Interaction between Anti Diabetic Drugs and Herbs: A Review

Ruchika Mourya*, Rajesh Sharma

School of Pharmacy, Devi Ahilya Vishwavidyalaya, Takshashila Parisar, Khandwa Road (Ring Road), Madhya Pradesh, INDIA.

ABSTRACT

Diabetes is a long-term medical disorder characterized by elevated blood glucose levels due to inadequate insulin use. It is caused by a pathogenic process that results in insulin insufficiency due to the death of β -cells. Uneven insulin secretion and use contribute to diabetes mellitus. Pancreatic beta cell loss is a primary cause of diabetes mellitus, which is associated with both insulin insufficiency and insulin resistance. Herbal medicines have been used for treating illness since ancient times, with over 800 plants having potential anti-diabetic properties. These plants have numerous pharmacological and therapeutic applications, including lowering blood glucose levels and enhancing beta-cell function. It is a common trend in Asian countries specifically in India that herbal medicine/s along with allopathic medicine/s is used. Herbal drug interactions can occur when conventional medications and herbal remedies are used together, leading to elevated toxicity or pharmacological effects. Xenobiotic substrates can affect the biological activity of xenobiotic substrates and other compounds, leading to increased oral bioavailability and decreased clearance and excretion. Cytochrome induction, triggered by AhR and PXR receptors, can improve the activity of intestinal and hepatic enzymes, affecting oral bioavailability and plasma concentration. The therapeutic benefit of herbal drugs that induce cytochrome induction is reduced when taken concurrently, hence in the present review interactions (pharmacodynamic and pharmacokinetic) of 27 plants having anti-diabetic property with oral anti-diabetic agents have studied.

Keywords: Anti diabetic effect, Cytochrome (cyp), Herbal drug interaction, *In vitro* study, *In vivo* study, Pharmacodynamic study, Pharmacokinetic study, Plant extract.

INTRODUCTION

Diabetes is a long-term medical disorder characterised by elevated blood glucose levels (hyperglycemia) brought on by inadequate insulin use or level. Elevated blood sugar levels are associated with organ damage and tissue breakdown.^[1] Diabetic Mellitus (DM) is caused by a pathogenic process that result in insulin insufficiency due to the death of β -cells Improper utilisation and metabolism of glucose results in insulin resistance. Uneven insulin secretion and use is a major contributor to diabetes mellitus.^[2] A WHO 2021 report states that between 1980 and 2014, there were 422 million more diabetic patients than there were in 1980. Between 2000 and 2016, the number of early deaths linked to diabetes increased by 5%.^[3] One of the primary causes of diabetes mellitus is the pancreatic beta cell loss. It is associated with both insulin insufficiency and insulin resistance. Insulin resistance may result from the down regulation of GLUT-4 in muscle and adipose



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Correspondence:

Ms. Ruchika Mourya

School of Pharmacy, Devi Ahilya Vishwavidyalaya, Eastern Ring Rd, DAVV Takshila Parisar, Indore-452001, Madhya Pradesh, INDIA. Email: mouryaruchika98@gmail.com

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tissue because it improves glucose absorption to these tissues. The main site of glucose utilisation is skeletal muscle. It is thought that oxidative stress contributes to the development of pancreatic-cell dysfunction in type 2 diabetes.^[4] Excessive formation of ROS destroys the beta cells in the pancreas and impacts cellular activity, leading to cell death in many organs and blood vessels. Insulin deficiency results from beta cell loss.^[5]

Since ancient times, herbal medicines have been utilised for treating illness. There are more than 800 plants with possible anti-diabetic properties. The natural remedies work to alleviate illnesses. There are numerous pharmacological and therapeutic applications for the phytochemicals that exist in plants, including polysaccharides, alkaloids, glycosides, lipids, terpenoids and steroids. The action of the plants' herbs lowers blood glucose levels and enhances beta-cell function (Table 1). Most synthetic medications are made from plants and herbs.^[6] Many plants have traditionally been used to treat DM because they have less adverse effects when used long term.^[7]

When a conventional medication and herbal remedy are used together, there may be positive or negative interactions between the two. These are known as herbal drug interactions. Drug interactions with herbal treatments may result in elevated toxicity or pharmacological effects.^[8] Pharmacokinetic interaction is the term for when cytochrome enzyme inhibition impacts biotransformation or pharmacokinetic factors such drug absorption, bioavailability, distribution, metabolism and elimination.^[9] The drug's clinical and therapeutic effects changed; this is known as pharmacodynamic interaction. These are two forms of herbal drug interactions: Pharmacokinetic interaction and pharmacodynamic interaction.^[10] Herbal drug interactions may occur due to a lack of knowledge about the pharmacological mechanisms of medicinal plants.^[11] Inhibition and induction of cytochrome have a role in herbal drug interaction processes. CYP450 enzymes are ham proteins that belong to a super family. Each one is identified by the letters CYP, followed by a number and a letter. Three of the 74 CYP gene families have been identified as being involved in drug metabolism in the human liver (CYP1, CYP2 and CYP3).^[12] By catalyzing the process, the cytochrome plays a critical role in drug metabolism. The cytochrome P450 enzyme has been responsible for the oxidative, per oxidative and reductive metabolic transformations of drugs and natural compounds.

The biological activity of xenobiotic substrates and other compounds can be affected when xenobiotic substrates impair CYP enzymatic activity.^[13] Because drug absorption and distribution increase while clearance and excretion decrease when cytochrome is inhibited, the pharmacokinetic properties of the drug are altered. This results in high oral bioavailability because drug absorption and distribution increase while clearance and excretion decrease. The accumulation of drugs is also a source of concern. This improves the drug's therapeutic impact, but co-administration of herbal drugs with drugs having a narrow therapeutic window can have substantial side effects.^[14] The activation of various receptors causes cytochrome induction. The AhR (Aryl hydrogen Receptor) and PXR (Pregnane Nuclear Receptor) receptors, which belong to the orphan nuclear receptors/steroids receptors super family, trigger CYP1A/B and CYP3A isozymes.^[9] Some xenobiotics improve the activity of intestinal and hepatic enzymes by enhancing mRNA transcription, resulting in higher enzyme levels than usual and speeding up drug metabolism. This has an impact on oral bioavailability as well as plasma concentration. The therapeutic benefit of herbal drugs that induce cytochrome induction is reduced when they are taken concurrently.[14]

Abelmoschus esculentus

Okra is the scientific name for *Abelmoschus esculentus*.^[15] The anti-diabetic action of the *A. esculentus* has been demonstrated through glycogenesis, delayed intestinal glucose diffusion, increased glucose adsorption capacity and pancreatic islet cell regeneration. In the study, all of these pathways were discovered to reduce the post-meal glucose level. The glucose level in

the blood is reduced by A. esculentus.^[16] The major chemical constituents of A. esculentus are flavonoids, which number seven in number. Rutin, hypersoside, hibifolin, isoqurectin, myricetin, quercetin and qurectin-3-o-robinobioside are among the compounds.^[17] In A. esculentus, β-sistostenol, oleanolic acid, myricetin and kaempferol have indeed been demonstrated to have anti-diabetic action. Myricetin has been discovered as an anti-diabetic compound which might be isolated from various plant components.^[15] The inhibitory concentrations of α -amylase and α -glucosidase are 125±2 µg/mL and 110±1 µg/ mL respectively. Metformin absorption from the small intestine is suppressed in an in vivo investigation of water-soluble okra extract with metformin. Dietary fibers are found in the highest concentration in okra, followed by carbs and protein. Metformin is entrapped by fibers, which inhibits absorption.^[18,19] It was indicated that metformin should not be used with okra in this case. Because it has the potential to lower metformin levels at the target location.

Phyllanthus emblica

In Hindi, Phyllanthus emblica is known as amla and in English, Indian gooseberry. Emblicanin A and Emblicanin B, pedunculagin and punigluconin are some of the primary tannins found in P. emblica. Gallic acids, amlaic acid, arginine, aspartic acid, astragallin, carotene, sitosterol, chebulagic acid, chebulic acid, chebulaginic acid, chebulinic acid, chebulinic acid, corilagic acid, corilagin, cysteine, ellagic acid, emblicol, kaempferol, leucodelphinidin^[20] Gallic acid, Ellagic acid, Estradiol, Sesamine, Kaempferol, Zeatin, Quercetin and Leucodelphinidin have all been identified as possible anti-diabetic substances in a computer simulation.^[21] Inhibition of diasaccharidase activity reduces glucose and sucrose absorption from the gut.^[22] P. emblica inhibits glycogenolysis and hepatic glucogenesis by increasing insulin secretion via pancreatic β -cell stimulation or by having an insulin-sensitizing action.^[23] In an *in vitro* investigation, aqueous extracts of P. emblica were found to inhibit cytochrome 450 isoforms CYP1A2 (IC₅₀ value=310.28±5.07µg/mL), CYP2C9 (IC₅₀ value=194.72±2.94 µg/mL), CYP2D6 (IC₅₀ value=589.52±14.32 $\mu g/mL),\ CYP2E1\ (IC_{_{50}}\ value=310.27\pm15.06\ \mu g/mL),\ CYP3A4$ $(IC_{50} value=325.54\pm7.44 \mu g/mL)$, the metabolisms of tolbutamide and metformin may affect due to inhibition of CYP2C9 and CYP3A4.^[24] The co-administration of metformin (200 mg/kg) with the herbal formulation of Nisha amlaki, a combination of turmeric and Indian gooseberry, increases C_{max}, AUC_t and AUC by 62.06%, 45.64% and 46.69%, respectively and decreases clearance and Vd by 109.6% and 119% in diabetic rats, according to Shengule et al.,^[25] but the AUC for glucose level reduces by 37.79% and decreases TC level by 25.7% in diabetic rats. The Nisha amlaki reduces metformin excretion by reducing metformin transportation by the Organic Action Transporter (OCT). This could be the cause of metformin pharmacokinetic parameter variation.

Andrographis paniculata

Kalmegh is a popular name for Andrographis paniculata. Kalmegh is a natural antidiabetic with insulin secretagogue action that has been shown to stimulate insulin secretion in vitro studies.[26] The HepG2 hepatoma cell line was used to investigate the effects of A. paniculata on cytochrome inhibition. The andrographolide (60 µM) reduced the CYP1A2 expression. CYP2D6 and CYP3A4 were both inhibited by andrographolide and 14 deoxy 11, 12 didehydro androgapholide.^[27] In treated human hepatocytes. The expression of CYP1A2, CYP2C9and CYP3A4 mRNAs was significantly reduced (>2-fold) by andrographolide and A. paniculata extract. The cytochrome inhibitory action of Andrographolide extracts in different solvents varies. The highest inhibitory activity shown in androgapholide ethanol and methanol extract IC₅₀ value 21.1±1.4 μg/mL and 27.8±1.9 μg/mL respectively for CYP3A4, IC₅₀ value 52.6±0.8 µg/mL and 87.8±2.6 µg/mL for CYP2D6, IC₅₀ value 41.3±3.5 µg/mL and 76.7±2.0 µg/mL for CYP2C9. In comparison to ethanol and methanol extract andrographolide aqueous and hexane extract had a modest inhibitory impact on CYP3A4, CYP2D6and CYP2C9. All solvent extractions have shown that CYP2C9-mediated tolbutamide 4 hydroxylation is Inhibited.^[28] The A. paniculata extract and andrographolide magnify the ethoxyresorufin-O-deethylation, methoxy resorufin-Odemethylation, diclofenac-4-hydroxylation and testosterone 6β -hydroxylation in rat liver. The andrographolide affects the pharmacokinetic of the tolbutamide, it lowers the plasma concentration of tolbutamide in serum and the A. paniculate extract reduces the AUC_{0-12h} by 18%. A. paniculate extract and andrographolide is a PXR and AhR activator, it increases the DNA binding activity of PXR and AhR which magnify the gene transcription and enzymatic activities of CYP1A1, CYP1A2, CYP2C6, CYP2C11, CYP3A1, CYP3A2. The increased action of CYP2C maximizes the metabolisms of tolbutamide. CYP isozymes and P-glycoprotein expression elicitation has been seen in A. paniculate extract (APE) treated rats. The co-administration of APE andrographolide with tolbutamide doesn't show any significant synergistic effect. They don't alter the blood glucose sugar.^[29] The APE inhibits the human liver microsomal CYP1A2, CYP2C, CYP3A4 cytochrome.^[30] As earlier shows that the andrographolide inhibits the CYPEA4. Co-administration of glyburide with andrographolide maximize the bioavailability and the pharmacokinetic parameters such as $C_{max_{i}}AUC_{0-n_{i}}AUC_{total_{i}}$ $t_{1/2}$ MRT. There is no change in T_{max} of glyburide, it indicate that andrographolide has no effect on rate of absorption of the glyburide. The tolbutamide and A. paniculate together shows a synergestic effect^[31] Co-medication of gliclazide with A. *paniculate* increases the C_{max} AUC, kd $t_{1/2}$ and bioavailability of gliclazide by 63.39%. The increase in pharmacokinetic parameter may be due to decreased metabolism of gliclazide via CYP2C9 and CYP3A4 enzyme inhibition by A. paniculate.^[32]

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Boswellia serrata

Salai guggal is the common name for Boswellia serrata (Family-Burseraceae). Boswellia serrata improves insulin sensitivity and reduces insulin resistance.^[33] Using baculovirus-infected cells and PHLM (pooled human liver microsome), the researchers have found that the 50 g/mL B. serrata extract has an 84% and 98% inhibitory impact on CYP2C9 and CYP3A414 respectively, with IC₅₀ values of 11 µg/mL and 1.4 µg/mL.^[34] Frankincense (10 g/mL), a Boswellia tree oleo gum resin, inhibits CYP1A2, CYP2D6, CYP2C8, CYP2C9, CYP2C19and CYP3A4. The CYP2C8, CYP2C9and CYP3A4 enzymes are inhibited by boswellic acid, KBA and AKBA, with IC₅₀ values in the 5-10 M range.^[35] In STZ induced diabetic rats, Samla S. and Veersham C.^[36] discovered that Boswellic Acid (BA) and extract of B. serrata (BSE) showed an increase in pharmacokinetic parameters such as C_{max} , AUC_{0-n} , AUC_{Total} , $T_{1/2}$ and MRT of glimepiride and a decrease in clearance and Vd of glimepiride, but no change in T_{max} of glimepiride. Glimepiride with BSE and glimepiride with BA reduced glucose levels by 52.95% and 53.04%, respectively. The BSE and BA with glimepiride lower triglyceride levels while increasing insulin levels in the blood. The BSE and BA may inhibit CYP2C9 and interact pharmacokinetically with glimepiride. In both normal and diabetic rats, boswellic acid had a somewhat stronger impact than B. serrata extract. In a pharmacokinetic study of metformin with boswellic acid, researchers discovered an increase in C_{max} , AUC_{0-n} , AUC_{Total} , $T_{1/2}$ and MRT, as well as a decrease in clearance and Vd in normal and diabetic rats. Because BA inhibits CYP3A4 in human liver microsomes and enhances metformin bioavailability, there is no effect on T_{max} and rate of absorption. After 28 days, BA and metformin lower GOT, GPT and glucose levels while increasing insulin levels in the blood.^[37]

Aloe barbadensis Miller

In India, Aloe barbadensis Miller is known as Aloe vera. Aloin, barbaloin, isobarbaloine, aloetic acid, aloe-emodin, emodin, cinnamic acid, crysophanic acid glucomannan, cellulose, mannose and glucosamines have antihyperglycemic action. Aloe vera raises insulin levels and lowers glucose absorption from the gut^[38] and inhibits the AGE formation.^[39] The two separate brands of Aloe vera juice A and B, respectively, with IC₅₀ values of 8.35±0.72 mg/mL and 22.4±5.4 mg/mL for CYP3A4 and 12.5±2.1 mg/mL and 43.0±2.0 mg/mL for CYP2D6.[40] The enzymes CYP3A4 and CYP2D6 may be inhibited by Aloe vera. The co-administration of glibenclamide and Aloe vera reduces glucose and lipid levels at modest doses within 2 to 4 weeks in a clinical investigation.^[41] The interaction of Aloe vera and other antihyperglycemic medications has potential. Because Aloe vera is a widely used herbal medicine, the interaction between the two should be investigated.

