, Chakrabarti R. Effect of Achyranthes aspera on the immunity and survival of *Labeo rohita* infected with Aeromonas hydrophila. Fish Shellfish Immunol. 2006;20(3):263-73. doi: 10.1016/j.fsi.2005.04.006, PMID 15961319.
- Chakrabarti R, Vasudeva RY. Achyranthes aspera stimulates the immunity and enhances the antigen clearance in Catla catla. Int Immunopharmacol. 2006;6(5):782-90. doi: 10.1016/j.intimp.2005.11.020, PMID 16546709.
- Mali RG, Hundiwale JC, Gavit RS, Patil KS, Kulkarni MV (2006). Effect of Achyranthes aspera extract on phagocytosis by human neutrophils. J of Natural Remedies. 2006;6(2):115-9.
- Jain NK, Tailang M. Green synthesis of zinc oxide nanoparticles and their biomedical applications in cancer treatment: current status and future perspectives. Appl Nanosci. 2023;13:6605-6629. doi:10.1007/s13204-023-02946-8
- Nelson VK, Nuli MV, Mastanaiah J, et al. Reactive oxygen species mediated apoptotic death of colon cancer cells: therapeutic potential of plant derived alkaloids. Front Endocrinol (Lausanne). 2023;14:1201198. https://www.frontiersin.org/ articles/10.3389/fendo.2023.1201198
- Paul D, Bera S, Jana D, Maiti R, Ghosh D. In vitro determination of the contraceptive spermicidal activity of a composite extract of Achyranthes aspera and Stephania hernandifolia on human semen. Contraception. 2006;73(3):284-8. doi: 10.1016/j.cont raception.2005.07.014, PMID 16472572.
- Shibeshi W, Makonnen E, Zerihun L, Debella A. Effect of Achyranthes aspera L. on fetal abortion, uterine and pituitary weights, serum lipids and hormones. on fetal abortion, uterine and pituitary weights, serum lipids and hormones. Afr Health Sci. 2006;6(2):108-12. doi: 10.5555/afhs.2006.6.2.108, PMID 16916302.
- Jain NK, Roy R, Pathan HK, Sharma A, Ghosh S, Kumar S. Formulation and evaluation of polyherbal aqueous gel from Psidium guajava, Piper betel and *Glycerrhiza glabra* extract for mouth ulcer treatment. Res J Pharmacogn Phytochem. 2020;12(3):145. doi:10.5958/0975-4385.2020.00024.2
- Sunita P, Jahan I, Jahan N, Alam MS, Pasha MK. Evaluation of anti-inflammatory activity of methanolic extract of *Achyranthes aspera* leaves. J Pharm Bioallied Sci. 2015;7(4):328-31. doi:10.4103/0975-7406.168035
- Wal A, Wal P, Vig H, et al. Treatment of Parkinson's Disease: Current Treatments and Recent Therapeutic Developments. Curr Drug Discov Technol. Published online May 2023. doi:10.2174/1570163820666230512100340
- 103. Jain NK, Agrawal A, Kulkarni GT, Tailang M. Molecular docking study on phytoconstituents of traditional ayurvedic drug tulsi (*Ocimum sanctum* Linn.) against COVID-19 Mpro enzyme: an *in silico* study. Int J Pharm Pharm Sci. Published online 2022:44-50. doi:https://doi.org/10.22159/ijpps.2022v14i4.43181
- 104. Wadhwa V, Singh MM, Gupta DN, Singh C, Kamboj VP. M, Gupta D. N, et al. Contraceptive and hormonal properties of Achyranthes aspera in rats and hamsters. Contraceptive and hormonal properties of Achyranthes aspera in rats and hamsters. Planta Med. 1986;5(3):231-3. PMID 3529148.
- Sandhyakumary K, Boby R. G, Indira M. Impact of feeding ethanolic extracts of Achyranthes aspera. on reproductive functions in male rats. Indian J Exp Biol. 2002;40(11):1307-9.

- Pakrashi A, Bhattacharya N. Abortifacient principle of Achyranthes aspera Linn. Indian J Exp Biol. 1977;15(10):856-8. PMID 606650.