Bridelia ferrugenia

'Ira' is the common name for *B. ferrugenia* (family: *Euphorbiaceae*). B. ferrugenia increases insulin sensitivity in tissue and decreases glucose absorption from the gut via stimulating insulin secretion and glp-1.^[42] a-Amyrin acetate, cymene, β-Amyrin 4-Phenylbenzophenone, Lupenon, Lupeol acetate, 2, 3, 6-Trimethylhept-3-en-1-oland 2, 3,6-Trimethylhept-3-en-1-ol properties.^[43] have antihyperglycemic Metformin pharmacokinetics was affected when metformin and B. ferrugenia were given together. C_{max}, T_{max} and AUC have all dropped. It means that the bioavailability of metformin in the body is reduced. Vd grew as the absorption rate constant (Ka) increased. Metformin's elimination constant, clearanceand T_{1/2} (half-life time) all increased. It is thought that B. ferrugenia reduces the impact of metformin via increasing metformin elimination.^[44] To corroborate this research, coadministration of metformin (250 mg/kg p.o. and 1000 mg/kg p.o.) with B. ferrugenia (30 mg/kg p.o. and 300 mg/kg p.o.) had a lesser antihyperglycemic impact in rats than *B. ferrugenia* (30 mg/kg and 300 mg/kg p.o.).^[45]

Cassia ariculata (L)

Cassia ariculata (L), popularly known as avaram senna, is a plant in the Fabaceaceae (Leguminosae) family. Qurecitin, cyanidin, spermidine and N8-acetylspermidine, hesperidine and nitexin-2-o-rhamnoside are phytochemicals found in C. ariculata that have anti-diabetic properties.[46] C. ariculata increases insulin levels in the blood and glycogen production, stimulates beta cell insulin release and regenerates them^[47] and inhibits AGE formation.^[48] With IC₅₀ values of 28.5 μ g/mL, 165.5 \pm 7.50 μ g/mL and 158.8±9.62 µg/mL, Cassia alata inhibits CYP1A2, CYP2D6 and CYP3A4 correspondingly.^[49] Metformin (90 mg/kg) and C. ariculata have a synergistic impact for glucose reducing action, while metformin (45 mg/kg) and C. ariculata had a similar effect to metformin alone. Metformin (90 mg/kg) enhanced the C_{max} by 11.46% and the AUC₀₋₂₄ by 4.6% in *C. ariculata*. Metformin's half-life was doubled, however T_{max} stayed the same. The half-dose of metformin (45 mg/kg) with C ariculata, on the other hand, resulted in a 17.72% reduction in $\mathrm{C}_{_{\mathrm{max}}}$ and a 42.42% reduction in AUC_{0.24}.^[50] The pharmacokinetic and pharmacodynamic interaction of a medicine may be affected by its dose. Therefore, dose determination should be studied in order to achieve a favorable therapeutic impact.

Catharanthus roseus

Periwinkle is a common name for *Catharanthus roseus* (family: *apocynacea*).^[51] *In vivo* and *in vitro* studies, vindoline,^[52] vincristine and vinblastine have significant anti diabetic and anticancer effects.^[53] The *C. roseus* maximize the glucose utilization via escalate the GLUT-2 and GLUT-4 gene expression in muscle tissue and alleviate the blood glucose level^[51] Augment

the insulin secretion from beta cells.^[54] Vindoline, an alkaloid found in *C. roseus*, is an inhibitor of CYP2D6 (IC₅₀ value 15.9 M) and CYP3A4 (IC₅₀ value 20.1 M). Ajmalicine is a potent reversible inhibitor of CYPD26. The methanol extract of *C. roseus* has been shown to inhibit CYP2D6 with an IC₅₀ of 11 g/mL.^[55] *C. roseus* can impact the pharmacokinetics and pharmacodynamics of drugs metabolised by CYP3A4 and CYP2D6, such as metformin, sitagliptin and sexagliptine. According to Ohadoma and Michael's research,^[56] *C. roseus* (250 mg/kg) plus metformin (100 mg/kg) exhibited a 64.86% drop in blood glucose levels when taken together. This occurred because of *C. roseus* inhibiting CYP3A6, which increased metformin bioavailability. However, when glibenclamide was combined with *C. rosea*, there was no significant difference in blood glucose levels compared to the normal medicine.

Table 1: Mechanisms of action of herbs.

| Mechanisms of action | Plants |
|--|--|
| Inhibition of AGE formation | Aloe barbadensis miller, Cassia ariculata. |
| Increase GLP secretion | Coctus pictus, Bridelia ferrugenia, Cinnamomum verum. |
| Regeneration of pancreatic cells | Cassia ariculata, Zingiberis offinalis roscoe, Glycyrrhiza glabra L. |
| Glycogen synthesis | Cassia ariculata, Cinnamomum verum, Curcuma longa, Momordica charantia. |
| Increase insulin secretion | Bridelia ferrugenia, Phyllanthus emblica, Andrographis paniculate, Cassia aricuata, Catharanthus rosea, Coctus pictus, Trigonella foenum graecum, Allium sativum L., Momordica charantia, Ginkgo biloba L., Gymnema sylvestre, Azadirachta indica. |
| decrease glucose absorption | Aloe barbadensis miller, Abelmoschus esculentus, Phyllanthus emblica, Bridelia ferrugenia, Cinnamomum verum, Trigonella foenum graecum, Tinospora cardiofolia, Opuntia spp., Gymnema sylvestre, Azadirachta indica, Ginkgo biloba L. |
| Regulation of glycolysis and Krebs cycle | Tinospora cordifolia, Syzigium cumini. |
| Insulin resistance | Boswellia serrata, Cinnamomum verum, Costus pictus, Curcuma longa, Trigonella foenum graecum, Allium sativum L., Zingibereris officinale roscoe, Gingko biloba, Opuntia spp., Gymnema sylvestre, Syzigium cumini, Azadirachta indica. |
| Inhibition of gluconeogenesis | Phyllantus emblica, Cinnamomum verum, Panax ginseng. |

Cinnamomum verum

Dalchini is the Indian name for Cinnamomum verum (family: lauraceace). C. verum is widely used as condiment all over the world. Cinnamaldehyde, eugenol, caryophyllene, cinnamyl acetate and -humulene are the primary phytochemicals.^[57] According to a placebo clinical study, C. verum increases GLUT 4 translocation and PPAR gene expression, improves insulin sensitivity and raises GLP-1 levels, while inhibiting glucose absorption through α - glucosidase inhibition and increasing glucose metabolism through stimulation of glycogen synthesis and inhibiting gluconeogenesis.^[58] C. verum suppresses the activity of the CYP2D1 enzyme in both type 1 and type 2 diabetes mellitus.^[59] An herbal-drug interaction may occur when a drug is processed by the CYP2D enzyme.^[60] The co-administration of metformin with C. verum in low and high doses improved G6PDH activity in diabetic rats, but that there was no significant difference in glucose reduction between the metformin-treated group and the metformin+cinnamon with low and high dose treated group. In comparison to the metformin-only group, the co-administration of metformin and C. verum improved HDL cholesterol levels in the blood. C. verum and metformin have been found to have a synergistic effect on cholesterol lowering.^[61]

Costus pictus

Spiral flag and step ladder are popular names for Costus pictus (family: Costaceae)[62] belongs to Central America and is known in South India as the insulin plant.^[63] The major phytochemicals present in C. pictus are isoquercetin, astragalin, kaempferol, quercetin, isovitexin, naringenin, galanin, genistin, licochalcone A, 2, 5-dihydroxy benzoic acid, gentisic acid, o-coumaric, melilotic, a-resorcyclic acid, 3,5-dihydroxy benzoic acid, p-hydroxy benzoic acid, cis and trans-p-coumaric acid.^[62] Insulin secretory action has been demonstrated in vitro and in vivo in several investigations and it is utilized as a herbal medication. Through activation of the incretin hormone GLP-1 release, the insulin plant reduces insulin resistance, enhances pancreatic insulin secretion and lowers postprandial blood glucose levels.^[63] The combination of metformin and the methanol extract of *C*. pictus (400 mg/kg) have a synergistic effect on the fasting blood sugar and lipid profile of diabetic rats. The extract and metformin have been shown to help with renal and liver problems.^[64] The herbal medication interaction of C. pictus with anti- diabetic drugs should be investigated in the future.

Curcuma longa

The rhizomes of *Curcuma longa (zingiberaceae*) are known as turmeric in English and haldi in Hindi. Curcumin, bisdemethoxycurcumin and demethoxycurcumin, as well as tumerone, atlantone and zingiberene, are diarylheptanoids found in curcuma species.^[65] *C. longa* increases hepatic glucose synthesis, GLUT2, GLUT3and GLUT4 gene expression, glucose uptake in tissue, activates AMPK and lowers insulin resistance by increasing hepatic glucose production, GLUT2, GLUT3and GLUT4 gene expression and decreasing insulin resistance.^[66] Both dexomethoxycurcumin and curcumin have been proven to inhibit CYP3A4, CYP1A2, CYP2C9 and CYP2D6. Demethoxycurcumin inhibited CYP3A4 (11.1±1.6 µM), CYP1A2 (1.4±0.2 µM), CYP2C9 (36.7±2.1 µM) and CYP2D6 (34.0±14.2 µM), while curcumin inhibited CYP3A4 (54.4±18.3 µM), CYP1A2 (6.0±1.4 μM), CYP2C9 (175.0±47.0 μM) and CYP2D6 (104.6±22.1 μM). In comparison to curcumin, demethoxycurcumin has a stronger inhibitory effect.^[67,68] Curcumin inhibited the production of p-gp in the intestine^[69] UGT and SLUT activities were suppressed by the curcuminoid extract.^[70] The curcuminoids may influence the enzymes that metabolize the drugs, resulting in a herbal drug interaction. In diabetic rats, the combination of curcumin from curcuma species plus glyburide, a sulfonyl urea derivative, had a considerable impact on blood glucose levels and lipid profiles. The bioavailability of glyburide is reduced by Pgp effluxand curcumin is a powerful Pgp inhibitor that increases glyburide bioavailability. The kinetics of glyburide is affected by curcumin. In contrast to the clearance, the AUC has increase.^[71] In diabetic rats, the combination of curcumin with glimepiride influences the pharmacokinetic characteristics of glimepiride, with an increase in C_{max} , AUC_{0-n} , AUC_{total} , $t_{1/2}$, MRT and a decrease in clearance and volume of distribution. This could be due to suppression of the CYP2C9 enzyme, but there was no change in T_{max} , indicating that the rate of glimepiride absorption has not changed.^[72] Curcumin (30 mg/kg) combined with metformin (50 mg/kg) resulted in a maximum decline in blood glucose levels and a positive rise in serum insulin levels. Undale V.R. et al.,[66] have observed beta cell reincarnation.

Trigonella foenum graecum L

Fenugreek (Trigonella foenum graecum) belongs to the Leguminosae family. Fenugreek seed contains mainly mucilaginous fiber (galactomannans), alkaloids mostly trigonelline, choline, gentianine and carpaine. Flavonoids are apigenin, luteolin, orientin, quercetin, vitexin and isovitexin. Saponins are diosgenin, yamogenin, tigogenin, neotigogenin, cholesterol and sitosterol. Other constituents are coumarin, fenugreekine, nicotinic acid, phytic acid, scopoletin, vitamins A, B1, C and nicotinic acid. GLUT4 translocation reduces glucose absorption and aberrant lipid metabolisms via inducing the insulin signaling pathway.^[73] The Trigonella foenum graecum extract had greater inhibitory activity on CYP3A4, CYP2D6 than trigonelline in cytochrome P450-carbon monoxide complex assay and fluorogenic assay. There are dose-dependent effects.^[74,75] The CYP2C11 activity was decreased by 43% at a high dose of fenugreek (600 mg/kg) and the protein expression was reduced by 80%. At a high dose, CYP2C9 was suppressed by 50%.^[76] The blood glucose level is reduced when fenugreek (1 g/kg) is combined with glimepiride (4 mg/kg) and insulin (4 u/kg). After 6 to 8 weeks of administration, they have a synergistic effect.^[73] Researchers discovered that

when metformin 300 mg/kg was compared to 500 mg/kg using non-compartment pharmacokinetic analysis, the drug's C_{max} and AUC increased by 74.68% and 148.55%, respectively. Vd and clearance fall, but metformin's T_{max} rises from 1 to 2 hr. The shift in T_{max} is attributable to a decrease in the drug's solubility. Metformin's bioavailability is increased by fenugreeks.^[76,77] After coadministration of aqueous extract of fenugreek and gliclazide, blood glucose levels in normal and diabetic rabbits were lowered by 56.96% and 45.81%, respectively. After coadministration of aqueous extract of fenugreek and gliclazide, blood glucose levels in normal and diabetic rabbits were lowered by 56.96% and 45.81%, respectively. The pharmacokinetic parameter of gliclazide has not been changed by fenugreek.[78] In clinical research conducted in China, the combination of fenugreek (0.35 g/pill) and sulfonyl urea reduced blood glucose levels by 80.43% more than the control group (43.48%) and the CSQS and HbA1c levels in diabetes patients decreased dramatically.^[79]

Allium sativum L.

Garlic is the common name for the Allium sativum L. (Alliaceae) plant. The major biological active constituents of A. sativum are Alliin, allicin, allixin, adenosine, Allyl1,5-hexadienyl trisulphide, Allyl methyl trisulphide, s-allyl2-pro pene thiosul phinate, ajoene, Diallyl disulfide, 1,5-hexa dianyl trisulfide, Methyl allyl trisulfide, 2-vinyl 1,3-dithiene, 3-vinyl 1,3-dithiene, S-allyl merc aptocysteine, Se-methyl selenocysteine, Allyl propyl disulfide, Sodium 2-propenyl thiosulphinate, s-methyl1-cysteine sulfoxide.[80] Garlic improves insulin sensitivity and production from beta cells, whereas allicin protects insulin from the SH group reaction and prevents insulin inactivation.[81] Several researches on various allium products in various solvents have been conducted. In an in vitro investigation, garlic products or extracts block CYP3A, CYP2A and Pgp trafficking.^[82] In a clinical trial, diabetic volunteers who ingested 10 g of raw garlic each day for 42 days had their blood cholesterol levels, FBS and HB1C levels drop dramatically, while their HDL-C levels raise significantly.^[83] Garlic raises intestinal Pgp expression by 131% and decreases the bioavailability of Pgp substrate saquiravair over 21 days in a clinical investigation. On CYP3A4, no effect was detected.^[84] Garlic 100 µg/mL decreased CYP2C9 activity but not CYP3A4 activity on the fourth day of therapy.^[85] Garlic oil helps with blood sugar control, diabetic nephropathy and blood pressure.^[86] The co-administration of metformin with A. sativum the AUC, 12 and C_{max} increases the bioavailability of the drug.^[81] In diabetic rats, co-administration of aqueous extract of A. sativum (500 mg/ kg) with metformin (50 mg) resulted in a greater reduction in blood glucose levels than metformin (100 mg).[87] Giving T2DM patients garlic tablets (300 mg) with metformin (500 mg) for 24 weeks resulted in a significant reduction in fasting blood glucose and total cholesterol levels.^[88] In STZ-induced diabetic rats, orally co-administration of aqueous extract of garlic (500 mg/kg) with glibenclamide (0.5 mg/kg) lowered blood glucose levels by 55.5%. It also helped diabetic rats lose weight, according to Tripathi P. *et al.*^[89]

Tinospora cordifolia

Tinospora cordifolia (family: Menispermaceae) is known in Sanskrit as amrita and in Hindi as giloy. Phytochemicals of giloy are tinosporine, cardiofolide, tinosporide, cardifole, columbin, barberin.^[90] magnoflorine, tinosporicide, manispermacide, tiniosinen.^[91] Amrita inhibits the synthesis of cholesterol and glycolysis. Inhibition of the DPP 4 enzyme enhances glucose transport and decreases carbohydrate digestion and absorption.^[90] T. cardifolia has been demonstrated to have an inhibitory effect on the CYP3A4, CYP2D6and CYP1A2 isoenzymes at high concentrations in DMSO and ethanol (5%), but no interaction was identified at low concentrations (0.1%), indicating that the effect is concentration dependant. CYP3A4 (IC₅₀=136.45 g/mL), CYP2D9 (144.37 g/mL), CYP29 (127.55 g/mL)and CYP1A2 (141.82 g/mL) have been demonstrated to be inhibited by T. cardifolia constituents and extracts, but the inhibitory impact is smaller than the positive control group.^[92] T. cardifoliaextract (400 mg/kg) combined with metformin (90 mg/kg), sitagliptin (10 mg/kg)and glibenclamide (1 mg/kg) lowered FBS levels to 148.8, 147.4and 188.5 mg/dL, respectively, with no hypoglycemia impact observed after 28 days. There was no interaction between metformin and sitagliptin pharmacokinetic parameters; however, T. cardifolia increases the C_{max} and AUC of glibenclamide by 1.2 times, a clinically insignificant impact. T. cardifolia combined with the above medicine lowered TC, TG and BUN levels in diabetic rats.^[93] The C_{max}, AUC and bioavailability of glibenclamide (1 mg/ kg) with T. cardifolia extract (400 mg/kg) increased at high dose, but not $\rm T_{_{1/2}}$ of gliben clamide significantly, $\rm T_{_{max}}$ increased from 3.5 hr to 6.5 hr significantly decreased in clearance at dose of 400 mg T. cardifolia extract with 1 mg/kg glibenclamide, according to Sahu R et al.^[94] T. cardifolia hydroalcoholic extract (100 mg/kg) plus glimepiride (20 mg/kg) Glimepiride's C_{max}, AUC_{0-t} and MRT_{0-t} all increased dramatically, but Vd and clearance decreased.[95]

Zingiberis officinale Roscoe

Zingiberis officinalis Roscoe (Zingiberaceae) is an herb that has been used to treat a variety of ailments. Zingiberis officinalis Roscoe is a plant that contains mostly gingerols and is often referred to as ginger. [6]-Gingerol, [8]-gingerol and [10]-gingerol are the most common gingerols. Methylgingerol and gingerdiol, dehydrogingerdione, [10]-dehydrogingerdione, gingerdiones, diarylheptanoids, diterpenlactones and galanolactone are some of the other gingerols. Gingerol has a pancreatic beta cell protective action, promotes glucose consumption in tissue and maintains blood glucose homeostasis by increasing insulin release and sensitivity and decreasing blood glucose levels.^[96] The ginger extract ethyl extract fraction inhibits the CYP2A6 and CYP2A13 enzymes with IC_{50} of $1.80\pm0.07 \mu g/mL$ and $11.81\pm0.18 \mu g/mL$, respectively.^[97] The gingerols in ginger suppressed the

activity of CYP2C9, CYP2C19, CYP3A4and CYP2D6. The 8-gingerol showed the most powerful inhibitory activity for CYP2C9 (6.8 µmol/l), CYP2C19 (12.5 µmol/l)and CYP2D6 (42.7 µmol/l).^[98] The 6-shogal a chemical constituent present in the ginger, inhibits the CYP2C9 (29.20 µM), CYP2C19 (18.78 μM), CYP2E1 (99.58 μM) CYP2E1 (99.58 μM).^[99] In vitro studies showed that 6-gingerol had a substantial inhibitory impact on CYP2C19 (IC₅₀ 36 μ M) and CYP1A2 (IC₅₀ 51 μ M), but a modest inhibitory effect on CYP3A4 (IC $_{50}$ 108µ M), CYP2D6 (IC $_{50}$ 235 μ M)and CYP2E1 (IC₅₀ 104 μ M).^[100] When ginger extract (4 mL/ kg) and sitagliptin (20 mg/kg) were given simultaneously, there was no interaction.^[101] Concomitant administration of (5 mg/ kg) glibenclamide and ginger extract at doses of 25 mg/kg and 50 mg/kg lowered blood glucose levels, but the higher doses of 100 mg/kg of extract raised blood glucose levels. In diabetic rats, ginger extract (50 mg/kg) reduced blood glucose level more than insulin.^[102]

Ginkgo biloba L.

Ginkgo biloba L. (family: Ginkgoaceae) is a living fossil that is used in traditional Chinese medicine. Flavone glycosides, ginkolides and bilobalides, rutin, quercitrin and hyperosid are the bioactive components of G. Biloba.^[103] (ginkgolides A, B, C and J and bilobalide) kaempferol, quercetin, apigenin, myricetin, tamarixetin, which yielded IC50 values for CYP1A2 or CYP3A of less than 10 µg/mL.^[104] The extract of G. biloba stimulates beta cell secretion and lowers insulin resistance.[105] G. biloba extract inhibits CYP2C9((K=14±4 µg/mL), CYP1A2 (K=106±24 µg/ mL), CYP2E1(K₁=127±42 µg/mL)and CYP3A4 (K₁=155±43 µg/mL), flavonoidic fraction of EGb 761 inhibited CYP2C9 (K_i=40±12 µg/mL), CYP2C9 (K_i=4.9±0.6 µg/mL), CYP3A4 (K_i=43±9 µg/mL)and CYP2E1 (K_i=55±11 µg/mL). And terpenoid fraction EGb 761 inhibited only CYP2C9 (K=15±6 µg/mL).^[106] A clinical study done by Geaorge B Kudolo,^[105] the G biloba extract (EGb 761, 120 mg for 3 month) have been reduced the glucose and insulin level in NIDDM subjects with pancreatic exhaustion (FBG 152±46 mg/dL, FPI 16±8 µU/mL) with oral hypoglycemic treatment. The G. biloba augmented the hepatic metabolic clearance rate of insulin and hypoglycemic agents; it may be anti-hyperinsulinemia in nature. The G. biloba extract inhibited CYP activity in small intestine (IC₅₀=50 μ g/mL) and in liver (IC₅₀=182±13µg/mL).^[107] In comparison to alpha amylase inhibitory effect, ethanolic and aqueous extracts (50 g/ mL) had a high alpha glucosidase inhibitory impact.^[108] After taking tolbutamide and GBE together, healthy volunteers' blood glucose levels dropped by 16%. There was no significant change in C_{max} and $AUC_{0-\infty}$.^[109] In a double-blind, placebo-controlled, cross-over study, co-administration of 120 mg of EGb 761 with metformin (500 mg) had no significant effect on metformin pharmacokinetic parameters in diabetic patients, but the highest dose of extract (9850 mg) had a significant effect on AUC and metformin excretion in 4 days.^[110]

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Panax ginseng

Asian or Korean red ginseng is the common name for Panax ginseng.[111] The main component of P. ginseng is ginsenoside, which is divided into two groups: protopanaxatriol (Re, Rg1, Rh1 and Rg2) and 20(S) protopanaxadiol (Rg3, Rd, Rb3, Rc, Rb1, Rb2). Improve perturbation of hepatic glucose uptake by GLUT 4 into peripheral tissue. Increases glucose absorption in muscles, lowers blood glucose and lowers HbA1c via activating AMPK and suppressing gluconeogenesis.^[112] Ginsenoside Re (20 mg/kg) decreased the fasting blood glucose level after 2 weeks of treatment in diabetic rat. It also has a protective effect against oxidative stress in the kidneys and eyes.^[113] After 1 hr of treatment with ginsenide Rh2 (1.0 mg/kg), plasma insulin levels rise from 211.42±13.84 to 342.34±12.23 pmol/L.^[111] In stz-induced diabetic rats, the extract and metformin reduce blood glucose levels up to 200 mg/dL after 35 days of treatment.^[112] In a 12-week randomised double-blind, placebo-controlled research of *P. ginseng* in diabetic patients, oral anti diabetic medication reduced blood glucose levels by 8-11% and plasma insulin levels by 33-38% after the oral glucose tolerance test.^[114] Compound K, a metabolite of protopanaxadiol ginsenosides, was combined with metformin (10 mg/kg) to lower plasma insulin and glucose levels.^[115] The pharmacokinetic parameter of metformin and the rate of absorption of metformin were not affected by the coadministration of P. ginseng 2g/kg with metformin 50 mg/kg.[112] The urine excretion rate is altered by repeated administration of red ginseng extract and the AUC and C_{max} of metformin are enhanced.^[116] BST204 is a ginseng extract that is dry. With IC₅₀ values of 17.4, 26.8, 31.5and 49.7l g/mL, BST204 mildly reduced the activity of CYP2C8, CYP2D6, CYP2C9and CYP2B6, respectively.[117]

Glycyrrhiza glabra L.

Liquorice plant (Glycyrrhiza glabra L.) belongs to the Fabaceae family. Glycyrrhizin, glycyrrhctinic acid, glabrene, formononetin, glabroi, isoliquiritigenin and liquiritigenin are phytoconstituents of G. glabra.^[118] Liquorice plant increases pancreatic cell number and improves tolerance to oral loading.^[119] G. Glabara and glycyrrhizin have an inhibiting impact on the cytochrome isozyme. Ethanol extract of G. Glabara affect the CYP3A4 and CYP2D6 with IC50 value 140.95±4.80 µg/mL and 132.49±1.07 μ g/mL and the IC₅₀ value for DMSO solvent of G. Glabra are 129.47±2.41 µg/mL and 125.16±0.88 µg/mL. The glycerrhizin in ethanol and DSMO inhibit CYP3A4 and CYP2D6 with IC₅₀ value174.62±2.30 µg/mL, 156.25±3.48 µg/mL and 172.33±1.92 μ g/mL, 153.38 \pm 1.98 μ g/mL. The crude extract has more potent action compared to bioactive compounds.[120] Licorice extract and glycerrhin increased the expression of CYP2B1, CYP2B9and CYP3A by 1.8-4.4 times, 1.8-2.2 times and 2 times, respectively. CYP2E1 and CYP1A1 activity have been reduced.^[121] The activities of CYP2C9, CYP2C19and CYP3A4 were all inhibited by glycyrhhetinic acid. On the activity of CYP1A2, CYP2D6and CYP2E1, a modest inhibitory effect has been seen, with an IC_{50}

| | | Table 2. Summary of Herbar Drug interaction. | | | | | | | | |
|------------|---------------------------------|---|---|---|-------------------------------------|-------------------------------------|--|--|--|--|
| SI. No. | Name of plant | Type of study | CYP/P-gp inhibition/induction | Interacting drug | Pharmaco- kinetic Interaction | Pharmaco- dynamic interaction | | | | |
| 1. | Abelmoschus esculentus | <i>In vitro</i> study and <i>in vivo</i> | - | Metformin | Yes | - | | | | |
| 2 | Phyllanthus emblica | <i>In vitro</i> study and <i>in vivo</i> | Inhibition of CYP1A2, CYP2C9, CYP2D6, CYP2E1, CYP3A4. | Metformin | Yes | Yes | | | | |
| 3. | Andrographis paniculata | <i>In vitro</i> study and <i>in vivo</i> | Inhibition of CYP3A4, CYP2D6 and CYP2C9. | Tolbutamide, Glyburide, gliclazide | Yes Yes Yes | No Yes - | | | | |
| 4 | Boswellia serrata | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP1A2, CYP2D6, CYP2C8, CYP2C9, CYP2C19 and CYP3A4. | Glimepride, metformin | Yes Yes | Yes Yes | | | | |
| 5 | Aloe barbadensis Miller | <i>In vitro, in vivo</i> study and human study | Inhibition of CYP3A4 and CYP2D6. | Glibenclamide | - | Yes | | | | |
| 6 | Bridelia ferrugenia | <i>In vitro, in vivo</i> study | - | Metformin | Yes | Yes | | | | |
| 7 | Cassia ariculata (L) | <i>In vitro, in vivo</i> study | Inhibition of CYP1A2, CYP2D6 CYP3A4. | Metformin | Yes | Yes | | | | |
| 8 | Catharanthus roseus | <i>In vitro, in vivo</i> study | Inhibition of CYP2D6 and CYP3A4. | Metformin, Glibenclamide | - | Yes No | | | | |
| 9 | Cinnamomum verum | In vitro, in vivo study | Inhibition of CYP2D1. | Metformin | - | Yes | | | | |
| 10 | Costus pictus | <i>In vitro, in vivo</i> study | - | Metformin | - | Yes | | | | |
| 11 | curcuma longa | In vitro, in vivo study | inhibition of CYP3A4, CYP1A2, CYP2C9 and CYP2D6, Pgp inhibitor. | Glyburide Glimepiride Metformin. | Yes Yes | Yes - Yes | | | | |
| 12 | Trigonella foenum graecum | <i>In vitro, in vivo</i> study and human study, | Inhibition of CYP2C9, CYP3A4, CYP2D6, CYP2C11. | Glimepiride Insulin Metformin Gliclazide. | - - Yes - | Yes Yes - Yes | | | | |
| 13 | Allium sativum L | <i>In vitro, in vivo</i> study and human study, | Inhibition of CYP3A, CYP2A CYP2C9, induction of Pgp. | Metformin | Yes | Yes | | | | |
| 14 | Tinospora cordifolia | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP3A4, CYP2D9, CYP1A2, CYP29. | Metformin Sitagliptin Glibenclamide Glimepiride. | No No Yes Yes | Yes Yes - | | | | |
| 15 | Zingiberis officinale Roscoe | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP2C9, CYP2C19, CYP3A4 and CYP2D6. | Sitagliptin Glibenclamide. | No - | No Yes | | | | |

Table 2: Summary of Herbal Drug Interaction.