- Raju GS, Moghal MM. A comprehensive review on Achyranthes aspera Linn.: Ethnobotany, phytochemistry and pharmacological activities. Pharmacogn Rev. 2007;1(1):143-50. doi:10.4103/0973-7847.31971
- Vasudeva N, Sharma S. K. Estrogenic and pregnancy interceptory effects of *Achyranthes aspera*. root. African Journal of Traditional, Complementary and Alternative Medicines. 2007;4(1):7-11.
- 109. Khare CP. Indian Medicinal Plants: An Illustrated Dictionary. Springer; 2007.
- 110. Nadkarni AK, Nadkarni KM. Indian Materia Medica. Popular Prakashan Pvt. Ltd.; 1976.
- 111. Mathur R, Chauhan S, Saxena V, Shukla S and, Prakash AO. Anti-implantation activity of some indigenous plants in rats. J Jiwaji Univ (Sci Technol Med). 1983;9:37-46.
- Khanna A. K, Chander R, Singh C, Srivastava A. K, Kapoor N. K. Hypolipidemic activity of *Achyranthes aspera*. in normal and triton-induced hyperlipidemic rats. Indian J Exp Biol. 1992;30:128-30.
- Priya K, Krishnakumari S (2007). Phytochemical analysis of Achyranthes aspera and its activity on sesame oil induced lipid peroxidation. Ancient Science of Life. 2007;27(1):6-10. PMID 22557252.
- 114. Girija S, Valarmathy N. Antifeedant effect of Achyranthes aspera on cauliflower borer (Hellula undalis), fruit and leaf borer of cauliflower (Spodoptera litura) and Brinjal fruit borer (Leucinodes arbonalis). Biosciences Biotechnology Research Asia. 2008;5(2):663-72.
- 115. Dhar M. L, Dhar M. M, Dhawan B. N, Mehrotra B. N, Ray C. Screening of Indian plants for biological activity. Part I,. Indian J Exp Biol. 1968;6(4):232-47. PMID 5720682.
- Akhtar M. S, Iqbal J. Evaluation of hypoglycemic effects of Achyranthes aspera in normal and alloxan-diabetic rabbits. J. Ethnopharmacol. 1991;31(1):49-5149-57. doi: 10.1016/0378-8741(91)90143-2, PMID 2030593.
- 117. Malarvili T, Gomathi N. Effect of Achyranthes aspera see on redox and oxidative status in plasma and selected tissues of rats fed with high doses of fructose see on redox and oxidative status in plasma and selected tissues of rats fed with high doses of fructose. Biosciences Biotechnology Research Asia. 2009;6(2):659-64.
- Szkudelski T. The mechanism of alloxan and streptozotocin action in cells of the rat pancreas. Physiol Res. 2001;50(6):537-46. PMID 11829314.
- Karunanayake EH, Hearse DJ, Mellows G. The metabolic fat and elimination of streptozotocin. Biochemical Society Transaction. 1975;3(3):410-4. doi: 10.1042/bst0 030410.
- Gupta S, Verma S. C, Ram A. K, Tripathi R. M. Diuretic effect of the saponin of Achyranthes aspera (Apamarga). Ind J Pharmacol. 1972;4:208-14.
- Ram A. K, Bhagwat A. W, Gupta S. S. Effect of saponin of Achyranthes aspera on the phosphorylase activity of rat heart. Ind J Physiol Pharmacol. 1971;15:107-10.
- Gambhir S. S, Sanyal A. K, Chowdhury N. K. Pharmacological study of Achyranthes aspera. A preliminary report. Ind J Physiol Pharmacol. 1965;9:185-8.
- Oliver -Bever B (1982). Medicinal plants in tropical West Africa. I. Plants acting on the cardiovascular system. J Ethnopharmacol pp: 5. 1982; 5(1):1-72. doi: 10.1016/ 0378-8741(82)90021-6, PMID 7033667.
- 124. Jayakumar T, Sridhar MP, Bharath Prasad TR, Ilayaraja M, Govindasamy S, et al. (2009) Experimental studies of *Achyranthes aspera*; 2009.
- 125. Sutar N. G, Sutar U. N, Sharma Y. P, Shaikh I. K, Kshirsagar S. S. Phytochemical investigation and pharmacological screening of leaves of *Achyranthus aspera* Linn. as analgesic and antipyretic. Biosciences Biotechnology Research Asia. 2008;5(2):841-4.