| SI. No. | Name of plant | Type of study | CYP/P-gp inhibition/induction | Interacting drug | Pharmaco- kinetic Interaction | Pharmaco- dynamic interaction |
|------------|---|--|--|---|-------------------------------------|-------------------------------------|
| 16 | Ginkgo biloba L. | <i>In vitro, in vivo</i> study and human study | Inhibition of CYP2C9, CYP1A2 CYP2E1, CYP3A4. | Tolbutamide Metformin. | Yes No | - |
| 17 | Panax ginseng | <i>In vitro, in vivo</i> study and human study | Inhibition of CYP2C8, CYP2D6, CYP2C9 and CYP2B6. | Metformin | Yes | Yes |
| 18 | Glycyrrhiza glabra L. | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP3A4, CYP2D6, CYP2E1. | Glibenclamide | Yes | - |
| 19 | <i>Opuntia</i> spp | Human study | - | Glipizide Metformin | - | Yes Yes |
| 20 | Gymnema sylvestre | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP1A2, CYP2C9, CYP3A4 and CYP2C8. | Sitagliptin Gliclazide Glimepiride Metformin Glibenclamide. | Yes Yes No Yes Yes | - No Yes Yes No |
| 21 | Syzigium cumini | <i>In vitro</i> and <i>in vivo</i> study | Inhibition of CYP3A4, CYP2D6 induction of Pgp. | Gliclazide Sitagliptin Glipizide. | - Yes Yes | Yes Yes Yes |
| 22 | Scutellaria baicalensis | <i>In vitro</i> and <i>in vivo</i> study | Inhibits only CYP1A2 | Metformin | - | Yes |
| 23 | withania somnifera | <i>In vitro in vivo</i> study and human study | Induction of CYP1A. | Glimepiride | Yes | Yes |
| 24 | Swertia chirata | <i>In vito</i> and <i>in vivo</i> study | Inhibition of CYP3A4, CYP2D6. | Tolbutamide | - | Yes |
| 25 | Azadirachta indica | <i>In vito</i> and <i>in vivo</i> study | Induction of CYP3A4. | Gliclazide Glipizide | - No | Yes Yes |
| 26 | Momordica charantia | <i>In vito</i> and <i>in vivo</i> study | Inhibition of Pgp, CYP2C9 CYP2C19, CYP1A2, CYP3A4, CYP2A6. | Rosiglitazone Metformin Glibenclamide. | - - | Yes Yes Yes |
| 27 | <i>Moringa</i> <i>oleifera</i> , Lam | <i>In vito</i> and <i>in vivo</i> study | Inhibition of CYP1A2, CYP2D6, CYP2E1, CYP3A4. | Metformin Pioglitazone | - Yes | Yes - |

value of around 500 M.^[122] When 18- glycyrrhizin (25 mg/kg i.p.) and glibenclamide (1 mg/kg i.g.) were given together, C_{max} , AUC₀₋₁₄ hand elimination half time $T_{1/2}$, ke were all elevated by 18%, 59% and 63%, respectively, while glibenclamide elimination was reduced by 38%.^[123]

Opuntia spp. (Prickly pear cactus)

Opuntia, popularly known as prickly pear cactus, is a cactus that belongs to the *cactaceae* family.^[124] *Opuntia spp*. is a medicinal plant that is used to treat a variety of chronic diseases. It thrives

in arid climates with harsh environmental conditions.^[125] *In vivo* research has discovered that *Opuntia spp.* have alpha glucosidase inhibitory action. The glucose absorption has declined in small intestine after the administration of *opuntia* spp. The blood glucose level of streptozocine induced rat has been reduced. After receiving an aqueous extract of *Opuntia ficus indica*, the AUC of blood glucose was reduced by 33.10%.^[126] The stimulation of the AMPK/P38 MAPK pathway in L6 myoblast cells by *Opuntia ficus indica* and enhanced GLUT-4 translocation and glucose absorption.^[127] The *opuntia* spp. contain 14.25%±0.062

pectin fiber^[128] which reduces the glucose absorption,^[129] phenols, flavonoids, dietary fibers, betalains, taurine linolic acid, vitamins, minerals and free amino acids are some of the bioactive compounds found in *Opuntia* spp.^[130] The alkaloids Indica xanthin, neobatanin, fibre, flavonoids and poly saccharides have been proven to have anti-diabetic and anti-glycation property.^[131] In clinical investigations, *Opuntia ficus indica* combined with leucine produced a synergistic effect in males after a vigorous workout.^[132] With the *Opuntia* spp., glipizide and metformin were given together. The results revealed an additive impact that resulted in hypoglycemia in diabetic patients.^[133]

Gymnema sylvestre

In Hindi, Gymnema sylvestre (family: Asclepiadaceae) is called as gur-mar.^[134] A new extract of G. sylvestre known as OSA improved glucose tolerance by reducing intestinal glucose absorption after an oral glucose load, decreasing insulin resistance, or increasing plasma insulin levels. OSA functioned to preserve insulin reserves by boosting insulin production through an increase in PPI mRNA. Gymnema sylvestre extracts have been shown to limit glucose absorption in the gut.^[135] Gymnemic acids, gymnemosides, gymnemasaponins, gurmarin, gymnemanol, stigmasterol, d-quercitol, amyrin related glycosides, anthraquinones, lupeol, hydroxycinnamic acids and coumarols are among the plant's bioactive ingredients. Anti-diabetic action has been found in gymnemic acids, gymnemasaponins and Gurmarin.^[136] The extracts of G. sylvetre in chloroform, n-hexane and ethyl acetate decrease the activities of CYP1A2, CYP2C9 and a modest inhibition of CYP3A4 and CYP2C8.[137] When sitagliptin (20 mg/ kg) was co-medicated with G. sylvestre extract (400 mg/kg), the bioavailability and AUC of sitagliptin (20 mg/kg) decreased.^[138] The reduction found in bioavailability by 43%, in C_{max} by 75%, the absorption rate constant by 95% and an increment in clearance of gliclazide (40 mg/kg p.o.) when combine with G. sylvestre extract (30 mg/kg p.o.). There was a maximum hypoglycemic effect observed but less than gliclazide alone.^[139] No significant pharmacokinetic interaction found between glimepiride (0.8 mg/kg) and G. sylvestre (400 mg/kg) in the study. On a 28-day treatment, the levels of FBGL and Hb1Ac were reduced by 63.38% and 4.82%, respectively, a more potent effect than the individual effects of glimepiride and G. sylvestre.^[140] G sylvestre (100 mg/kg and 500 mg/kg p.o.) and metformin (50 mg/kg and 100 mg/kg p.o.) interact, G. sylvestre (100 mg/kg) and metformin (50 mg/kg) decreased $\mathrm{AUC}_{_{0-\infty}}, \mathrm{C}_{_{\mathrm{max}}}$ and $\mathrm{K}_{_{\mathrm{a}}}$ by 27%, 34% and 37%, respectively and increased clearance by 38%, while G. sylvestre 500 mg/kg and metformin 100 mg/kg decreased AUC, C_{max} and Ka by 47%, 53% and 41%, respectively and increased clearance by 88%. The combination of metformin and G. sylvestre dramatically reduced the FBGL to 140.4±3.4 mg/dL.^[139] Taking G. sylvestre 500 mg/kg with glibenclamide 0.5 mg/kg together reduced AUC $_{\scriptscriptstyle \infty}$ by 17%, C $_{\scriptscriptstyle \rm max}$ by 19%, Ka by 24% and increased clearance by

14%. The second combination of G. sylvestre 500 mg/kg and a

high dose of glibenclamide (0.6 mg/kg) lower AUC_{0-∞} by 8-7% increases C_{max} by 13%, Ka by 9% and increases clearance by 29%. This combination lowers blood glucose and cholesterol levels, but it has a smaller effect than glibenclamide (0.6 mg/kg).^[141]

Syzigium cumini

Eugenia jambolana Lam. (syn. Syzigium cumini (L.), Family: Myrtaceae), often known as black plum in English or Jamun in Hindi, is a plant that grows in the Myrtaceae family. Ellagic acids, isoquercetin, quercetin, kampferol, myricetin anthocyanins, delphinidin, petunidin, malvidin-diglucosides, jambosine, gallic acid, corilagin, -sitosterol, betulinic acid, mycaminose are phytochemicals found in plants.^[142] Mycaminose has anti-diabetic properties. Aqueous extract of seed (2.5 g/kg) was given to diabetic rats for 30 days, the results were observed that, seed extract increases the glycolysis and decreases glucose formation by increment in hexokinase activity and reduces insulin sensitivity by augmentation of glucose utilization. Extract administration also decreases the tissue damaged by oxidative stress.^[143] The interaction of crude seed extract of S. cumini with CYP isoenzymes was examined .S. cumini crude extract inhibited CYP3A4 with an IC₅₀ of 76.69 µg/mL, whereas CYP3A4 $(IC_{50}=359.02 \ \mu g/mL)$ and CYP2D6 $(IC_{50}=493.05 \ \mu g/mL)$ were only weakly inhibited.^[144] The methanolic extract of S. cumini (500 mg/kg) with gliclazide 2 mg/kg reduced the blood glucose level by 35% in 12 hr of administration.^[145] Combining sitagliptin and aqueous seed extract of E. jambolana (400 mg/kg) effectively lowered FBL to 298.74 mg/dL after 28 days of treatment. Cmax and AUC_{0.24}, both pharmacokinetic parameters, were lowered by 38.70% and 22.40%, respectively. The decrease in absorption could be related to increased P-gp expression or stimulation of the CYP enzyme. The pancreatic tissue was able to recover from oxidative injury in a considerable way.^[146] The combination of S. cumini seed extract (250 mg/kg and 400 mg/kg p.o.) and glipizide enhances the AUC, C_{max} and $T_{1/2}$ of glipizide, while the *S. cumini* (100g) extract suppresses the CYP3A enzymatic activity. The S. cumini extract at 250 mg/kg and 400 mg/kg in combination with glipizide dramatically lowers blood glucose levels.[147]

Scutellaria baicalensis

Scutellaria baicalensis, often known as Scutellariae radix or skullcap root. Baicalin, baicalein, wogonin, chalones, flavanonols and anthocyanidines are active phytoconstituents of *S. Baicalensis*.^[148] *S. baicalensis* (3.52 g) and metformin (500 mg) together enhanced insulin resistance and hepatic enzymatic activity, as well as glucose tolerance.^[149] The *S. baicalensis* extract inhibits solely CYP1A2 with an IC₅₀ value of 0.5 μ M to 19.9 μ M.^[150] When *S. baicalensis* is used with a drug metabolized by CYP1A2, it may induce an herbal drug interaction. To test this, more research is required. A pharmacokinetic interaction between metformin (500 mg/kg) and *S. baicalensis* (400 mg/kg) given to STZ induced diabetic rats for 30 days, which reduced 38.2% blood glucose levels and plasma TC levels (99.733 mg/dL) with a greater effect than the metformin and *S. baicalensis* groups treated separately.^[151]

Withania somnifera

Ashwagandha or Indian ginseng is the way of referring to Withania somnifera (L.) Dunal (family: solenaceae). Isopelletierine, anaferine (alkaloids), tamanolides', withaferins (steroidal lactones), sitoindoside VII and VIII and sitoindoside IX and X are the biologically active chemical ingredients.^[152] A research study found increases of 2.13, 1.95, 1.35and 1.20 times in C_{max} , AUC_{0- ∞}, $T_{_{\rm I\!\!\!\!\!\!\!\!\!\!\!\!\!\!\!\!\!}}$ and MRT and decreases of 0.51 and 0.69 folds in clearance and volume of distribution, in glimepiride pharmacokinetic parameter, the oral bioavailability by 33.5%. The combination of ashwagandha (500 mg/kg) with glimepiride (1 mg/kg) resulted in a 55.46% reduction in glucose levels after 6 hr, compared to glimepiride (46.06%).^[153] The inhibitory impact of crude extract of W. somnifera (10-640 µg/mL) and active ingredient (1-32 µM) on cytochrome isoenzyme. In vitro studies revealed that crude extract and phytoconstituent had no inhibitory effect on CYP1A, with IC₅₀ values of 500 µg/mL and 32 µM, respectively. In vivo studies revealed that the methanolic extract of W. somnifera and phenacetin reduced $AUC_{0-\infty}$ by 31%, increased elimination constant by 48% and decreased half-life $T_{1/2}$ by 43%, with no significant difference in $\mathrm{C}_{_{\mathrm{max}}}$ of phenacetin. These findings could point to an increase in the CYP1A isoenzyme, which could reduce the therapeutic efficacy of a medicine metabolized by the CYP1A enzyme.^[154] In a clinical trial on patients with mild hyperglycemia and hypercholesteremia, after 30 days of treatment, NIDDM patients who were administered ashwagandha 500 mg capsules, 6 capsules per day (3 g/day), saw a 12% reduction in blood glucose levels. Ashwagandha has been shown to boost insulin levels. Serum cholesterol, triglycerides, LDL and VLDL cholesterol levels were reduced by 10%, 15%, 6% and 15%, respectively.^[155]

Swertia chirata

Chirata or Buch-Ham is the common name for *Swertia chirata* (family: Gentianaceae). It's a type of Chinese medicine that dates back thousands of years.^[156] Amarogentin, mangiferin and swertia merin are phytochemicals of *Swertia chirata*. Mangiferin is proven to lower blood glucose levels while also lowering lipid levels.^[157] The ethanlic and DSMO extract of *Swertia chirata* inhibited CYP3A4 (197.49±2.68 and 193.63±2.87) and CYP2D6 (211.45±3.54 and 208.34±1.26) respectively. The hexane extract of *S. chirata* (250 mg/kg) given to tolbutamide pre- treated rats, a decrement in blood sugar level at 4 hr.^[158]

Azadirachta indica

Azadirachta indica (family: *Meliaceae*) is a plant that is generally referred to as neem in Hindi and is utilized in Ayurveda and Unani medicine.^[159] *A. indica* stimulates beta cell insulin release, improves glucose utilization and insulin sensitivity and prevents

glucose absorption in the gut.^[160] The aqueous and acetone extracts of A. *indica* inhibited α amylase and α glucosidase with IC₅₀ values of 9.15 mg/mL and 5.00 mg/mL, respectively.^[161] After 21 days of treatment, chloroform extract of A. indica lowers fasting blood glucose (109.65 mg/dL). The activity of the glucosidase enzyme was reduced by 51% and 35%, respectively, after administration of chloroform and aqueous neem extract. After 21 days of A. indica administration, plasma insulin secretion from cells and G6PD activity were shown to be enhanced.^[162] In diabetic rats and normal rabbits, the co-administration of aqueous neem extract (30 mg/ kg) with gliclazide (2 mg/kg) lowered blood glucose levels by 28.1 and 1.2%, respectively. After administration of A. indica, there was no significant pharmacokinetic interaction in gliclazide.^[163] The co-administration of A. indica (250 mg) with glipizide (5 mg/kg) had no significant effect on gliclazide pharmacokinetic parameters, but A. indica (500 mg/kg p.o.) decreased AUC and $\mathrm{T_{_{1/2}}}$ but had no effect on $\mathrm{C_{_{max}}}$ MRT significantly increased and T_{max} significantly increased, indicating that A. indica. The A. indica extract (100 µg) caused induction of CYP3A4 enzymatic activity. No significant difference was found in the blood glucose level when treated with combination of A. indica 250 mg+5 mg/ kg and 500 mg/kg+5 mg compared to gliclazide (5 mg/kg p.o.) alone. The AST and ALT level decreased in A. indica (500 mg/kg) and gliclazide (5 mg/kg). Glucose load has been decreased after 120 min of treatment.^[160] Because of the elevation of CYP3A4 enzymatic activity, A. indica has an herbal medication interaction with gliclazide.

Momordica charantia

Momordica charantia (Family: Cucurbitaceae) is commonly known as bitter gourd. Neuropathy, retinopathy, cardiomyopathy, nephropathy, exocrine gland insufficiency and various other diabetes problems are all treated with Momordica charantia. Momordin charantin and momordicin are bioactive phytochemicals. These compounds have anti-diabetic properties.^[164] M. charantia stimulated beta cells, which increased glucose uptake and insulin levels.^[165] In H411EC3 hepatoma cells, an ethyl acetate extract of M. charantia activates PPARy and PPARa.^[166] Glycogen synthesis was induced by M. Charantia.^[167] M. charantia inhibits P-gp with IC₅₀ value of 16±0.4 and raise the induction of PXR activity by 2 times. Methanolic extract of M. charantia inhibits CYP2C9 and CYP2C19 strongly, CYP1A2, CYP3A4 and CYP2A6 weakly.^[168] The glucose load and blood glucose level have been reduced by the combination of rosiglitazone (2 mg and 5 mg) and M. charantia (500 mg/kg) methanolic extract. The potent effect shown by the high dose of rosiglitazone (5 mg) with MC.^[169] In clinical study, the carbon tetrachloride+hexene extract of M. charantia with metformin (2.5 mg/kg.po.) reduced 11% FBS and 17% PPBS and extract with glibenclamide (2.5mg/kg p.o.) reduced 13% FBS and 15% PPBS in diabetic patient, the combination of *M. charantia* extract with metformin (2.5 mg) and glibenclamide 2.5 mg have reduced 13% FBS and 21% PPBS, *Momordica charantia*+antihyperglycemic drug showed synergistic effect.^[170]

Moringa oleifera, Lam

Moringa oleifera, Lam (family: Moringaceae) is commonly known as drumsticks.^[171] The phytochemicals are 4(a L-rhamnosyloxy)-benzyl isothiocyanate, niazimicin, 3-O-(6'-O-oleoyl-β-D-glucopyranosyl)-β-sitosterol, β-sitosterol-3-O-β-D-glucopyranoside, niazirin, β-sitosterol and glycerol-1-(9-octadecanoate),[172] chlorogenic acid, rutin, quercetin glucoside and kaempferol rhamnoglucoside.[173] Ethanolic and aqueous extract of M. oleifera inhibited human CYP1A2, CYP2D6, CYP2E1 and CYP3A4 activities with IC₅₀ value 13.8 to 1500 µg/mL. Ethanolic extract of M. oleifera shown specific inhibition of CYP1A2 with IC_{50} 13.8 µg/mL and CYP3A4 with $IC_{_{50}}$ 101 $\mu g/mL^{_{[174]}}$ and A study has found a synergistic effect of met (150 mg/kg) and ethanolic extract of MO 375, 750 and 1500 rmg/kg in alloxan induced diabetic rat, the blood glucose level and serum lipid level have been reduced in 28 days.^[175] Aqueous extract of *M. oleifera* leaves (200 µg/mL) inhibited α -amylase and α - glucosidase activity by 80.5% and 75.65% respectively. Intestinal absorption inhibited by extract at 100 µg/mL by 86.25%.the glucose uptake increased by 24.3% at the dose of 100 µg/mL.^[171] Mukharjee et al.^[176] resulted that, hydroalcoholic extract of Moringa oleifera and chlorogenic acid weak inhibitor of CYP3A4 and CYP2D6 compared to positive inhibitors. The co- administration of pioglitazone (3 mg/kg) and moringa (800 mg/kg) have reduced the glucose levels and body weights on contrast the half dose of pioglitazone 1.5 mg/ kg and 400 mg/kg have produced significant interaction with the Pharmacokinetic parameter AUC, T_{1/2}, Ke, CL, Vd administered to diabetic and normal rabbit. The single dose decreased AUC and in opposite multiple doses increased AUC.[177]

DISCUSSION

Herbal medicines are widely utilised and various formulations are available as Over-The-Counter (OTC) and prescription pharmaceuticals. Patients are encouraged to take herbal medications in addition to their allopathic treatment. This could result in an herbal drug interaction and a lack of understanding of HDI could worsen the patient's condition. Cytochrome inhibition or induction is the mechanism of pharmacodynamic and pharmacokinetic interactions. Andrographis paniculata and Catharanthus roseus exhibited no impact when taken together with tolbutamide and glibenclamide respectively. Metformin's bioavailability and therapeutic and pharmacological effects were lowered by Bridelia ferugenia. Before combining allopathy with herbs, they should be researched thoroughly. Plant phytochemicals interfere with cytochrome activity, promoting cytochrome induction or inhibition. It is necessary to investigate phytoconstituents and cytochrome induction or inhibition activity to improve therapeutic and pharmacological effects,

safety, efficacy and dose adjustment. The phytoconstituents present in medicinal plants are responsible for the herbal drug interaction. The phytoconstituents alter the cytochrome activities by inhibition or induction. Cytochrome induction and inhibition distress the pharmacokinetic and pharmacodynamic effects. These are major concerns when we combine herbs with allopathy. The safety and efficacy of drug should be study for better therapeutic effect. The researcher should pay attention towards this. Herbal drug interaction should be measured for better treatment and mitigation of diabetes Mellitus.

CONCLUSION

The exploration of interactions between herbal remedies and conventional antidiabetic medications presents significant implications for diabetes management. Herbal treatments, with a long-standing history of use in managing chronic conditions, offer promising benefits when integrated properly with allopathic therapies. As highlighted in the findings of the study, numerous plants exhibit pharmacological properties that can enhance glycemic control and improve metabolic profiles in diabetic patients.

However, the potential for herb-drug interactions remains a critical concern, primarily due to the involvement of cytochrome P450 enzymes and other metabolic pathways. Understanding the pharmacokinetics of both herbal and conventional medications is essential for optimizing therapeutic efficacy while minimizing adverse effects. The research indicates that certain herbal compounds can modulate the bioavailability and effectiveness of antidiabetic drugs, such as metformin and glimepiride.

Future research should focus on elucidating the mechanisms underlying these interactions and establishing comprehensive guidelines for patients seeking to incorporate herbal remedies into their diabetes management strategies. By fostering a collaborative approach between traditional and modern medicine, we can create a more effective, safe, and holistic framework for treating diabetes, ultimately enhancing patient outcomes and quality of life.

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CONFLICT OF INTEREST

The authors declare that there is no conflict of interest.

ABBREVIATIONS

AUC: Area Under the Curve; **AUC**₀: Area Under the Curve from 0 to the last measurable concentration; **AUC**_{total}: Total Area Under the Curve; **AMPK:** AMP-Activated Protein Kinase; **C**_{max}:

Maximum Plasma Concentration/Peak Plasma Concentration; CYP: Cytochrome; DSMO: Dimethyl Sulfoxide; FBS: Fasting Blood Sugar; GOT: Glutamic Oxaloacetic Transaminase; GLUT: Glucose Transporter; GPT: Glutamic Pyruvate Transaminase; HbA1c: Hemoglobin A1c; LDL: Low-Density Lipoprotein; MRT: Mean Residence Time; PgP: P-Glycoprotein; PPAR: Peroxisome Proliferator-Activated Receptor; T1/2: Half-life; T2DM: Type 2 Diabetes Mellitus; T_{max}: Time to reach maximum concentration; TC: Total Cholesterol; Vd: Volume of distribution.

SUMMARY

This review article investigates the complex interplay between herbal remedies and oral antidiabetic medications. Focusing on 27 antidiabetic plants highlights the increasing prevalence of diabetes mellitus globally, characterized by high blood glucose levels due to insufficient insulin function. The extensive use of herbal supplements alongside conventional treatments necessitates a thorough understanding of potential Herb-Drug Interactions (HDIs). The review details the diverse mechanisms by which these 27 plants may impact blood glucose, including AGE inhibition, GLP-1 stimulation, beta-cell regeneration and glucose metabolism modulation (Table 1). However, this article emphasizes the critical role of Cytochrome P450 (CYP) enzymes in mediating pharmacokinetic and pharmacodynamic HDIs. CYP inhibition can enhance drug bioavailability, potentially causing adverse effects, while induction diminishes therapeutic efficacy. The study meticulously examines the individual interactions of each plant with specific antidiabetic drugs, documenting CYP involvement and resultant changes in pharmacokinetic and pharmacodynamic parameters. (Summarized in Table 2). The authors conclude that a comprehensive understanding of HDIs is paramount for safe and effective diabetes management. They stress the need for further research focusing on dose-dependent effects and identifying specific phytochemicals responsible for these interactions to optimize therapeutic outcomes and minimize adverse events. This necessitates a shift towards more rigorous safety and efficacy testing of herbal remedies used concomitantly with conventional antidiabetic medications.

REFERENCES

- No author listed. What is diabetes? [cited Oct 27 2023] Available from: https://www.i df.org/aboutdiabetes/what-is-diabetes.shtml.
- Expert Committee on the Diagnosis and Classification of Diabetes Mellitus. Report of the expert committee on the diagnosis and classification of diabetes mellitus. Diabetes Care. 2003;26 Suppl 1:S5-20. doi: 10.2337/diacare.26.2007.s5, PMID 12502614.
- No author listed. Diabetes. [cited Oct 27 2023] Available from: https://www.who.int/ news-room/fact-sheets/detail/diabetes.
- 4. Das SK, Elbein SC. The genetic basis of type 2 diabetes. Cellscience. 2006;2(4):100-31. doi: 10.1901/jaba.2006.2-100, PMID 16892160.
- Darenskaya MA, Kolesnikova LI, Kolesnikov SI. Oxidative stress: pathogenetic role in diabetes mellitus and its complications and therapeutic approaches to correction. Bull Exp Biol Med. 2021;171(2):179-89. doi: 10.1007/s10517-021-05191-7, PMID 34173093.
- Grover JK, Yadav S, Vats V. Medicinal plants of India with anti-diabetic potential. J Ethnopharmacol. 2002;81(1):81-100. doi: 10.1016/s0378-8741(02)00059-4, PMID 12020931.

- Prabhakar PK, Doble M. Mechanism of action of natural products used in the treatment of diabetes mellitus. Chin J Integr Med. 2011;17(8):563-74. doi: 10.1007/ s11655-011-0810-3, PMID 21826590.
- Choi JG, Eom SM, Kim J, Kim SH, Huh E, Kim H, et al. A comprehensive review of recent studies on herb-drug interaction: a focus on pharmacodynamic interaction. J Altern Complement Med. 2016;22(4):262-79. doi: 10.1089/acm.2015.0235, PMID 27003511.
- Manikandan P, Nagini S. Cytochrome P450 structure, function and clinical significance: a review. Curr Drug Targets. 2018;19(1):38-54. doi: 10.2174/1389450118 666170125144557, PMID 28124606.
- Bijnsdorp IV, Giovannetti E, Peters GJ. Analysis of drug interactions. Methods Mol Biol. 2011;731:421-34. doi: 10.1007/978-1-61779-080-5_34, PMID 21516426.
- 11. Sharma N, Sharma M, Bindal MC. Potential antidiabetic herbal drugs: a comparative review of marketed products. Res J Pharmacogn Phytochem. 2010;2(2):115-21.
- 12. Ritter JM, Rang HP, Flower RJ, Henderson G. Rang and Dale's pharmacology. Elsevier Health Sciences; 2014.
- Rendic S, Carlo FJ. Human cytochrome P450 enzymes: a status report summarizing their reactions, substrates, inducers and inhibitors. Drug Metab Rev. 1997;29(1-2):413-580.
- Shaikh AS, Thomas AB, Chitlange SS. Herb-drug interaction studies of herbs used in treatment of cardiovascular disorders—A narrative review of preclinical and clinical studies. Phytother Res. 2020;34(5):1008-26. doi: 10.1002/ptr.6585, PMID 31908085.
- Prabhune A, Sharma M, Ojha B. Abelmoschus esculentus (Okra) potential natural compound for prevention and management of diabetes and diabetic induced hyperglycemia. Tamil Nadu. Int J Herb Med. 2017;5(2):66-8.
- Abbas AY, Muhammad I, AbdulRahman MB, Bilbis LS, Saidu Y, Onu A. Possible antidiabetic mechanism of action of ex-maradi okra fruit variety (*Abelmoscus* esculentus) on alloxan induced diabetic rats. Niger J Basic Appl Sci. 2017;25(2):101-13.
- Li N, Tang H, Wu L, Ge H, Wang Y, Yu H, *et al.* Chemical constituents, clinical efficacy and molecular mechanisms of the ethanol extract of *Abelmoschus manihot* flowers in treatment of kidney diseases. Phytother Res. 2021;35(1):198-206. doi: 10.1002/ptr.6 818, PMID 32716080.
- Romdhane MH, Chahdoura H, Barros L, Dias MI, Carvalho Gomes Corrêa RC, Morales P, et al. Chemical composition, nutritional valueand biological evaluation of Tunisian okra pods (*Abelmoschus esculentus* L. Moench). Molecules. 2020;25(20):4739. doi: 10. 3390/molecules25204739, PMID 33076530.
- Khatun H, Rahman A, Biswas M, Islam AU. Water-soluble fraction of *Abelmoschus* esculentus L. interacts with glucose and metformin hydrochloride and alters their absorption kinetics after coadministration in rats. ISRN Pharm. 2011; 2011:260537. doi: 10.5402/2011/260537, PMID 22389848.
- Gaire BP, Subedi L. Phytochemistry, pharmacology and medicinal properties of *Phyllanthus emblica* Linn. Chin J Integr Med. 2014:1-8. doi: 10.1007/s11655-014-1984-2, PMID 25491539.
- Sharma P, Joshi T, Joshi T, Chandra S, Tamta S. In silico screening of potential antidiabetic phytochemicals from *Phyllanthus emblica* against therapeutic targets of type 2 diabetes. J Ethnopharmacol. 2020;248:112268. doi: 10.1016/j.jep.2019.11226 8, PMID 31593813.
- Sultana Z, Jami SI, Ali E, Begum M, Haque M. Investigation of antidiabetic effect of ethanolic extract of *Phyllanthus emblica* Linn. fruits in experimental animal models. 2014.
- Huang HZ, Qiu M, Lin JZ, Li MQ, Ma XT, Ran F, et al. Potential effect of tropical fruitsPhyllanthus emblica L. for the prevention and management of type 2 diabetic complications: a systematic review of recent advances. Eur J Nutr. 2021;60(7):3525-42. doi: 10.1007/s00394-020-02471-2.
- 24. Anannarukan N, Niwattisaiwong N, Warisnoicharoen W, Winitthana T, Pramyothin P, Chaichantipyuth C, et al. Inhibition of human cytochrome P450 *in vitro* by*Phyllanthus amarus* and*Phyllanthus emblica* aqueous extracts. Thai J Pharm Sci. 2012;36(4):135-43. doi: 10.56808/3027-7922.2142.
- Shengule S, Kumbhare K, Patil D, Mishra S, Apte K, Patwardhan B. Herb-drug interaction of Nisha Amalaki and curcuminoids with metformin in normal and diabetic condition: a disease system approach. Biomed Pharmacother. 2018;101:591-8. doi: 1 0.1016/j.biopha.2018.02.032, PMID 29518605.
- Wibudi A, Kiranadi B, Manalu W, winarto A, Suyono S. The traditional plant and Rographis paniculata (Sambiloto), exhibits insulin-releasing actions in vitro. Acta Med Indones. 2008;40(2):63-8. PMID 18560025.
- Ooi JP, Kuroyanagi M, Sulaiman SF, Muhammad TS, Tan ML. Andrographolide and 14-deoxy-11,12-didehydroandrographolide inhibit cytochrome P450s in HepG2 hepatoma cells. Life Sci. 2011;88(9-10):447-54. doi: 10.1016/j.lfs.2010.12.019, PMID 21219911.
- Pan Y, Abd-Rashid BA, Ismail Z, Ismail R, Mak JW, Pook PC, et al. *In vitro* determination of the effect of *Andrographis paniculata* extracts and andrographolide on human hepatic cytochrome P450 activities. J Nat Med. 2011;65(3-4):440-7. doi: 10.1007/ s11418-011-0516-z, PMID 21365364.
- Chen HW, Huang CS, Liu PF, Li CC, Chen CT, Liu CT, et al. Andrographis paniculata extract and andrographolide modulate the hepatic drug metabolism system and plasma tolbutamide concentrations in rats. Evid Based Complement Alternat Med. 2013; 2013:982689. doi: 10.1155/2013/982689, PMID 23997806.
- Pekthong D, Martin H, Abadie C, Bonet A, Heyd B, Mantion G, et al. Differential inhibition of rat and human hepatic cytochrome P450 by Andrographis paniculata

extract and andrographolide. J Ethnopharmacol. 2008;115(3):432-40. doi: 10.1016/ j.jep.2007.10.013, PMID 18053665.