- 126. Jain NK, Baghel KS. Selective cyclooxygenase-2 inhibitor etoricoxib attenuated hypoxic cancer milieu induced m2-polarization of macrophages and acquisition of pro-angiogenic and pro-invasive attributes. Res J Pharm Technol. 2019;12(12):5871-5877. doi:10.5958/0974-360X.2019.01018.7
- 127. Kumar H, Singh D, Kushwaha SKS, Gupta AK. Comparison of leaf and root extract of Achyranthes aspera for its analgesic activity. Der Pharmacia Lettre. 2009;1(2):193-8.
- 128. Chakraborty A, Brantner A, Mukainaka T, Nobukuni Y, Kuchide M, Konoshima T, Tokuda H, Nishino H, et al. Cancer chemo preventive activity of Achyranthes aspera leaves on Epstein-Barr virus activation and two stage mouse skin carcinogenesis. Cancer Lett. 2002;177(1):1-5. doi: 10.1016/s0304-3835(01)00766-2, PMID 11809524.
- 129. Subbarayan PR, Sarkar M, Impellizzeri S, Raymo F, Lokeshwar BL, Kumar P, et al. (2010) Anti-proliferative and anti-cancer properties of Achyranthes aspera: Specific inhibitory activity against pancreatic cancer cells. J Ethnopharmacol. 2010;131(1):78-82. doi: 10. 1016/j.jep.2010.06.002, PMID 20541002.
- Geetha P, Narayanan KR, Murugesan AG (2010). Screening the anti-cancerous efficacy of Achyranthes aspera. using animal model Swiss Albino mice. using animal model Swiss Albino mice. J Biomed Sci Res. 2010;2(4):231-5.
- Pareta SK, Patra KC, Harwansh R (2011). In vitro calcium oxalate crystallization inhibition by Achyranthes indica Linn. hydroalcoholic extract: an approach to anti-lithiasis. Int J Pharma Biol Scienc. 2011;2(1):432-7.
- 132. Anantha D. In vitro anti helminthic activity of aqueous and alcoholic extracts of Aerva lanata seeds and leaves. J Pharmaceut Sci Resc. 2010;2(5):317-21.
- Barua CC, Talukdar A, Begum SA, Buragohain B, Roy JD, et al. (2009) Antidepressant like effects of Achyranthes aspera. in animals models of depression. in animals models of depression. Pharmacology. 2009;2:587-94.

- Tahiliani P, Kar A. Achyranthes aspera elevates thyroid hormone levels and decrease a hepatic lipid peroxidation in male rats. J. EthanopharmacolJ Ethnopharmacol. 2000;71(3):527-32. doi: 10.1016/s0378-8741(00)00170-7, PMID 10940593.
- Basu N. K, Neogi N. C, Srivastava V. P. Biological investigation of Achyranthes aspera. and its constituent achyranthine. and its constituent achyranthine. J Proc Inst Chem. 1957;29:161-5.
- Aswal B. S, Goel A. K, Kulshrestha D. K, Mehrotra B. N, Patnaik G. K. Screening of Indian plants for biological activity: Part XV. Ind J Exp Biol. 1996;34(5):444-67. PMID 9063078.
- 137. Jain NK, Tailang M, Chandrasekaran B, et al. Integrating network pharmacology with molecular docking to rationalize the ethnomedicinal use of Alchornea laxiflora (Benth.) Pax & K. Hoffm. for efficient treatment of depression. Front Pharmacol. 2024;15:1290398. doi:10.3389/fphar.2024.1290398
- Singh A, Singh SK. Phytochemical investigation and evaluation of anti-inflammatory activity of *Achyranthes aspera* Linn. in rats. Indian J Exp Biol. 2013;51(9):741-5. PMID: 24151751.

Cite this article: Jain NK, Anand S, Keshri P, Chopra N, Bajhaiya MK, Harsha GSS, *et al.* A Comprehensive Review of Ethnomedicinal, Phytochemical and Pharmacological Activity Profile of *Achyranthes aspera*. Pharmacog Res. 2024;16(3):472-82.