- Samala S, Veeresham C. Pharmacokinetic and pharmacodynamic interaction of boswellic acids and andrographolide with glyburide in diabetic rats: including its PK/PD modeling. Phytother Res. 2016;30(3):496-502. doi: 10.1002/ptr.5556, PMID 26762235.
- 32. Mouid MG. Effect of ethanolic extract of aerial parts of *Andrographis paniculata* on the pharmacokinetics of gliclazide in rats. Asian J Biomed PharmSci. 2015;5(51):21-4. doi: 10.15272/ajbps.v5i51.755.
- 33. Gomaa AA, Makboul RM, Al-Mokhtar MA, Nicola MA. Polyphenol-richBoswellia serrata gum prevents cognitive impairment and insulin resistance of diabetic rats through inhibition of GSK3β activity, oxidative stress and pro-inflammatory cytokines. Biomed Pharmacother. 2019;109:281-92. doi: 10.1016/j.biopha.2018.10.0 56, PMID 30396086.
- Roe AL, Wilcox R, Price JM, Li L, Dai H, Freeman KM, et al. An evaluation of potential inhibition of CYP3A4/5 and CYP2C9 enzymatic activity byBoswellia serrata extract. Appl in vitro Toxicol. 2019;5(1):34-46. doi: 10.1089/aivt.2018.0023.
- Frank A, Unger M. Analysis of frankincense from various*Boswellia* species with inhibitory activity on human drug metabolising cytochrome P450 enzymes using liquid chromatography mass spectrometry after automated on-line extraction. J Chromatogr A. 2006;1112(1-2):255-62. doi: 10.1016/j.chroma.2005.11.116, PMID 16364338.
- Samala S, Veeresham C. Enhanced bioavailability of glimepiride in the presence of boswellic acids in streptozotocin-induced diabetic rat model. Nat Prod Chem Res. 2013;1(4). doi: 10.4172/2329-6836.1000116.
- Samala S, Veeresham C. Boswellic acids pretreatment enhances the bioavailability and hypoglycemic action of metformin in rats: involvement of CYP3A inhibition. Int J Pharm Pharmacol. 2018;2(3):1-8. doi: 10.31531/2581-3080.1000132.
- Beppu H, Shimpo K, Chihara T, Kaneko T, Tamai I, Yamaji S, et al. Antidiabetic effects of dietary administration of Aloe arborescens Miller components on multiple low-dose streptozotocin-induced diabetes in mice: investigation on hypoglycemic action and systemic absorption dynamics of aloe components. J Ethnopharmacol. 2006;103(3):468-77. doi: 10.1016/j.jep.2005.10.034, PMID 16406411.
- Muñiz-Ramirez A, Perez RM, Garcia E, Garcia FE. Antidiabetic activity of *Aloe vera* leaves. Evid Based Complement Alternat Med. 2020; 2020:6371201. doi: 10.1155/2 020/6371201, PMID 32565868.
- 40. Djuv A, Nilsen OG. *Aloe vera* juice: IC50 and dual mechanistic inhibition of CYP3A4 and CYP2D6. Phytother Res. 2012;26(3):445-51. doi: 10.1002/ptr.3564, PMID 21842479.
- Bunyapraphatsara N, Yongchaiyudha S, Rungpitarangsi V, Chokechaijaroenporn O. Antidiabetic activity of *Aloe vera* L. juice II. Clinical trial in diabetes mellitus patients in combination with glibenclamide. Phytomedicine. 1996;3(3):245-8. doi: 10.1016/ S0944-7113(96)80061-4, PMID 23195078.
- Adewale O, Oloyede O. Hypoglycemic activity of aqueous extract of the bark of Bridelia ferruginea in normal and alloxan-induced diabetic rats. Prime Res Biotechnol. 2012;2:53-6.
- Oyebode O, Erukainure OL, Zuma L, Ibeji CU, Koorbanally NA, Islam MS. *In vitro* and computational studies of the antioxidant and anti-diabetic properties of *Bridelia ferruginea*. J Biomol Struct Dyn. 2020:1-15.
- 44. Sakyiamah MM, BrewDaniels H, Appiah AA, Edoh D. Herb-drug interaction: effect of aqueous extract of *Bridelia ferruginea* leaves on the pharmacokinetics of metformin. J Med Herbs Ethnomed. 2015;1(1):84-8. doi: 10.5455/jmhe.2015.09.019.
- 45. Thomford KP, Edoh DA, Sarkodie JA, Thomford AK, Essuman K, Owusu D. Re-evaluating the efficacy of the aqueous leaf extract of *Bridelia ferrugenia* and its potential combination with metformin in the management of diabetes mellitus. J Med Herbs Phytochem. 2015.
- 46. Nambirajan G, Karunanidhi K, Ganesan A, Rajendran R, Kandasamy R, Elangovan A, et al. Evaluation of antidiabetic activity of bud and flower of Avaram Senna (*Cassia* auriculata L.) in high fat diet and streptozotocin induced diabetic rats. Biomed Pharmacother. 2018;108:1495-506. doi: 10.1016/j.biopha.2018.10.007, PMID 30372851.
- Gupta S, Sharma SB, Singh UR, Bansal SK, Prabhu KM. Elucidation of mechanism of action of *Cassia auriculata* leaf extract for its antidiabetic activity in streptozotocin-induced diabetic rats. J Med Food. 2010;13(3):528-34. doi: 10.1089/ jmf.2009.1253, PMID 20521978.
- 48. Thilakarathna GC, Navaratne SB, Wickramasinghe I, Ranasinghe P, Samarkoon SR, Samarasekera JK. The effect of *Salacia reticulata*, *Syzygium cumini*, *Artocarpus heterophyllus* and *Cassia auriculata* on controlling the rapid formation of advanced glycation end-products. J Ayurveda Integr Med. 2021;12(2):261-8. doi: 10.1016/j.jaim .2020.10.010, PMID 33731265.
- Appiah-Opong R, Commandeur JN, Axson C, Vermeulen NP. Interactions between cytochromes P450, glutathione S-transferases and Ghanaian medicinal plants. Food Chem Toxicol. 2008;46(12):3598-603. doi: 10.1016/j.fct.2008.09.002, PMID 18822337.
- Elango H, Ponnusankar S, Sundaram S. Assessment of pharmacodynamic and pharmacokinetic interaction of aqueous extract of *Cassia auriculata* L. and metformin in rats. Pharmacogn Mag. 2015;11 Suppl 3:S423-6. doi: 10.4103/0973-1296.168986, PMID 26929576.
- Al-Shaqha WM, Khan M, Salam N, Azzi A, Chaudhary AA. Antidiabetic potential of *Catharanthus roseus* Linn. and its effect on the glucose transport gene (GLUT-2 and GLUT-4) in streptozotocin-induced diabetic Wistar rats. BMC Complement Altern Med. 2015;15(1):379. doi: 10.1186/s12906-015-0899-6, PMID 26490765.

- Tiong SH, Looi CY, Hazni H, Arya A, Paydar M, Wong WF, et al. Antidiabetic and antioxidant properties of alkaloids from*Catharanthus roseus* (L.) G. Don. Molecules. 2013;18(8):9770-84. doi: 10.3390/molecules18089770, PMID 23955322.
- Kumar S, Singh B, Singh R. Catharanthus roseus (L.) G. Don: a review of its ethnobotany, phytochemistry, ethnopharmacology and toxicities. J Ethnopharmacol. 2022;284:114647. doi: 10.1016/j.jep.2021.114647, PMID 34562562.
- Zhang L, Wei G, Liu Y, Zu Y, Gai Q, Yang L. Antihyperglycemic and antioxidant activities of total alkaloids from *Catharanthus roseus* in streptozotocin-induced diabetic rats. J For Res. 2016;27(1):167-74. doi: 10.1007/s11676-015-0112-2.
- Usia T, Watabe T, Kadota S, Tezuka Y. Cytochrome P450 2D6 (CYP2D6) inhibitory constituents of *Catharanthus roseus*. Biol Pharm Bull. 2005;28(6):1021-4. doi: 10.124 8/bpb.28.1021, PMID 15930738.
- Ohadoma SC, Michael HU. Effects of co-administration of methanol leaf extract of *Catharanthus roseus* on the hypoglycemic activity of metformin and glibenclamide in rats. Asian Pac J Trop Med. 2011;4(6):475-7. doi: 10.1016/S1995-7645(11)60129-6, PMID 21771702.
- Singh N, Rao AS, Nandal A, Kumar S, Yadav SS, Ganaie SA, *et al.* Phytochemical and pharmacological review of *Cinnamomum verum* J. Presl—a versatile spice used in food and nutrition. Food Chem. 2021;338:127773. doi: 10.1016/j.foodchem.2020.1 27773, PMID 32829297.
- Sahib AS. Anti-diabetic and antioxidant effect of cinnamon in poorly controlled type-2 diabetic Iraqi patients: A randomized, placebo-controlled clinical trial. J Intercult Ethnopharmacol. 2016;5(2):108-13. doi: 10.5455/jice.20160217044511, PMID 27104030.
- Neyshaburinezhad N, Rouini MR, Entezari H, Lavasani H, Hosseinzadeh Ardakani YH. Evaluation of changes in cytochrome P450 2C19 activity in type 2 diabetic rats before and after treatment, by using isolated perfused liver model. Iran J Basic Med Sci. 2020;23(5):629-35. doi: 10.22038/ijbms.2020.40836.9642, PMID 32742601.
- Taheri A, Lavasani H, Kasirzadeh S, Sheikholeslami B, Ardakani YH, Rouini MR. Changes in CYP2D enzyme activity following induction of type 2 diabetesand administration of cinnamon and metformin: an experimental animal study. Xenobiotica. 2018;48(10):984-9. doi: 10.1080/00498254.2017.1390626, PMID 29092654.
- 61. Ashoor LA, Qusti SY. Potential interactions between cinnamon and metformin treatment in diabetic rats. Biosci Biotechnol Res Asia. 2016;7(2):607-16.
- Ashwini S, Bobby Z, Joseph M, Jacob SE, Padmapriya R. Insulin plant (*Costus pictus*) extract improves insulin sensitivity and ameliorates atherogenic dyslipidaemia in fructose-induced insulin-resistant rats: molecular mechanism. J Funct Foods. 2015;17:749-60. doi: 10.1016/j.jff.2015.06.024.
- Patibandla C, Khan ZI, MacGregor L, Campbell MJ, Patterson S. Costus pictus D. Don leaf extract stimulates GLP-1 secretion from GLUTag L-cells and has cytoprotective effects in BRIN-BD11 β-cells. J Ethnopharmacol. 2020;260:112970. doi: 10.1016/j.jep. 2020.112970, PMID 32422353.
- Naik A, Adeyemi SB, Vyas B, Krishnamurthy R. Effect of co-administration of metformin and extracts of *Costus pictus* D. Don leaves on alloxan-induced diabetes in rats. J Tradit Complement Med. 2022;12(3):269-80. doi: 10.1016/j.jtcme.2021.08. 007, PMID 35493313.
- Akram M, Shahab-Uddin AA, Usmanghani KH, Hannan A, Mohiuddin E, Asif M. *Curcuma longa* and curcumin: a review article. Rom J Biol Plant Biol. 2010;55(2):65-70.
- Undale VR, Kurkute SS, Jadhav SS. Curcumin potentiates therapeutic efficacy of metformin: a preclinical study in STZ-NA induced hyperglycemia in Wistar rats. Res J Pharm Technol. 2020;13(6):2591-6. doi: 10.5958/0974-360X.2020.00461.8.
- Bamba Y, Yun YS, Kunugi A, Inoue H. Compounds isolated from*Curcuma aromatica* Salisb. inhibit human P450 enzymes. J Nat Med. 2011;65(3-4):583-7. doi: 10.1007/ s11418-011-0507-0, PMID 21287405.
- Appiah-Opong R, Commandeur JN, van Vugt-Lussenburg B, Vermeulen NP. Inhibition of human recombinant cytochrome P450s by curcumin and curcumin decomposition products. Toxicology. 2007;235(1-2):83-91. doi: 10.1016/j.tox.2007.0 3.007, PMID 17433521.
- Graber-Maier A, Büter KB, Aeschlimann J, Bittel C, Kreuter M, Drewe J, et al. Effects of *Curcuma* extracts and curcuminoids on expression of P-glycoprotein and cytochrome P450 3A4 in the intestinal cell culture model LS180. Planta Med. 2010;76(16):1866-70. doi: 10.1055/s-0030-1249980, PMID 20509107.
- Volak LP, Ghirmai S, Cashman JR. Curcuminoids inhibit multiple human cytochromes P450, UDP-glucuronosyltransferase and sulfotransferase enzymes, whereas piperine is a relatively selective CYP3A4 inhibitor. Drug Metab Dispos. 2008;36(8):1594-605.
- Neerati P, Devde R, Gangi AK. Evaluation of the effect of curcumin capsules on glyburide therapy in patients with type-2 diabetes mellitus. Phytother Res. 2014;28(12):1796-800. doi: 10.1002/ptr.5201, PMID 25044423.
- Rani TS, Sujatha S, Veeresham C. Pharmacokinetic and pharmacodynamic interaction of curcumin with glimepiride in normal and diabetic rats. Pharmacogn Commun. 2012;2(3):14-21.
- Haritha C, Reddy AG, Reddy YR, Anilkumar B. Pharmacodynamic interaction of fenugreek, insulin and glimepiride on sero-biochemical parameters in diabetic Sprague-Dawley rats. Vet World. 2015;8(5):656-63. doi: 10.14202/vetworld.2015. 656-663, PMID 27047152.
- 74. Ahmmed SM, Mukherjee PK, Bahadur S, Kar A, Mukherjee K, Karmakar S, *et al.* Interaction potential of*Trigonella foenum graceum* through cytochrome P450 mediated inhibition. Indian J Pharmacol. 2015;47(5):530-4. doi: 10.4103/0253-7613 .165179, PMID 26600643.

- Al-Jenoobi FI, Al-Thukair AA, Alam MA, Abbas FA, Al-Mohizea AM, Alkharfy KM, et al. Effect of *Trigonella foenum-Graecum* L. Forsch Komplementmed. 2015;22(3):180-4. doi: 10.1159/000432412, PMID 26335391.
- Korashy HM, Al-Jenoobi FI, Raish M, Ahad A, Al-Mohizea AM, Alam MA, et al. Impact of herbal medicines likeNigella sativa, Trigonella foenum-graecum and Ferula asafoetida, on cytochrome P450 2C11 gene expression in rat liver. Drug Res. 2015;65(7):366-72. doi: 10.1055/s-0034-1384604, PMID 25099385.
- Abdelwahab NS, Morsi A, Ahmed YM, Hassan HM, AboulMagd AM. Ecological HPLC method for analyzing an antidiabetic drug in real rat plasma samples and studying the effects of concurrently administered fenugreek extract on its pharmacokinetics. RSC Adv. 2021;11(8):4740-50. doi: 10.1039/d0ra08836f. PMID 35424379.
- Satyanarayana S, Eswar Kumar KE, Rajasekhar J, Thomas L, Rajanna S, Rajanna B. Influence of aqueous extract of fenugreek-seed powder on the pharmacodynamics and pharmacokinetics of gliclazide in rats/rabbits. Therapy. 2007;4(4):457-63. doi: 1 0.2217/14750708.4.4.457.
- Lu FR, Shen L, Qin Y, Gao L, Li H, Dai Y. Clinical observation on *Trigonella foenum-Graecum* L. total saponins in combination with sulfonylureas in the treatment of type 2 diabetes mellitus. Chin J Integr Med. 2008;14(1):56-60. doi: 10.10 07/s11655-007-9005-3, PMID 18219452.
- Singh DK, Singh VK. Pharmacological effects of Allium sativum L. (garlic). Annu Rev Biomed Sci. 2008;10:6-26.
- Chourey S, Narsinghani T, Soni LK. Effect of *Allium sativum* on the pharmacokinetic of metformin in rat plasma: a herb-drug interaction study. Pharm Chem. 2011;3(2):287-92.
- Foster BC, Foster MS, Vandenhoek S, Krantis A, Budzinski JW, Arnason JT, et al. An in vitro evaluation of human cytochrome P450 3A4 and P-glycoprotein inhibition by garlic. J Pharm Pharm Sci. 2001;4(2):176-84. PMID 11466175.
- 83. Mahmoodi M, Zijoud SM, Hassanshahi GH, Toghroli MA, Khaksari M, Hajizadeh MR, et al. The effects of consumption of raw garlic on serum lipid level, blood sugar and a number of effective hormones on lipid and sugar metabolism in hyperglycemic and/or hyperlipidemic individuals-Benefit of raw garlic consumption. Adv Biol Chem. 2011;1(2):29-33. doi: 10.4236/abc.2011.12005.
- Hajda J, Rentsch KM, Gubler C, Steinert H, Stieger B, Fattinger K. Garlic extract induces intestinal P-glycoprotein, but exhibits no effect on intestinal and hepatic CYP3A4 in humans. Eur J Pharm Sci. 2010;41(5):729-35. doi: 10.1016/j.ejps.2010.09.016, PMID 20933082.
- Ho BE, Shen DD, McCune JS, Bui T, Risler L, Yang Z, et al. Effects of garlic on cytochromes P450 2C9-and 3A4-mediated drug metabolism in human hepatocytes. Sci Pharm. 2010;78(3):473-81. doi: 10.3797/scipharm.1002-11, PMID 20936048.
- Liu CT, Wong PL, Lii CK, Hse H, Sheen LY. Antidiabetic effect of garlic oil but not diallyl disulfide in rats with streptozotocin-induced diabetes. Food Chem Toxicol. 2006;44(8):1377-84. doi: 10.1016/j.fct.2005.07.013, PMID 16690190.
- Tripathi P, Gupta PP, Lal VK. Effect of co-administration of *Allium sativum* extract and metformin on blood glucose of streptozotocin-induced diabetic rats. J Intercult Ethnopharmacol. 2013;2(2):81-4. doi: 10.5455/jice.20130530123748.
- Ashraf R, Khan RA, Ashraf I. Garlic (Allium sativum) supplementation with standard antidiabetic agent provides better diabetic control in type 2 diabetes patients. Pak J Pharm Sci. 2011;24(4):565-70. PMID 21959822.
- Poonam T, Prakash GP, Kumar LV. Influence of Allium sativum extract on the hypoglycemic activity of glibenclamide: an approach to possible herb-drug interaction. Drug Metab Drug Interact. 2013;28(4):225-30. doi: 10.1515/dmdi-2013-0 031, PMID 24114899.
- Bharti SK, Krishnan S, Kumar A, Kumar A. Antidiabetic phytoconstituents and their mode of action on metabolic pathways. Ther Adv Endocrinol Metab. 2018;9(3):81-100. doi: 10.1177/2042018818755019, PMID 29492244.
- Srinivasan GV, Unnikrishnan KP, Rema Shree AB, Balachandran I. HPLC estimation of berberine in*Tinospora cordifolia* and*Tinospora sinensis*. Indian J Pharm Sci. 2008;70(1):96-9. doi: 10.4103/0250-474X.40341, PMID 20390090.
- Bahadur S, Mukherjee PK, Milan Ahmmed SK, Kar A, Harwansh RK, Pandit S. Metabolism-mediated interaction potential of standardized extract of *Tinospora* cordifolia through rat and human liver microsomes. Indian J Pharmacol. 2016;48(5):576-81. doi: 10.4103/0253-7613.190758, PMID 27721546.
- Vora A, Varghese A, Kachwala Y, Laddha AP, Bhaskar M, Akhtar J, et al. Pharmacokinetic and pharmacodynamic interactions of *Tinospora cordifolia* aqueous extract and hypoglycemic drugs in streptozotocin-induced diabetes in rats. Pharmacogn Mag. 2020;16(68):47. doi: 10.4103/pm.pm_272_19.
- Sahu R, Ahmed T, Sangana R, Punde R, Subudhi BB. Effect of *Tinospora cordifolia* aqua-alcoholic extract on pharmacokinetic of glibenclamide in rat: an herb-drug interaction study. J Pharm Biomed Anal. 2018;151:310-6. doi: 10.1016/j.jpba.2018.0 1.010, PMID 29413979.
- Thomas AB, Shaikh AS, Raje A, Lokhande KB, Swamy KV, Nagore DH, et al. Herb drug interaction of *Tinospora cordifolia* (Willd.) Miers extract and glimepiride: *in vivo* and *in silico* studies. Med Plants Int J Phytomed Relat Ind. 2020;12(4):555-67. doi: 10.5958/ 0975-6892.2020.00068.4.
- Li Y, Tran VH, Duke CC, Roufogalis BD. Preventive and protective properties of Zingiber officinale (ginger) in diabetes mellitus, diabetic complications and associated lipid and other metabolic disorders: a brief review. Evid Based Complement Alternat Med. 2012; 2012:516870. doi: 10.1155/2012/516870, PMID 23243452.

- Rattanapun N, Buppanhasamai N, Kangkan S, Sarapusit S. Inhibitory of the human cytochrome P450 2A6 and cytochrome P450 2A13 by ginger extracts and star fruits extracts. Burapha Sci J. 2017;22:163-72.
- Li M, Chen PZ, Yue QX, Li JQ, Chu RA, Zhang W, et al. Pungent ginger components modulate human cytochrome P450 enzymes in vitro. Acta Pharmacol Sin. 2013;34(9):1237-42. doi: 10.1038/aps.2013.49, PMID 23770984.
- Kim J. Effects of 6-shogaol, a major component of *Zingiber officinale* Roscoe, on human cytochrome P450 enzymes *in vitro*. Korean soc Med Crop Sci. 2016;24(1):7-13. doi: 10.7783/KJMCS.2016.24.1.7.
- Joo SY, Lim YC. Inhibitory effects of 6-gingerol on cytochrome P450 in human liver microsomes. J Korean Soc Clin Pharmacol Ther. 2011;19(1):52-8. doi: 10.12793/jkscp t.2011.19.1.52.
- Dhande S, Patil A, Kadam L. Study of effect of supplementation of *Zingiber officinale* on pharmacokinetic profile of sitagliptin phosphate on streptozotocin-induced type II DM rat model. Pharmacia *Tutor*. 2015;3(12):40-5.
- Al-Omaria IL, Afifib FU, Salhaba AS. Therapeutic effect and possible herb drug interactions of ginger (*Zingiber officinale* Roscoe, Zingiberaceae) crude extract with glibenclamide and insulin. *World*. 2012;9(11).
- Trumbeckaite S, Bernatoniene J, Majiene D, Jakštas V, Savickas A, Toleikis A. Effect of *Ginkgo biloba* extract on the rat heart mitochondrial function. J Ethnopharmacol. 2007;111(3):512-6. doi: 10.1016/j.jep.2006.12.028, PMID 17258877.
- von Moltke LL, Weemhoff JL, Bedir E, Khan IA, Harmatz JS, Goldman P, et al. Inhibition of human cytochromes P450 by components of *Ginkgo biloba*. J Pharm Pharmacol. 2004;56(8):1039-44. doi: 10.1211/0022357044021, PMID 15285849.
- 105. Kudolo GB. The effect of 3-month ingestion of *Ginkgo biloba* extract (EGb 761) on pancreatic β -cell function in response to glucose loading in individuals with non-insulin-dependent diabetes mellitus. J Clin Pharmacol. 2001;41(6):600-11. doi: 10.1177/00912700122010483, PMID 11402628.
- Gaudineau C, Beckerman R, Welbourn S, Auclair K. Inhibition of human P450 enzymes by multiple constituents of the *Ginkgo biloba* extract. Biochem Biophys Res Commun. 2004;318(4):1072-8. doi: 10.1016/j.bbrc.2004.04.139, PMID 15147983.
- 107. Ohnishi N, Kusuhara M, Yoshioka M, Kuroda K, Soga A, Nishikawa F, et al. Studies on interactions between functional foods or dietary supplements and medicines. I. Effects of *Ginkgo biloba* leaf extract on the pharmacokinetics of diltiazem in rats. Biol Pharm Bull. 2003;26(9):1315-20. doi: 10.1248/bpb.26.1315, PMID 12951478.
- Pinto MD, Kwon YI, Apostolidis E, Lajolo FM, Genovese MI, Shetty K. Potential of *Ginkgo bilobaL*. leaves in the management of hyperglycemia and hypertension using *in vitro* models. Bioresour Technol. 2009;100(24):6599-609. doi: 10.1016/j.biort ech.2009.07.021, PMID 19665890.
- Uchida S, Yamada H, Li XD, Maruyama S, Ohmori Y, Oki T, et al. Effects of *Ginkgo* biloba extract on pharmacokinetics and pharmacodynamics of tolbutamide and midazolam in healthy volunteers. J Clin Pharmacol. 2006;46(11):1290-8. doi: 10.1177 /0091270006292628, PMID 17050793.
- 110. Kudolo GB, Wang W, Javors M, Blodgett J. The effect of the ingestion of *Ginkgo biloba* extract (EGb 761) on the pharmacokinetics of metformin in non-diabetic and type 2 diabetic subjects—a double blind placebo-controlled, crossover study. Clin Nutr. 2006;25(4):606-16. doi: 10.1016/j.clnu.2005.12.012, PMID 16698134.
- Lee WK, Kao ST, Liu IM, Cheng JT. Increase of insulin secretion by ginsenoside Rh2 to lower plasma glucose in Wistar rats. Clin Exp Pharmacol Physiol. 2006;33(1-2):27-32. doi: 10.1111/j.1440-1681.2006.04319.x, PMID 16445695.
- Nam SJ, Han YJ, Lee W, Kang B, Choi MK, Han YH, et al. Effect of red ginseng extract on the pharmacokinetics and efficacy of metformin in streptozotocin-induced diabetic rats. Pharmaceutics. 2018;10(3):80. doi: 10.3390/pharmaceutics10030080, PMID 29970815.
- 113. Cho WC, Chung WS, Lee SK, Leung AW, Cheng CH, Yue KK. Ginsenoside Re ofpanax ginseng possesses significant antioxidant and antihyperlipidemic efficacies in streptozotocin-induced diabetic rats. Eur J Pharmacol. 2006;550(1-3):173-9. doi: 10 .1016/j.ejphar.2006.08.056, PMID 17027742.
- 114. Vuksan V, Sung MK, Sievenpiper JL, Stavro PM, Jenkins AL, Di Buono M, et al. Korean red ginseng (*Panax ginseng*) improves glucose and insulin regulation in well-controlled, type 2 diabetes: results of a randomized, double-blind, placebo-controlled study of efficacy and safety. Nutr Metab Cardiovasc Dis. 2008;18(1):46-56. doi: 10.1016/j.num ecd.2006.04.003, PMID 16860976.
- 115. Yoon SH, Han EJ, Sung JH, Chung SH. Anti-diabetic effects of compound K versus metformin versus compound K-metformin combination therapy in diabetic db/ db mice. Biol Pharm Bull. 2007;30(11):2196-200. doi: 10.1248/bpb.30.2196, PMID 17978500.
- 116. Jin S, Lee S, Jeon JH, Kim H, Choi MK, Song IS. Enhanced intestinal permeability and plasma concentration of metformin in rats by the repeated administration of red ginseng extract. Pharmaceutics. 2019;11(4):189. doi: 10.3390/pharmaceutics11040 189, PMID 31003498.
- 117. Zheng YF, Bae SH, Choi EJ, Park JB, Kim SO, Jang MJ, et al. Evaluation of the in vitro/in vivo drug interaction potential of BST204, a purified dry extract of ginseng and its four bioactive ginsenosides through cytochrome P450 inhibition/induction and UDP-glucuronosyltransferase inhibition. Food Chem Toxicol. 2014;68:117-27. doi: 1 0.1016/j.fct.2014.03.004, PMID 24632066.
- Fenwick GR, Lutomski J, Nieman C. Liquorice, *Glycyrrhiza glabra* L.—composition, uses and analysis. Food Chem. 1990;38(2):119-43. doi: 10.1016/0308-8146(90) 90159-2.

- Al-Snafi AE. *Glycyrrhiza glabra*: a phytochemical and pharmacological review. IOSR J Pharm. 2018;8(6):1-17.
- Pandit S, Ponnusankar S, Bandyopadhyay A, Ota S, Mukherjee PK. Exploring the possible metabolism-mediated interaction of *Glycyrrhiza glabra* extract with CYP3A4 and CYP2D6. Phytother Res. 2011;25(10):1429-34. doi: 10.1002/ptr.3426, PMID 21351298.
- 121. Paolini M, Pozzetti L, Sapone A, Cantelli-Forti G. Effect of licorice and glycyrrhizin on murine liver CYP-dependent monooxygenases. Life Sci. 1998;62(6):571-82. doi: 10.10 16/s0024-3205(97)01154-5, PMID 9464470.
- Zhao K, Ding M, Cao H, Cao ZX. *In vitro* metabolism of glycyrrhetinic acid by human and rat liver microsomes and its interactions with six CYP substrates. J Pharm Pharmacol. 2012;64(10):1445-51. doi: 10.1111/j.2042-7158.2012.01516.x, PMID 22943175.
- 123. Ao Y, Chen J, Yue J, Peng RX. Effects of 18α-glycyrrhizin on the pharmacodynamics and pharmacokinetics of glibenclamide in alloxan-induced diabetic rats. Eur J Pharmacol. 2008;587(1-3):330-5. doi: 10.1016/j.ejphar.2008.03.043, PMID 18462715.
- Al-Naqeb G, Fiori L, Ciolli M, Aprea E. Prickly pear seed oil extraction, chemical characterization and potential health benefits. Molecules. 2021;26(16):5018. doi: 10. 3390/molecules26165018, PMID 34443606.
- Díaz MD, de la Rosa AP, Héliès-Toussaint C, Guéraud F, Nègre-Salvayre A. Opuntia spp.: characterization and benefits in chronic diseases. Oxid Med Cell Longev. 2017.
- 126. Hwang SH, Kang IJ, Lim SS. Antidiabetic effect of fresh nopal (*Opuntia ficus-indica*) in low-dose streptozotocin-induced diabetic rats fed a high-fat diet. Evid Based Complement Alternat Med. 2017; 2017:4380721. doi: 10.1155/2017/4380721, PMID 28303158.
- 127. Leem KH, Kim MG, Hahm YT, Kim HK. Hypoglycemic effect of *Opuntia ficus-indica* var.*saboten* is due to enhanced peripheral glucose uptake through activation of AMPK/p38 MAPK pathway. Nutrients. 2016;8(12):800. doi: 10.3390/nu8120800, PMID 27941667.
- 128. Anwar MM, Sallam EM. Utilization of prickly pear peels to improve quality of pan bread. Arab J Nucl Sci Appl. 2016;49(2):151-63.
- Nuñez-López MA, Paredes-López O, Reynoso-Camacho R. Functional and hypoglycemic properties of nopal cladodes (*O. ficus-indica*) at different maturity stages using *in vitro* and *in vivo* tests. J Agric Food Chem. 2013;61(46):10981-6. doi: 1 0.1021/jf403834x, PMID 24164385.
- Hassan F, El-Razek A, Hassan AA. Nutritional value and hypoglycemic effect of prickly cactus pear (*Opuntia ficus-indica*) fruit juice in alloxan-induced diabetic rats. Aust J Basic Appl Sci. 2011;5(10):356-77.
- El-Mostafa K, El Kharrassi Y, Badreddine A, Andreoletti P, Vamecq J, El Kebbaj MS et al. Nopal cactus (Opuntia ficus-indica) as a source of bioactive compounds for nutrition, health and disease. Molecules. 2014;19(9):14879-901. doi: 10.3390/molecules19091 4879, PMID 25232708.
- Deldicque L, Van Proeyen K, Ramaekers M, Pischel I, Sievers H, Hespel P. Additive insulinogenic action of *Opuntia ficus-indica* cladode and fruit skin extract and leucine after exercise in healthy males. J Int Soc Sports Nutr. 2013;10(1):45. doi: 10.1186/ 1550-2783-10-45, PMID 24144232.
- 133. Sobieraj DM, Freyer CW. Probable hypoglycemic adverse drug reaction associated with prickly pear cactus, Glipizide and Metformin in a patient with type 2 diabetes mellitus. Ann Pharmacother. 2010;44(7-8):1334-7. doi: 10.1345/aph.1P148, PMID 20516361.
- Liu HM, Kiuchi F, Tsuda Y. Isolation and structure elucidation of gymnemic acids, antisweet principles of *Gymnema sylvestre*. Chem Pharm Bull (Tokyo). 1992;40(6):1366-75. doi: 10.1248/cpb.40.1366, PMID 1327559.
- Al-Romaiyan A, King AJ, Persaud SJ, Jones PM. A novel extract of *Gymnema sylvestre* improves glucose tolerance *in vivo* and stimulates insulin secretion and synthesis *in vitro*. Phytother Res. 2013;27(7):1006-11. doi: 10.1002/ptr.4815, PMID 22911568.
- Tiwari P, Mishra BN, Sangwan NS. Phytochemical and pharmacological properties of *Gymnema sylvestre*: an important medicinal plant. BioMed Res Int. 2014; 2014:830285. doi: 10.1155/2014/830285, PMID 24511547.
- 137. Rammohan B, Samit K, Chinmoy D, Arup S, Amit K, Ratul S, et al. Human cytochrome P450 enzyme modulation byGymnema sylvestre: a predictive safety evaluation by LC-MS/MS. Pharmacogn Mag. 2016;12 Suppl 4:S389-94. doi: 10.4103/0973-1296.19 1441, PMID 27761064.
- Dhande SR, Lokegaonkar DV, Bhutkar SP. Effect of *Gymnema sylvestre* on the pharmacokinetics of sitagliptin phosphate in type II diabetes mellitus. Int J Pharm Sci Res. 2017;8(3):1160.
- Dholi SK, Raparla R. Effect of *Gymnema sylvestre* on the pharmacokinetic and pharmacodynamic of oral hypoglycemic drug-gliclazide in streptozotocin-induced diabetic rats. Annu Res Rev Biol. 2014;4(22):3373-85. doi: 10.9734/ARRB/2014/8166.
- 140. Kamble B, Gupta A, Moothedath I, Khatal L, Janrao S, Jadhav A, et al. Effects of *Gymnema sylvestre* extract on the pharmacokinetics and pharmacodynamics of glimepiride in streptozotocin-induced diabetic rats. Chem Biol Interact. 2016;245:30-8. doi: 10.1016/j.cbi.2015.12.008, PMID 26721197.
- 141. Dholi SK, Raparla R. Effect of *Gymnema sylvestre* on the pharmacokinetics and pharmacodynamics of 0.5 mg and 0.6 mg glibenclamide in diabetic rats. Int J Pharmacol Res. 2015;5:172-8.
- Baliga MS, Bhat HP, Baliga BR, Wilson R, Palatty PL. Phytochemistry, traditional uses and pharmacology of *Eugenia jambolana* Lam. Food Res Int. 2011;44(7):1776-89. doi: 10.1016/j.foodres.2011.02.007.

- Prince PS, Menon VP, Pari L. Effect of *Syzigium cumini* extracts on hepatic hexokinase and glucose-6-phosphatase in experimental diabetes. Phytother Res. 1997;11(7):529-31. doi: 10.1002/(SICI)1099-1573(199711)11: 7<529::AID-PTR140> 3.0.CO;2-M.
- 144. Chinni S, Dubala A, Kosaraju J, Khatwal RB, Satish Kumar MN, Kannan E. Effect of crude extract of *Eugenia jambolana* Lam. on human cytochrome P450 enzymes. Phytother Res. 2014;28(11):1731-4. doi: 10.1002/ptr.5137, PMID 24590863.
- 145. Mastan SK, Latha TB, Latha TS, Srikanth A, Chaitanya G, Kumar KE. Influence of methanolic extract ofSyzygium cumini seeds on the activity of gliclazide in normal and alloxan-induced diabetic rats. Pharmacol Online. 2009;3:845-50.
- 146. Vora A, Varghese A, Kachwala Y, Bhaskar M, Laddha A, Jamal A, et al. Eugenia jambolana extract reduces the systemic exposure of sitagliptin and improves conditions associated with diabetes: a pharmacokinetic and a pharmacodynamic herb-drug interaction study. J Tradit Complement Med. 2019;9(4):364-71. doi: 10.101 6/j.jtcme.2018.10.001, PMID 31453133.
- Chaudhari S, Zambad S, Ali M. Pharmacokinetic and pharmacodynamics interaction between Syzygium cumini and glipizide: role of cytochrome P450 enzyme. Indian J Pharm Educ Res. 2019;53(3):S273-9.
- 148. Han K, Bose S, Wang JH, Lim SK, Chin YW, Kim YM, et al. *In vivo* therapeutic effect of combination treatment with metformin and *Scutellaria baicalensis* on maintaining bile acid homeostasis. PLOS One. 2017;12(9):e0182467. doi: 10.1371/journal.pone.0 182467, PMID 28877164.
- 149. Shin NR, Gu N, Choi HS, Kim H. Combined effects of *Scutellaria baicalensis* with metformin on glucose tolerance of patients with type 2 diabetes via gut microbiota modulation. Am J Physiol Endocrinol Metab. 2020;318(1):E52-61. doi: 10.1152/ajpen do.00221.2019, PMID 31770016.
- 150. Kim JY, Lee SY, Kim DH, Kim BR, Park R, Lee BM. Effects of flavonoids isolated fromScutellariae radix on cytochrome P-450 activities in human liver microsomes. J Toxicol Environ Health A. 2002;65(5-6):373-81. doi: 10.1080/15287390252808046, PMID 11936218.
- 151. Waisundara VY, Hsu A, Huang D, Tan BK. Scutellaria baicalensis enhances the anti-diabetic activity of metformin in streptozotocin-induced diabetic Wistar rats. Am J Chin Med. 2008;36(3):517-40. doi: 10.1142/S0192415X08005953, PMID 18543386.
- Mishra LC, Singh BB, Dagenais S. Scientific basis for the therapeutic use of *Withania* somnifera (ashwagandha): a review. Altern Med Rev. 2000;5(4):334-46. PMID 10956379.
- 153. B N, C V. Effect of ashwagandha on pharmacokinetic and pharmacodynamic parameters of glimepiride in streptozotocin-induced diabetic rats. Asian J Pharm Clin Res. 2018;11(4):207-10. doi: 10.22159/ajpcr.2018.v11i4.23960.
- 154. Savai J, Pandita N, Chintamaneni M. Investigation of CYP1A interaction potential of *Withania somnifera* in rat and human liver microsomes. Indian J Pharm Sci. 2014;76:138-47.
- Andallu B, Radhika B. Hypoglycemic, diuretic and hypocholesterolemic effect of winter cherry (*Withania somnifera*, Dunal) root. Indian J Exp Biol. 2000;38(6):607-9. PMID 11116534.
- 156. Ahmmed SM, Mukherjee PK, Bahadur S, Harwansh RK, Kar A, Bandyopadhyay A, et al. CYP450 mediated inhibition potential of *Swertia chirata*: an herb from Indian traditional medicine. J Ethnopharmacol. 2016;178:34-9. doi: 10.1016/j.jep.2015.11.0 46, PMID 26657265.
- Rajesh CS, Holla R, Patil V, Anand AS, Prasad KH. Anti-hyperglycemic effect of Swertia chirata root extract on indinavir-treated rats. Natl J Physiol Pharm Pharmacol. 2017;7(6):569.
- Sekar BC, Mukherjee B, Chakravarti RB, Mukherjee SK. Effect of different fractions of *Swertia chirayita* on the blood sugar level of albino rats. J Ethnopharmacol. 1987;21(2):175-81. doi: 10.1016/0378-8741(87)90127-9, PMID 3437768.
- 159. Bhowmik D, Chiranjib YJ, Tripathi KK, Kumar KS. Herbal remedies of *Azadirachta indica* and its medicinal application. J Chem Pharm Res. 2010;2(1):62-72.
- 160. Chaudhari S, Zambad S, Ali M. Effect of aqueous extract of *Azadirachta indica* leaves on pharmacokineics and pharmacodynamics of glipizide. Drug Metab Lett. 2019;13(1):19-24. doi: 10.2174/1872312812666181106115247, PMID 30398126.
- 161. Kazeem MI, Dansu TV, Adeola SA. Inhibitory effect of *Azadirachta indica* A. Juss leaf extract on the activities of α-amylase and α-glucosidase. Pak J Biol Sci. 2013;16(21):1358-62. doi: 10.3923/pjbs.2013.1358.1362, PMID 24511747.
- 162. Bhat M, Kothiwale SK, Tirmale AR, Bhargava SY, Joshi BN. Antidiabetic properties of *Azardiracta indica* and *Bougainvillea spectabilis: in vivo* studies in murine diabetes model. Evid Based Complement Alternat Med. 2011; 2011:561625. doi: 10.1093/eca m/nep033, PMID 19389871.
- 163. Satyanarayana S, Eswar Kumar K, Cooty T, Rajanna S, Rajanna B. Influence of an aqueous extract of *Azadirachta indica* leaf on the pharmacodynamics and pharmacokinetics of gliclazide in rats and rabbits. J Herbs Spices Med Plants. 2009;15(1):16-23. doi: 10.1080/10496470902787451.
- 164. Singh J, Cumming E, Manoharan G, Kalasz H, Adeghate E. Medicinal chemistry of the anti-diabetic effects of *Momordica charantia*: active constituents and modes of actions. Open Med Chem J. 2011;5 Suppl 2: 70-7. doi: 10.2174/1874104501105010 070, PMID 21966327.
- Welihinda J, Karunanayake EH. Extra-pancreatic effects of Momordica charantia in rats. J Ethnopharmacol. 1986;17(3):247-55. doi: 10.1016/0378-8741(86)90112-1, PMID 3807387.

- 166. Chao CY, Huang CJ. Bitter gourd (*Momordica charantia*) extract activates peroxisome proliferator-activated receptors and upregulates the expression of the acyl CoA oxidase gene in H4IIEC3 hepatoma cells. J Biomed Sci. 2003;10(6 Pt 2):782-91. doi: 10 .1159/000073966, PMID 14631118.
- 167. Sarkar S, Pranava M, MARITA R. Demonstration of the hypoglycemic action of *Momordica charantia* in a validated animal model of diabetes. Pharmacol Res. 1996;33(1):1-4. doi: 10.1006/phrs.1996.0001, PMID 8817639.
- 168. Fasinu PS, Manda VK, Dale OR, Egiebor NO, Walker LA, Khan SI. Modulation of cytochrome P450, P-glycoprotein and pregnane X receptor by selected antimalarial herbs—implication for herb-drug interaction. Molecules. 2017;22(12):2049. doi: 10.3 390/molecules22122049, PMID 29168799.
- Nivitabishekam SN, Asad M, Prasad VS. Pharmacodynamic interaction of *Momordica charantia* with rosiglitazone in rats. Chem Biol Interact. 2009;177(3):247-53. doi: 10.1 016/j.cbi.2008.09.034, PMID 18983991.
- Tongia A, Tongia SK, Dave M. Phytochemical determination and extraction of *Momordica charantia* fruit and its hypoglycemic potentiation of oral hypoglycemic drugs in diabetes mellitus (NIDDM). Indian J Physiol Pharmacol. 2004;48(2):241-4. PMID 15521566.

- 171. Khan W, Parveen R, Chester K, Parveen S, Ahmad S. Hypoglycemic potential of aqueous extract of *Moringa oleifera* leaf and *in vivo* GC-MS metabolomics. Front Pharmacol. 2017;8:577. doi: 10.3389/fphar.2017.00577, PMID 28955221.
- 172. Anwar F, Latif S, Ashraf M, Gilani AH. Moringa oleifera: a food plant with multiple medicinal uses. Phytother Res. 2007;21(1):17-25. doi: 10.1002/ptr.2023, PMID 17089328.
- 173. Atawodi SE, Atawodi JC, Idakwo GA, Pfundstein B, Haubner R, Wurtele G, et al. Evaluation of the polyphenol content and antioxidant properties of methanol extracts of the leaves, stemand root barks of*Moringa oleifera* Lam. J Med Food. 2010;13(3):710-6. doi: 10.1089/jmf.2009.0057, PMID 20521992.
- 174. Taesotikul T, Navinpipatana V, Tassaneeyakul W. Selective inhibition of human cytochrome P450 1A2 by*Moringa oleifera*. Thai J Pharmacol. 2010;32(1):256-8.
- 175. Idakwoji PA, Salawu OA, Maiha BB, Obidike I, Tijani AY. Co-administeration of ethanolic leaf extract of *Moringa oleifera* and metformin improves glucose, lipid and protein profiles of diabetic Wistar rats. Biokemistri. 2015;27(3):129-38.
- 176. Ahmmed SK, Mukherjee PK, Bahadur S, Kar A, Al-Dhabi NA, Duraipandiyan V. Inhibition potential of *Moringa oleifera* Lam. on drug metabolizing enzymes. 2015.
- 177. Bharathi K, Sandhya M, Prasad KV. Effect of *Moringa oleifera* on the pharmacokinetics and pharmacodynamics of pioglitazone. FASEB J. 2017;31(S1):822-5. doi: 10.1096/fa sebj.31.1_supplement.822.5.

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